

Fully Automated Synthesis of DNA-Binding Py-Im Polyamides Using a Triphosgene Coupling Strategy

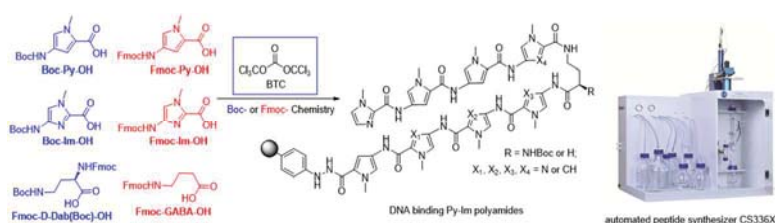
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Supporting Information Placeholder



ABSTRACT: The fully automated solid phase synthetic strategy of hairpin pyrrole-imidazole polyamides is described. A key advance described herein is the development of methodology for the application of triphosgene as a coupling agent in the automated synthesis of hairpin polyamides without racemization. This automated methodology is compatible with all the typical building blocks, enabling the facile synthesis of polyamide libraries in good yield (9-15%) and crude purity.

N-methylpyrrole-N-methylimidazole (Py-Im) polyamides are cell-permeable small molecules capable of recognizing pre-determined sequences of double-stranded DNA (dsDNA) by binding in the minor groove (Figure 1).¹ Sequence selectivity is achieved by precisely oriented pairs of aromatic Py and Im amino acids, such that an anti-parallel Py/Py preferentially targets A•T or T•A base pairs; Im/Py targets a C•G base pair while a Py/Im pair targets a C•G base pair.² As a consequence of their predictable binding and nanomolar binding affinity for target dsDNA sequences, Py-Im polyamides have considerable potential as exogenous agents that can regulate transcription by disrupting transcription factor-dsDNA binding interactions *in vivo*.³ Additionally, the ability of Py-Im polyamides to discriminate between match and mismatched dsDNA sequences renders these molecules as excellent molecular probes for diagnostic applications⁴ and in the design of DNA-based nanophotonic materials.⁵

Over the past two decades, much attention has been devoted to the total synthesis of Py-Im polyamides.⁶ The synthesis of these molecules are challenging as a consequence of the low inherent nucleophilicity of the immobilized aromatic amine. The uses of conventional coupling reagents such as HATU or HOAt/DCC produce the corresponding coupled product in poor yield. This problem is exacerbated when Im building blocks are used.^{6c,g} To circumvent this issue, a number of strategies have been employed such as the preparation of

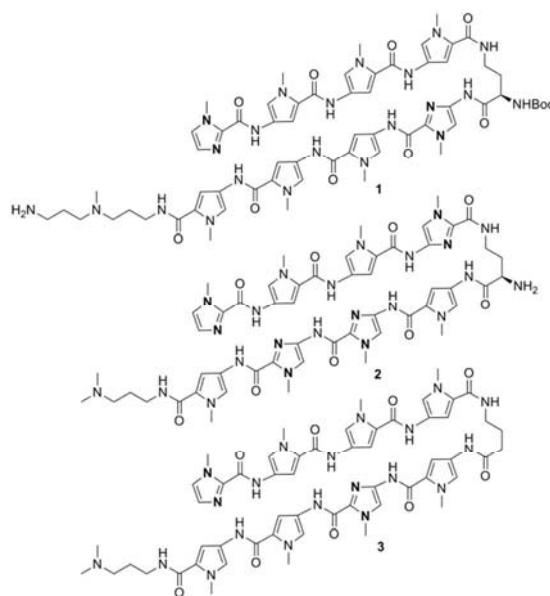


Figure 1. Structure of Py-Im polyamides

dimeric, trimeric and tetrameric carboxylic acids in solution phase^{6a,d,j} or using solution-phase total synthesis of the Py-Im polyamide core^{6c,f}. Recently, microwave-assisted synthesis

