

1 Integrated lignocellulosic biorefinery for efficient production of 2 furans and photothermal materials

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24 Abstract

25 Integrated lignocellulosic biorefineries offer a great potential to valorize all the
26 components in lignocellulose into products, including fuels, chemicals, and materials.

27 However, because of lignocellulose recalcitrance, conversion of bioresources remains

28 a techno-economic challenge for many lignocellulosic biorefineries. In this work, we

29 have proposed a sustainable and profitable biorefinery strategy for lignocellulose

30 fractionation and conversion. In this design, a biphasic solvent consisting of a molten

31 salt hydrate LiCl·4H₂O and γ -valerolactone (GVL) was initially used for separating

32 hemicellulose from lignocellulose. More interestingly, 100 wt% of biorefinery products

33 from lignin were directly converted to functional photothermal materials by
34 coordinating with Fe^{3+} for solar-thermal-electricity conversion. Attributed to this
35 rational design, the techno-economic analysis predicts a revenue of 439.3 USD by
36 processing 100 kg of lignocellulosic biomass using the above developed method.

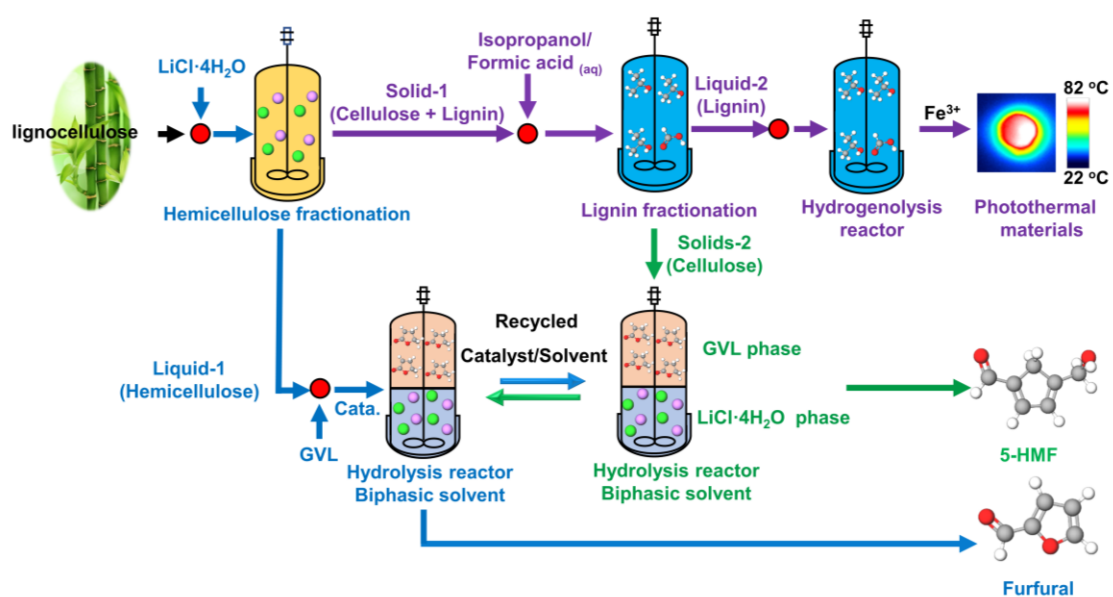
37 **Keywords:** lignocellulosic biomass, molten salt hydrate, solvent effects, photothermal
38 materials

39 **1. Introduction**

40 The core of the chemical industry is the conversion of petroleum resources into value-
41 added products, including fuels, platform chemicals, polymers, materials and
42 pharmaceuticals [1-3]. In the last century, fossil resources have constituted the primary
43 feedstock for a multitude of chemical processes and transformations [4-6]. However,
44 sustainability is becoming a global imperative due to the considerable depletion of
45 fossil resources and environmental concerns [7, 8]. As a result, the biorefinery paradigm
46 has been proposed as a solution for sustainable development [9, 10]. In a biorefinery,
47 sustainable biomass feedstocks are refined/upgraded to energy products, polymeric
48 materials, platform chemicals, and functional materials. Lignocellulose bioresources,
49 due to their abundance and renewability, are regarded as one of the most promising
50 sources for biorefineries [11-13]. Lignocellulose is primarily composed of cellulose
51 (35-50%), hemicellulose (25-30%), and lignin (15-30%) [14-16]. Amongst them,
52 cellulose and hemicellulose have been shown to exhibit potential as raw resources for
53 HMF, levulinic acid, furfural and other furan derivatives [17-19]. Lignin is frequently
54 proposed as a suitable source for guaiacol, phenol, phenolic resins, and other functional

55 materials [20-23]. The traditional lignocellulose biorefinery is heavily cellulose-
56 centered, and it faces significant challenges nowadays [24]. To obtain readily
57 hydrolysable cellulose, lignocellulose is often delignified initially prior to enzymatic
58 hydrolysis, employing high pretreatment severities. In this process, a major part of
59 hemicellulose could be lost especially under acidic conditions. Equally concerning, the
60 extracted lignin often contains relatively inert C-C interunit linkages, rendering a low-
61 value lignin resource for producing aromatic compounds [25, 26]. To address this
62 problem, the lignin-first biorefinery was proposed as described in two works by Abu-
63 Omar and Sels's group in 2015 [27, 28]. In this strategy, lignin is initially dissolved and
64 depolymerized to yield phenolic/aromatic monomers, dimers, and oligomers, while
65 cellulose and hemicellulose are retained in the solid residue for further treatment (e.g.,
66 enzymatic hydrolysis). This strategy represents significant progress in an integrated
67 lignocellulose biorefinery, and for a more elaborate discussion on the development and
68 guidelines of the lignin first approaches, the reader is referred to these dedicated studies
69 and reviews [29-31]. Nevertheless, several critical challenges remained in the proposed
70 process. Firstly, although the lignin-derived bio-oil has great potential, applications are
71 still limited by the complicated isolated components. Moreover, a large amount of
72 hemicellulose is lost or isomerized in lignin-first biorefinery, preventing their further
73 conversion to chemicals [32, 33]. These considerations, combined with low efficient
74 recovery of expensive catalysts, rendered a reduced efficiency and techno-economic
75 balance for a cost-effective integrated lignocellulose biorefinery.
76 To address these issues, we have proposed a sustainable and profitable biorefinery

77 strategy (**Figure. 1**). In this design, a recyclable biphasic solvent system consisting of
 78 a molten salt hydrate ($\text{LiCl}\cdot 4\text{H}_2\text{O}$) and biomass-derived γ -valerolactone (GVL) solvent
 79 was used for hemicellulose fractionation from lignocellulose with a separation
 80 efficiency of 93.2 wt%. After that, 90.5 wt % of cellulose and 88.7 wt% of lignin were
 81 separated from the solid residue using isopropanol/formic acid. As-obtained
 82 hemicellulose and cellulose were subsequently converted to furfural and HMF with a
 83 conversion yield of 76.2 mol% and 77.9 mol% in the biphasic solvent, respectively,
 84 catalyzed by $\text{Al}_2(\text{SO}_4)_3$. Finally, the lignin fraction was hydrogenized to phenolic
 85 monomers and oligomers. Instead of separating and purifying the phenolic mixtures,
 86 nearly 100 wt% of them were directly converted to functional photothermal materials
 87 by coordinating with Fe^{3+} for solar-thermal-electricity conversion. Attributed to the
 88 rational design, techno-economic analysis suggests a revenue of 439.3 USD by
 89 processing 100 kg of lignocellulose using the as-developed method.



90

91 **Figure. 1.** Comprehensive catalytic strategy for lignocellulose powder full-component
 92 conversion.

93

94 **2. Experimental section**

95 **2.1. Chemicals**

96 Bamboo was purchased from a farm (Sichuang, China), which was ground and sieved
97 to average particle size of 50 μm prior to use. The chemical composition (wt%) of the
98 pubescens was 40.5 % cellulose, 32.8 % hemicellulose, 23.2 % acid insoluble lignin
99 and 2.6 % acid soluble lignin. The composition (wt%) of the pubescens was determined
100 according the method of National Renewable Energy Laboratory (NREL, 2008a,
101 Preparation of samples for Compositional Analysis; 2008b, Determination of Structural
102 Carbohydrates and Lignin in Biomass.). LiCl, $\text{Al}_2(\text{SO}_4)_3$, formic acid and
103 microcrystalline cellulose were purchased from Sinopharm Chemical Reagent Co. γ -
104 valerolactone and isopropanol were got from Aladdin Reagent Co. Guaiacol, 4-
105 ethylguaiacol, 2-methoxy-4-methylphenol, syringic acid, 2,6-dimethoxy-4-
106 methylphenol, 2,6-dimethoxyphenol, 4-methylphenol, phenol, syringaldehyde, 2-
107 methoxy-4-propylphenol, 4-ethylphenol, eugenol and 4-hydroxy-3-
108 methoxyacetophenone were supplied by Sigma-Aldrich company. Commercial 5 wt%
109 Ru/C, Pt/C, Pd/C and Rh/C catalyst were purchased from Shanghai Macklin
110 Biochemical Co., Ltd. All chemicals were analytical grade and used as received without
111 further purification.

112 **2.2. Preparation of molten salt hydrate**

113 The purchased LiCl and deionized water were mixed according to the required
114 water/salt molar ratio, then added to a 500 ML 316L stainless steel autoclave with
115 Teflon lining, followed by stirring (800 rpm) at 60 $^{\circ}\text{C}$ for 30 min. The $\text{LiCl}\cdot 4\text{H}_2\text{O}$ was
116 obtained after cooling to room temperature.

117

118 **2.3. Hemicellulose fractionation and hemicellulose-to-furfural conversion**

119 Typically, 3.0 g bamboo and 20.0 g LiCl·4H₂O were added into a 50 mL 316L stainless
120 steel autoclave with Teflon lining, followed by stirring at 800 rpm. Then, the reactor
121 was pressurized by 2 Mpa of N₂ and heated at 130 °C for 60 min. The zero time was
122 defined as the mixed reaction solution reaches the target temperature. Upon completing
123 the reaction, the reactor was rapidly cooled in an ice bath, and the mixture was filtered
124 through a 0.45 μm membrane to collect the solid and liquid products. The main
125 components of the solid product are cellulose and lignin, which were defined as **Solid-**
126 **1** (1.9 g). The solid products were washed 3-5 times with deionized water, freeze-dried,
127 and stored in a constant temperature drying oven for further treatment. The liquid
128 products were defined as **Liquid-1** (21.1 g, containing hemicellulose-derived
129 oligomers). Additionally, the above-obtained **liquid-1**, 10.0 g GVL and 0.1 g
130 Al₂(SO₄)₃·18H₂O were added to a 50 mL 316L stainless steel autoclave with Teflon
131 lining. N₂ was charged into the reactor at an initial pressure of 2.0 MPa to remove air,
132 followed by programmed heating (heating rate =7.5 °C/min) up to the target
133 temperature (120–170 °C) and stirring with 800 rpm for a period of time (15–90 min).
134 The products were analyzed by GC-MS and HPLC.