was utilized to facilitate the manual synthesis of Py-Im polyamides.^{6h,i} Although this method could significantly enhance the coupling yields and reduce coupling times, the difficulty in conducting immobilized imidazole-amine couplings was noted even at elevated temperatures (60–80°C). Machine-assisted syntheses have been employed by the Dervan and Sugiyama laboratories using Boc- and Fmoc- chemistry.^{6a,b,j} In these cases, the use of Boc-Py-Im-OH/Fmoc-Py-Im-OH dimers was a necessary requirement to overcome the difficulties aforementioned.^{6a,b,i,j} We previously reported a facile and highly efficient solid phase synthesis of hairpin DNA-binding polyamides using the cost effective BTC [bis(trichloromethyl) carbonate or triphosgene] activating agent.^{6g} This strategy was employed with great effect to produce Py-Im polyamides in 8–33% yield by manual solid phase synthesis. At present, a flexible, modular and facile protocol for the fully automated synthesis of Py-Im polyamides using monomeric building blocks much akin to solid phase peptide synthesis is not currently available. In this communication, we report such a strategy to produce a palette of hairpin Py-Im polyamides using BTC as the condensing agent for all coupling steps.

BTC putatively forms highly electrophilic acid chlorides in the presence of carboxylic acids *in situ*, which undergo facile amidation of sterically-hindered and electronically less-reactive amines.⁷ One limitation with current BTC-mediated coupling protocols is the strong exothermic nature of reaction, resulting in the emission of CO₂ accompanied by large amount of collidine hydrochloride precipitate (Figure 2). In order to render BTC-couplings amenable to automation, methods were therefore required to mitigate the exotherm and to prevent clogging of the lines of the automated synthesizer by the precipitate. Pre-activation of the amino acids with BTC outside the peptide synthesizer was employed by König *et al* in the machine-assisted synthesis of aromatic oligoamides as the solution could be freed of insoluble particles by syringe filtration.⁸ However, the pre-activated amino acid chloride has limited stability, which is not feasible to be used in the automated synthesis of Py-Im polyamides. Therefore, development of a milder and solid free activation method is crucial for the automation of the BTC strategy.

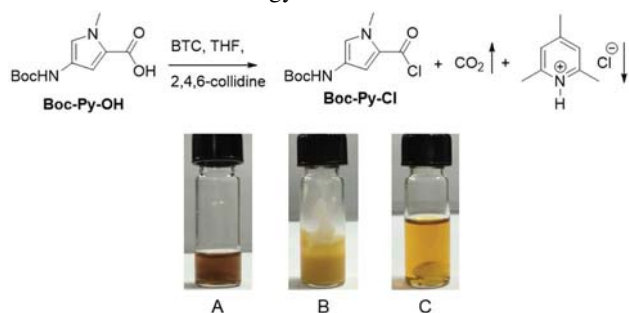


Figure 2. Activation of Boc-Py-OH with BTC. (A. the solution of Boc-Py-OH and BTC in THF; B. the slurry generated by addition of 2,4,6-collidine to A; C. the clear solution obtained by addition of a 10% DIEA/DMF solution to B)

In a primary study, the reaction conditions of BTC-mediated *in situ* activation were investigated in order to meet the requirements of commercially available peptide synthesizers. As observed by Fuse *et al* in the formation of acid chloride in a microflow system,⁹ the combinations of BTC, organic bases (Et₃N, *i*-Pr₂NEt, 2,4,6-collidine, pyridine) and solvents (THF,

1,4-dioxane, DMF) generated precipitations. The only combination of BTC, *i*-Pr₂NEt and CH₂Cl₂ generating no precipitation resulted in low coupling yields as well as epimerization of the chiral amino acids. Because Py-Im polyamides syntheses usually involved the coupling of a chiral amino acid as γ -turn unit, the retention of the chirality during the incorporation of this amino acid had to be concerned (Figure 3). Recently, we developed a reproducible, highly efficient and racemization free protocol for triphosgene-mediated solid-phase coupling of proteinogenic amino acids.¹⁰ The key features of the procedure include: (1) the preactivation of the carboxyl acid is carried out with collidine in THF, racemization-free conditions validated by the research of Jung and Falb;⁷ (2) the resulting precipitation is converted to the soluble DIEA hydrochloride salt by the addition of a DIEA/DMF solution. According to this procedure, Boc-Py-OH (1.0 equiv.) and BTC (0.33 equiv.) were dissolved in dry THF (Figure 2, A), collidine was added slowly to form a slurry (Figure 2, B).¹¹ Upon addition of a 10% DIEA/DMF mixture, the insoluble collidine hydrochloride disappeared very quickly and a clear solution was generated (Figure 2, C). To avoid the potential clogging of the peptide synthesizer pipelines, this special combination of bases for the activation of amino acids could be utilized in the automated synthesis of Py-Im polyamides.

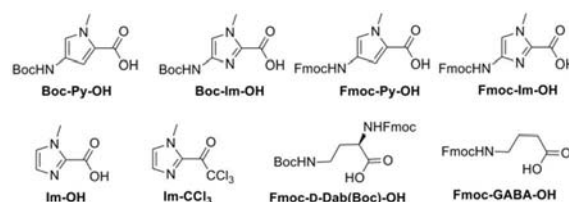


Figure 3. Building blocks for the synthesis of Py-Im polyamides

We then investigated whether the optimized BTC activation procedure could be conducted on a CS336X peptide synthesizer with a computer-controlled operation system. The synthesizer was programmed in the standard hardware configuration for DIC/HOBt (or HBTU/DIEA) protocols. To evaluate the automated activation efficiency of the BTC method, Boc-Py-OH (1.0 equiv) and BTC (0.33 equiv.) were loaded as solids into an amino acid reservoir, which were set up to be dissolved by dry THF (1.25 mL), activated by 2,4,6-collidine/dry THF (15%, 1.25 mL), followed by DIEA/dry DMF (10%, 2.5 mL). During each step, nitrogen was bubbled into the amino acid reservoir to blend the reaction mixture and facilitate the emission of CO₂. As a model study for the HPLC analysis, excess 2-(4-methoxyphenyl)-ethylamine was added to generate the corresponding amide. As shown in Figure 4, we found

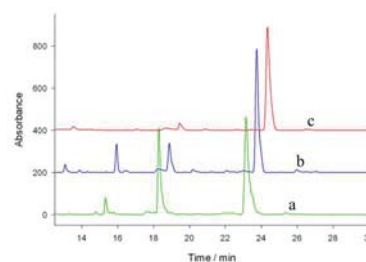
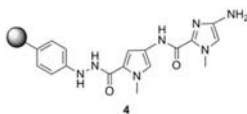


Figure 4. HPLC analysis of automated activation of Boc-Py-OH with BTC (285 nm): (a) 0.33 equiv. of BTC; (b) 0.41 equiv. of BTC; (c) 0.50 equiv. of BTC. Rt = 18.29 (Boc-Py-OH), Rt = 23.16 (the resulting amide).

that 0.33 equivalent of BTC, typically used in the activation step to release equal molar of phosgene, was not sufficient for full conversion of the acid chloride (trace a). We assume that this might be due to the emission of phosgene in the nitrogen bubbling step. The activation efficiency was significantly improved when 0.50 equiv. BTC was used (trace c) without any new by-product detected. Similar results were achieved when Boc-Im-OH or Fmoc-D-Dab(Boc)-OH was used, except for Im-OH which has poor solubility in THF. Therefore, 0.50 equiv. BTC is an optimal ratio for the efficient activation of both aromatic amino acids and aliphatic amino acids.