135
$$\text{Furfural yield} = \frac{\text{mole of furfural}}{\text{mole of hemicellulose}} \times 100\%$$

136
$$\text{Energy consumption (kWh)} = p \text{ (kW)} \times t \text{ (h)}$$

137 **2.4. Lignin fraction and Lignin depolymerization**

138 The above-obtained **Solid-1**, 3.0 g formic acid, 20.0 g isopropanol and 20.0 g H₂O were
139 added to a 100 mL 316L stainless steel autoclave with Teflon lining, followed by stirring

140 speed was fixed at 800 rpm. Then, the reactor was pressurized by 2 MPa of N₂ and
141 heated at 160 °C for 60 min. After the reaction was completed, the reactor was rapidly
142 cooled in an ice bath, and the mixture was filtered through a 0.45 µm membrane to
143 collect the solid and liquid products. The main component of the solid product is
144 cellulose, which was defined as **Solid-2** (1.08 g, rich in cellulose). The solid products
145 were washed 3-5 times with deionized water, freeze-dried, and stored in a constant
146 temperature drying oven for further treatment. The liquid products were defined as
147 **Liquid-2** (43.8 g, contains lignin-derived oligomers). Furthermore, the above-obtained
148 **Liquid-2** and 0.2 g commercial catalyst (e.g., Ru/C) were put together in a 100 mL
149 316L stainless steel autoclave with Teflon lining, followed by stirring speed was fixed
150 at 800 rpm. Then, the reactor was pressurized by 2 Mpa of H₂ and heated at 210 °C for
151 4.0 h. After reaction, the reactor was rapidly cooled down in an ice bath, and the mixture
152 was filtered through a 0.45 µm membrane to collect the catalyst and liquid products.
153 The liquid product is qualitatively and quantitatively analyzed by GC-MS and GC.

154 ***2.5. Depolymerized lignin-to-photothermal materials conversion***

155 1.2 g of hydrogenolyzed lignin was dissolved in 15.0 mL of ethanol. A sodium
156 hydroxide solution (10.0 mL, pH=12) was the added. The solution was stirred for 10
157 min, then an aqueous solution containing 0.4 g of ferric chloride was added (5.0 mL).
158 The mixed solution was stirred for 10 min and freeze-dried to give a solid powder (~1.6
159 g).

160 ***2.6. Cellulose-to-HMF conversion***

161 0.54 g of the above-obtained **Solid-2** (Contains 0.5g of cellulose), 10.0 g LiCl·4H₂O,

162 20.0 g GVL, and 0.05g $\text{Al}_2(\text{SO}_4)_3$ were placed in a 100 mL 316L stainless steel
163 autoclave with Teflon lining, followed by stirring speed was fixed at 800 rpm. It should
164 be noted that the solvent and catalyst can be fresh or recycled. Then, the reactor was
165 pressurized by 2 MPa of N_2 and heated at 160-200 °C for 60-120 min. Zero time was
166 defined as the mixed reaction time for the solution to reach the target temperature. After
167 the reaction, the reactor was rapidly cooled in an ice bath, and the mixture was filtered
168 through a 0.45 μm membrane to collect the solid and liquid products. The liquid product
169 is qualitatively and quantitatively analyzed by GC-MS and GC.

170 ***2.7. Computational simulations***

171 Forcite and CASTEP module of the Materials Studio software was employed for the
172 quantum chemistry calculations. In the molecular dynamics calculations, liquid-phase
173 molecular systems were prepared by placing a reaction molecule, solvent molecules in
174 a nearly $50 \times 50 \times 50$ periodic box to simulate the effect of ratios between $\text{LiCl} \cdot 4\text{H}_2\text{O}$ and
175 GVL on the reaction, where the number of reactants is constant, and the solvent
176 molecules were changed to simulate the concentrations. The operating temperature is
177 set at 413 K and controlled by means of the Nose thermostat method to match the actual
178 experimental procedure. The prepared molecular systems were equilibrated for 1 ns by
179 NVT (constant molecule numbers, volume, and temperature) ensemble. The Van der
180 Waals interaction is calculated by atom-based method with a cutoff radius of 18.5 Å to
181 avoid the errors of relatively short-range non-bond interactions, while electrostatic
182 summation is calculated by Ewald method to avoid the errors of long-range non-bond
183 energy in periodic systems.

184 Activation free energy are calculated by the following equation:

$$185 \quad \Delta E = E[\text{Product}] - E[\text{Reaction}]$$

186 where E[Product] and E[Reaction] is the energy of single molecule after optimized in
187 corresponding solution; For example, the activation free energy from Cellulose to
188 Glucose is calculated by E[Glucose] - E[Cellulose].