In regards to the poor nucleophilicity of the resin bound Im amines (Resin-Im-NH₂), we anticipated that the successful synthesis of Py-Im polyamides on a peptide synthesizer relied on the efficient coupling of Boc-Py-OH, Boc-Im-OH or Fmoc-D-Dab(Boc)-OH (γ -turn) to Im amines. An oligomer core (4) terminating with an Im nucleophile was used as a model study on the aryl hydrazine resin. We found that 40 min coupling time was optimal for complete coupling of Boc-Im-OH monomer to amine **4**. To ensure complete coupling of Boc-Py-OH to **4**, two successive rounds of coupling were conducted (25 min each). The coupling of Fmoc-D-Dab(Boc)-OH to the Im nucleophile was surprisingly slow and required an extended coupling step (200 min) to achieve acceptable yields. In optimizing the γ -Im coupling condition, we found that neither the reagent set DIC/HOAt/DIEA nor PyBOP/DIEA, which could be applied in the peptide synthesizer, were efficient in this amide bond formation. Further research revealed that the addition of one equivalent of HOAt to the BTC activated intermediate dramatically accelerated the rate of coupling with complete conversion observed by HPLC within 20 min. Using the combination of BTC/HOAt, the Im-Im coupling was also complete in 20 min. (Table 1)

Table 1. Optimized conditions for the coupling of monomers to Im amine **4**

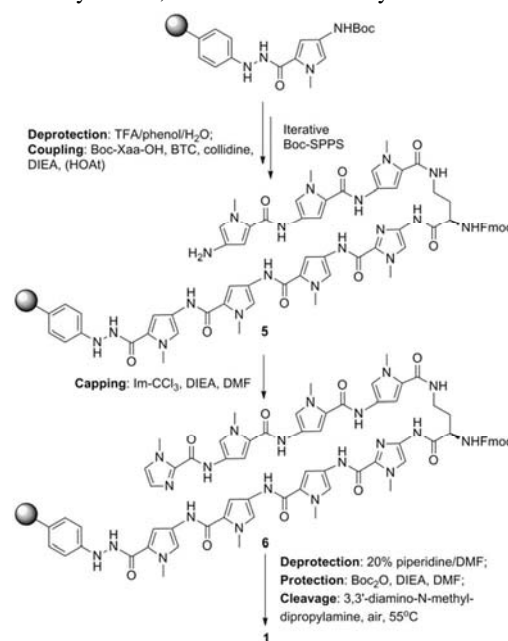


entry	monomer	monomer/BTC equiv/equiv	HOAt equiv ^a	coupling time (min) ^b
1	Boc-Py-OH	4/2	-	50 ^c
2	Boc-Im-OH	4/2	-	40
3	Fmoc-D-Dab(Boc)-OH	4/2	-	200
4	Fmoc-D-Dab(Boc)-OH	4/2	4	20
5	Boc-Im-OH	4/2	4	20

^aAfter Fmoc-D-Dab(Boc)-OH was activated using standard BTC procedure, a solution of HOAt/DMF/DIEA was added and the resulting mixture was transferred to the pretreated resin for the coupling reaction. ^bThe optimal coupling times were determined by the chloranil test for free Im amines of **4**. ^ctwo cycles, 25 min each.

Encouraged by this result, we then investigated the automated preparation of a biologically relevant polyamide **1**, targeting the 5'-WGWWC-3' (where W = A/T) subset of the consensus androgen and glucocorticoid response element.¹² The synthesis program on the CS336X peptide synthesizer was divided into three modules: (1) deprotection of the resin-

bound polyamide, including removal of the Boc group with TFA/Phenol/H₂O (92.5:5:2.5) cocktail, DCM washes, DIEA/DMF wash and DMF washes; (2) monomer coupling to the resin-bound polyamide, including BTC activation (5 min), delivery of the activated monomer, coupling (25 min) and DMF washes; (3) the terminal Im capping, including Im-CCl₃ coupling and DMF washes. For a typical synthesis on resin (400 mg, 0.15 mmol/g), Boc-amino acid (4 equiv.) and BTC (2 equiv.) were activated just prior to the coupling step and transferred to the pretreated resin to generate the corresponding polyamide. The automated solid phase synthesis of polyamide **1** involved eight coupling cycles plus a protecting group exchange step. All the program proceeded smoothly using the optimized BTC protocol, requiring no special precautions beyond those used for general solid phase peptide synthesis.¹³ Polyamide **1** was then cleaved from the resin using 3,3'-diamino-N-methyl-dipropylamine after oxidation of the hydrazide by N-bromosuccinimide or atmospheric O₂ in the presence of Cu(II) salts to form a transient diazene species.^{6g, 14} However, better yields were obtained by direct aminolysis of resin-bound polyamides under air at 55 °C for 16 h.^{6k} After semi-preparative HPLC purification, polyamide **1** was obtained in 15% yield with a purity of 95% (Scheme 1). Using the modular program described above, polyamide **2** comprising a diverse range of couplings encountered in polyamide synthesis was attempted. These included Py-Py, Im-Py, Im-Im, γ -Py and Im- γ couplings in addition to the known challenging Py-Im (Boc-Py-OH/Resin-Im-NH₂) coupling in the synthesis. After cleavage from the resin using dimethylamino-propylamine (Dp) and purification by semi-preparative HPLC, polyamide **2**, which was one of the most difficult sequences in polyamides synthesis, was obtained in 9% yield.