189 **2.8. Product analysis**

190 FAL and 5-HMF in the GVL was analyzed by gas chromatography (GC, 2010Plus,
191 Shimadzu) for qualitative and quantitative tests. The gas chromatograph uses the SH-
192 RTX-5 column (30m×0.32mm×0.5um), and the flame ionization detector (FID) were
193 used to determine the volatile components with 0.43 mL min⁻¹ of gaseous helium as the
194 carrier gas. The temperature of the injector was set to 260 °C, and the temperature of
195 the detector was 280 °C. The column temperature programming was adopted as follows:
196 The initial temperature of the chromatographic column was set at 100 °C for 2 min,
197 then heated up to 280 °C at the rate of 8.0 °C min⁻¹, and held at 280 °C for 10 min.
198 Likewise, phenolic monomers are detected with the above-mentioned gas phase
199 equipment. The initial temperature of the chromatographic column was set at 50 °C for
200 2 min, and the oven temperature program was raised with a ramp of 20 °C min⁻¹ from
201 50 °C to 280 °C for 5 min. The injector and detector temperatures was kept at 275 °C
202 and 280 °C, respectively. Besides, the FAL, 5-HMF and sugars in water phase were
203 measured by HPLC (Agilent 1200 series) equipped with an amines column (Model
204 HPX-87H, 300 mm×7.8 mm, Bio-Rad), a variable-wavelength detector (Model VWD-
205 3×00(RS)), and a refractive index (RI) detector (Model RI-101, Shodex). The mobile

206 phase was 5 mM H₂SO₄ at a flow rate of 0.6 mL/min. The temperature of the detector
207 and column oven was maintained at 35 °C and 50 °C, respectively. Moreover, the
208 chemical composition of HDO is determined by Anion Exchange Chromatography
209 (HPACE, Dionex ICS-3000) equipped with a Carbopac PA-100 column (4×250 mm,
210 Dionex), the mobile phase is 150 mM sodium hydroxide solution containing 100 mM
211 sodium acetate, the flow rate is 0.4 mL/min, and the column temperature is 30 °C.

212 ***2.9. Characterization of Solid-2 and microcrystalline cellulose***

213 The crystallinity of the sample was analyzed with an X-ray diffractometer (XRD) at 30
214 mA and 40 kV, respectively, the scanning rate was 5 °/min, and the scanning range was
215 5° to 50°. The crystallinity index (CrI) is calculated as follows:

$$216 \quad \text{CrI} = \frac{I_{002} - I_a}{I_{002}} \times 100\%$$

217 I₀₀₂ represents the diffraction intensity of the crystalline region (2θ = 22.5°) of cellulose
218 in the biomass component; I_a represents the diffraction intensity of the amorphous
219 region (2θ = 18.5°) of cellulose in the biomass component.

220 The composition and structure of the substrates and residues were determined by a
221 PerkinElmer Spectrum One Fourier transform infrared spectroscope (FTIR). The
222 attenuated total reflectance (ATR) method was adopted here over the scan area range
223 from 4000 to 500 cm⁻¹ with resolution of 4 cm⁻¹ and accumulation of 4 times.

224 ***2.10. Characterization of Liquid-1 and hydrogenolysis products***

225 Molecular weights of the Liquid-1 and hydrogenolysis products were measured by gel
226 permeation chromatography (GPC). The instrument used was a Waters high-
227 performance liquid chromatography with a refractive index (RI) detector. The mobile

228 phase is THF, and the constant column temperature is 30 °C. A calibration curve was
229 acquired by polystyrene as standards.