Scheme 1. Automated synthesis of Py-Im polyamide **1**

To further confirm whether the stereochemical information encoded in the turn unit would survive several additional coupling steps in the automated synthesis, we synthesized the two enantiomers of polyamide **6**th from Fmoc-L-Dab(Boc)-OH and Fmoc-D-Dab(Boc)-OH, respectively. Mosher amide derivatives **7**-(R,S) and **7**-(S,S) were formed by the reaction of both

enantiomers of **6** with (*S*)-Mosher's acid chloride. **7-(R,S)**, **7-(S,S)** and a mixture of them were subjected to HPLC analysis using a CHIRALPAK® ID column to separate the two diastereomers. As shown in Figure 5, no epimerization products could be detected for each diastereomer (trace b and c), indicating the chirality of the γ -turn unit was retained during the automated Py-Im polyamides synthesis.

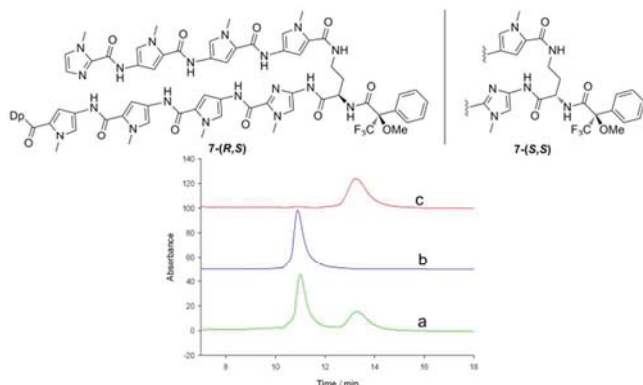


Figure 5. HPLC analysis of polyamide diastereomers (310 nm): (a) a mixture of **7-(R,S)** and **7-(S,S)**; (b) **7-(R,S)**, $R_t = 10.90$ min; (c) **7-(S,S)**, $R_t = 13.25$ min.

Finally, a further application of the optimized BTC protocol was explored *via* the preparation of polyamide **3** using an Fmoc-strategy. The deprotection step was conducted by twice deblocking (10 min) with 20% piperidine/DMF to remove Fmoc group from the resin-bound polyamide. Fmoc-Py-OH, Fmoc-Im-OH and Fmoc-GABA-OH were activated by the same BTC activation procedure. It should be noted that two coupling cycles were needed to drive the reaction of Fmoc-Py-OH/Resin-Im-NH₂ to completion (60 min each). All other couplings were carried out with a single-coupling cycle with extended reaction time (40 min). After capping with Im-CCl₃, cleavage from the resin and semi-preparative HPLC purification, polyamide **3** was obtained in 12% yield.

In summary, an automated protocol to solid phase Py-Im polyamides synthesis using BTC as the activating agent has been developed. This protocol was facile and fully amenable for automation on a CS336X peptide synthesizer using monomeric *N*-protected carboxylic acids regardless of the required sequence or *N*-protection strategy. The automation of the BTC strategy represented a significant step forward for the multiple, parallel synthesis of structurally diverse peptoids for biomedical studies and also for the large-scale production of this family of molecules for further exploration as therapeutic scaffolds.

ASSOCIATED CONTENT

Supporting Information

Experimental procedures and analysis data. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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Notes

The authors declare no competing financial interest.

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