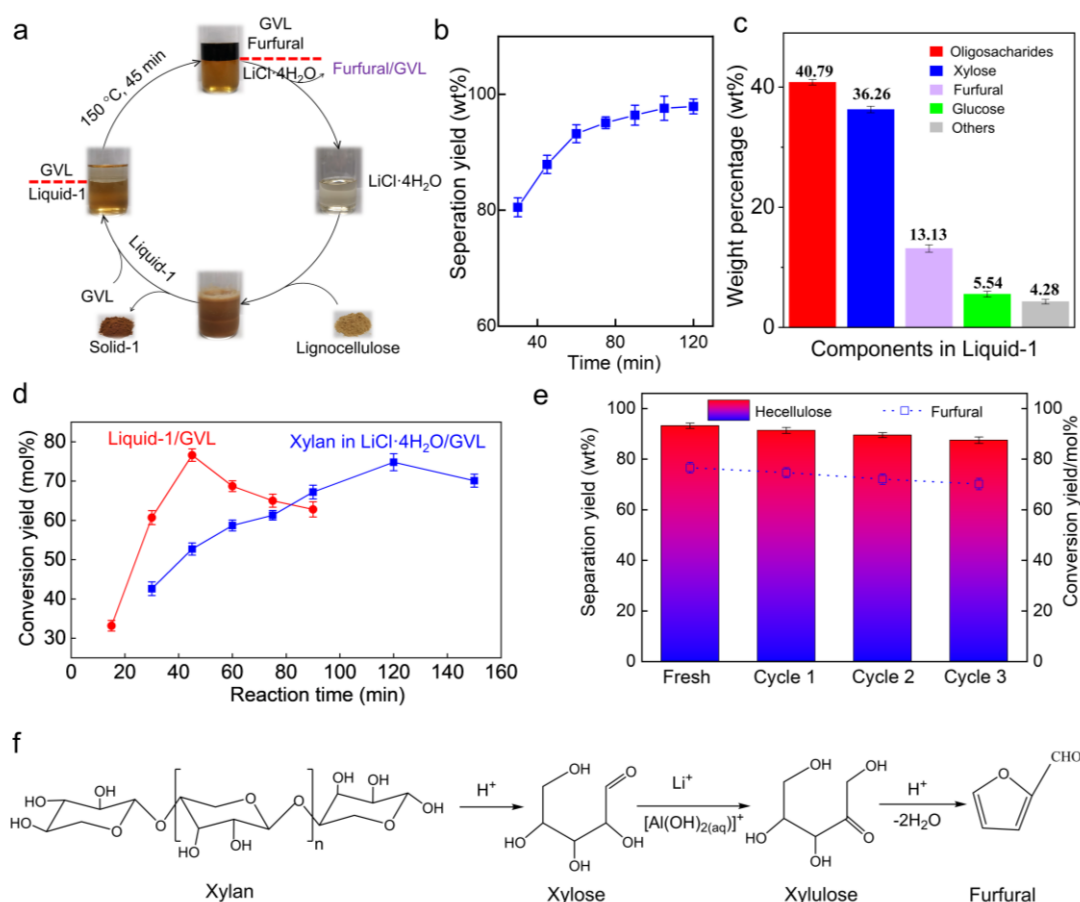
230 **3. Results and Discussion**

231 **3.1. Step 1: Hemicellulose-to-furfural conversion**

232 The hemicellulose fractionation from the native plant cell wall and its subsequent
233 conversion to furfural are illustrated in **Figure. 2a**. Specifically, 93.2 wt% of
234 hemicelluloses were separated from biomass at optimized conditions (130 °C for 60
235 min) in molten LiCl·4H₂O (**Figure. 2b**). Meanwhile, the molten salt hydrate LiCl·4H₂O
236 showed good selectivity as a hemicellulose solvent, while 90.1 wt% of cellulose and
237 96.3 wt% of lignin were not dissolved and remained in Solid-1 for further separation.
238 This result may be attributed to the following two points: 1) Metal cations can promote
239 the break of glycosidic bonds and melt salt hydrate has a good dissolution effect on
240 hemicellulose and cellulose[34, 35]; 2). Cellulose is encapsulated in hemicellulose and
241 lignin and has a dense crystalline structure, so it has higher stability than hemicellulose
242 and requires more demanding reaction conditions for dissolution/degradation[36, 37].
243 High-performance anion exchange chromatography (HPACE) and liquid
244 chromatography (HPLC) were used to determine the components in **Liquid-1** [38] .
245 The results showed that the main components of **Liquid-1** are oligosaccharides (40.79
246 wt%, derived from hemicellulose), xylose (36.26 wt%), furfural (13.13 wt%), glucose
247 (5.54 wt%), and others (4.28 wt%, *e.g.*, acetic acid, arabinose and galactose) (**Figure.**
248 **2c**). Subsequently, GVL aqueous solution was added to the **Liquid-1** to form a biphasic
249 solvent for the subsequent catalytic reactions. Al₂(SO₄)₃ was then added to the biphasic

250 solvent as a catalyst. A 76.2 mol% of furfural conversion in the biphasic solvent was
251 achieved under optimized conditions (150 °C for 45 min), which is comparable to the
252 maximum xylan-to-furfural conversion yield, but significantly less process time
253 (**Figure. 2d**). As a control, only 37.6 mol% (150 °C for 60 min) of furfural conversion
254 was achieved in the presence of $\text{Al}_2(\text{SO}_4)_3$ without the addition of GVL (**Figure. S1**).
255 The result was attributed to the fact that GVL could provide a protective shell for
256 furfural to avoid its condensation, thus having a positive effect on furfural
257 production[39]. A plausible conversion mechanism was then proposed. $\text{Al}_2(\text{SO}_4)_3$ is
258 simultaneously hydrolyzed to H^+ and Lewis acid catalytic active center $[\text{Al}(\text{OH})_2(\text{aq})]^+$
259 in $\text{LiCl} \cdot 4\text{H}_2\text{O}/\text{GVL}$. Oligosaccharides in **Liquid-1/GVL** were then depolymerized to
260 xylose under the acidic condition. Our previous studies showed that $[\text{Al}(\text{OH})_2(\text{aq})]^+$
261 could complex with the oxygen atoms of xylose at C1 and C2 position [40]. Such
262 interactions accelerated hydrogen transfer between the two carbon atoms, followed by
263 isomerization of xylose into xylulose. Meanwhile, $\text{LiCl} \cdot 4\text{H}_2\text{O}$ also showed catalytic
264 performance converting xylose to xylulose. This synergistic catalytic effect of $\text{LiCl} \cdot$
265 $4\text{H}_2\text{O}$ and $\text{Al}_2(\text{SO}_4)_3$ enabled a high yield of furfural. In conclusion, oligosaccharides
266 and xylose in **Liquid-1** were converted to sugar intermediates catalyzed by $\text{LiCl} \cdot 4\text{H}_2\text{O}$
267 and $\text{Al}_2(\text{SO}_4)_3$, which subsequently yielded furfural (**Figure. 2f**). The accumulated
268 furfural was solubilized in GVL layer, which significantly alleviated the side reactions
269 of sugar intermediates and resulted in a high conversion yield[41, 42]. To investigate
270 its potential in a practical application, catalyst and solvent recycling experiments were
271 conducted. Firstly, $\text{Al}_2(\text{SO}_4)_3$ is insoluble in $\text{LiCl} \cdot 4\text{H}_2\text{O}$ at room temperature, thus it

272 could be easily separated from the reaction mixture by filtration. GVL on the upper
 273 layer was recycled by removing furfural via a vacuum distillation. LiCl·4H₂O was
 274 recovered by employing the activated carbon to adsorb the incorporated byproducts
 275 [43]. It was shown that the recycled solvent did not show an obvious change in
 276 performance in separating hemicellulose and hemicellulose conversion after three
 277 cycles, when compared with the original solvent and catalyst (**Figure. 2e**).



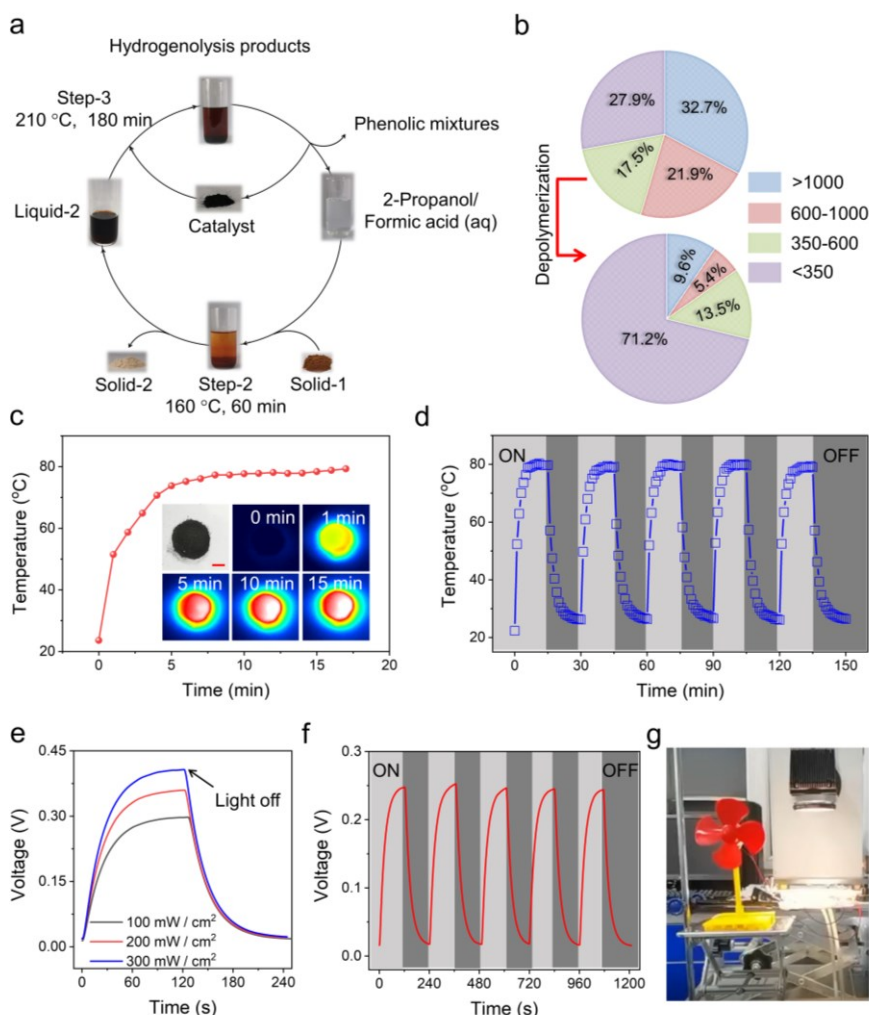
278
 279 **Figure. 2. Hemicellulose fractionation and conversion.** a) Schematic illustration of
 280 hemicellulose fractionation and conversion. b) Weight percentage of fractionated
 281 hemicellulose from lignocellulose at different reaction times, reaction conditions: 3.0 g
 282 bamboo, 20.0 g LiCl·4H₂O, 2 Mpa N₂, 130 °C. c) Components in fractionated
 283 hemicellulose. d) Conversion yield of Liquid-1/GVL and Xylan in LiCl·4H₂O/GVL to
 284 furfural at different times, reaction conditions: 21.1 g Liquid-1 or 0.89 g xylan, 2 Mpa
 285 N₂, 10.0 g GVL at 150 °C catalyzed by 0.1 g Al₂(SO₄)₃, calculation based on the
 286 hemicellulose in the lignocellulose. e) Reusability of the biphasic system
 287 (GVL/LiCl·4H₂O) and Al₂(SO₄)₃, the reaction parameters were consistent with the
 288 corresponding reaction steps above. f) Hydrolysis of xylan to furfural.

289 **3.2. Step 2: Lignin-to-photothermal materials conversion**

290 After hemicellulose fractionation, lignin was separated from **Solid-1** (Detailed
291 chemical components shown in Table S1) and depolymerized to a bio-oil (**Figure. 3a**).
292 Three main steps are involved: (i) lignin dissolution and cellulose separation; (ii) lignin
293 hydrogenolysis; and (iii) solvent and catalyst recovery. In the first step, **Solid-1** was
294 treated with isopropanol/formic acid (aq). 89 wt% of lignin was dissolved in the solvent
295 while most of cellulose (90.5 wt%) precipitated in solid state. After removing cellulose
296 via filtration, the lignin solution (**Liquid-2**) was hydrogenized assisted by various
297 catalysts (*e.g.*, Pd/C, Pt/C, Rh/C, and Ru/C). A crude oil, which consists mainly of
298 phenolic compounds, was obtained after recovering catalyst and isopropanol/formic (aq)
299 via simple liquid/solid separation and flash evaporation. GC-MS analysis showed that
300 Ru/C showed the best catalytic performance (**Table S2**). The conversion yield of mono-
301 and oligo- phenolics was 71.2 wt% (**Figure. 3b**). GPC analysis showed that the Mw/Mn
302 of lignin (in Liquid-2) decreased from 1298/786 Da (Mw > 600 Da accounting for
303 56.7%) to 916/575 Da (Mw > 600 Da accounting for 15.0%) (**Figure. 3b**). These results
304 suggested that lignin was efficiently converted into phenolic monomers/oligomers
305 catalyzed by Ru/C. It is worth noting that Ru/C still showed satisfactory catalytic
306 activity after 5 cycles (**Figure S2**). The recovered monomer-/oligomer- phenolics were
307 fully converted to photothermal reagents via coordinating with Fe³⁺. As shown in
308 **Figure S3 a-c**, the photothermal material is a crystal with obvious granular surface and
309 rich in hydroxyl group. UV-vis-NIR (**Figure S3d**) verified the following characteristics
310 of photothermal materials: 1) Extremely wide absorption spectra covering the whole

311 solar spectrum; 2) photothermal materials showed absorbance over a wide range of
312 wavelengths (200–2000 nm), which overlaps well with the solar spectrum; 3) It shows
313 strong absorption throughout the solar spectrum. These results show that the prepared
314 photothermal materials have excellent solar energy absorption capacity. By promoting
315 the absorbance and non-radiative migration of phenolics, such phenolics-iron complex
316 showed an efficient photothermal conversion [44]. The temperature of the photothermal
317 reagents increased from 24 °C to 79 °C during 15 min irradiation, suggesting that the
318 material is able to carry out efficient photothermal conversion (**Figure. 3c**). The effect
319 of irradiation on the materials was reversible, and there was no noticeable change in
320 photothermal conversion after five cycles (**Figure. 3d**). Encouraged by these results,
321 we then investigated the possibility of using the photothermal reagents to drive a
322 thermoelectric generator. Photothermal reagents, a thermoelectric module, and a
323 cooling system were combined to create a solar-powered thermoelectric generator. The
324 photothermal reagents absorbed simulated solar light (100 mW cm^{-2}) and heated the
325 thermoelectric module on one side, creating a temperature difference from the other
326 side nearest to the cooling system, and generating a voltage of 0.25 V (**Figure. 3e**).
327 Further increasing the mimetic solar intensity to 200 mW cm^{-2} and 300 mW cm^{-2}
328 triggered a voltage output of 0.35 V and 0.40 V, respectively. The as-prepared
329 thermoelectric generator was sensitive to photo irradiation. Switching off the light
330 source immediately reduced the voltage generated by the thermoelectric generator
331 (**Figure. 3e**), demonstrating good controllability of the device. The voltage output did
332 not show obvious change after 5 cycles of switching on/off the solar sources, suggesting

333 the superior stability of such solar-thermal-electricity conversion (**Figure. 3f**). More
 334 interestingly, a motion of engine was triggered by as-prepared solar-driven
 335 thermoelectric generators (**Figure. 3g** and **Video S1**).



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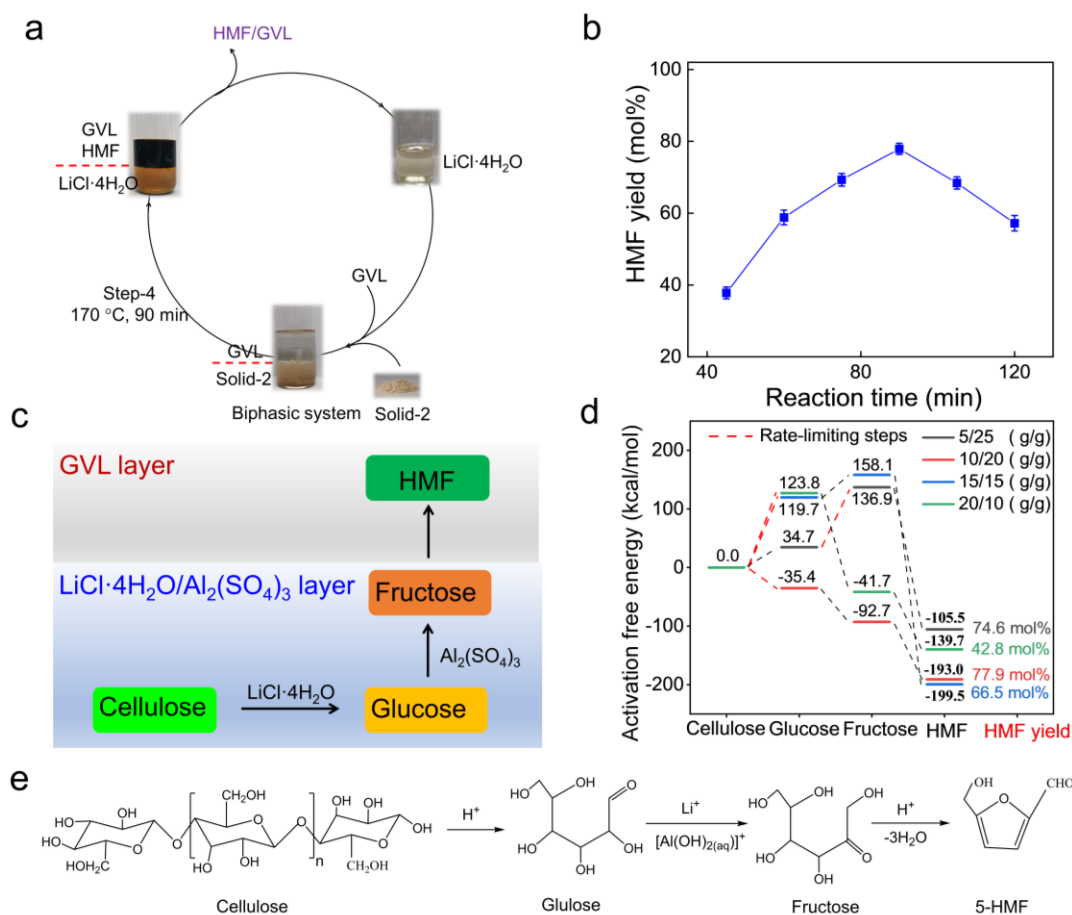
337 **Figure. 3. Depolymerization of lignin and functional conversion.** a) Schematic
 338 illustration of lignin fractionation and depolymerization. b) The components of lignin
 339 and depolymerized lignin, reaction conditions: 43.8 g Liquid-2, 0.2 g Ru/C, 2 Mpa H₂,
 340 210 °C for 4 h. c) Temperature changes of depolymerized lignin-derived photothermal
 341 materials, inset: the digital and IR images of depolymerized lignin-derived
 342 photothermal materials upon mimetic solar irradiation (100 mW cm⁻²), scale bar = 1
 343 cm. d) Recycled temperature changes of depolymerized lignin-derived photothermal
 344 materials upon switching on/off mimetic solar irradiation (100 mW cm⁻²). e) Voltage
 345 output of thermoelectric generators upon switching on/off mimetic solar irradiation at
 346 different intensities. f) Recycled voltage changes of thermoelectric generators upon
 347 switching on/off mimetic solar irradiation (100 mW cm⁻²). g) Digital images of fans
 348 driven by thermoelectric generators upon mimetic solar irradiation, scale bar = 2 cm.

349 **3.3. Step 3: Cellulose-to-HMF conversion**

350 Cellulose-rich **Solid-2** (92.2 wt%) was obtained after lignin fractionation from **Solid-1**
351 (**Figure. 4a**). **Solid-2** was further explored for its conversion to HMF in
352 LiCl·4H₂O/GVL. Catalyzed by the recycled Al₂(SO₄)₃ from the hemicellulose
353 conversion step, up to 77.9 mol% conversion yield of HMF was achieved (170 °C for
354 90 min) (**Figure. 4b**). As obtained HMF was well dissolved in the upper GVL layer and
355 could be easily obtained via a simple distillation. In order to explore the mechanism for
356 such high conversion yield, product evolution was analyzed. Glucose and fructose were
357 observed as intermediates in the reaction (**Figure. S4**), indicating that the reaction has
358 undergone cellulose hydrolysis, glucose isomerization, and fructose dehydration [45,
359 46]. To further explore the reaction mechanism, several control experiments were
360 conducted. In the molten salt hydrate LiCl·4H₂O solvent without Al₂(SO₄)₃, a 26.8 wt%
361 conversion yield of glucose was achieved (**Figure. S5**). Most of the glucose existed in
362 the LiCl·4H₂O, and no glucose was observed in GVL (**Figure. S6**). In contrast, only
363 5.3 wt% conversion yield of glucose was produced in Al₂(SO₄)₃ aqueous without the
364 LiCl·4H₂O at the same condition (**Figure. S7**). These results suggested that
365 depolymerization of cellulose is mainly promoted by LiCl·4H₂O rather than Al₂(SO₄)₃
366 and GVL. Moreover, 8.2 wt% and 23.7 wt% of fructose were obtained in the molten
367 LiCl·4H₂O and Al₂(SO₄)₃ aqueous, respectively. Similarly, no fructose was detected in
368 GVL (**Figure. S6**). These results suggest that the isomerization of glucose to fructose
369 is mainly catalyzed by Al₂(SO₄)₃. Furthermore, only a trace amount of HMF loss and
370 no structural change was observed when HMF was dissolved in GVL upon heating at

371 170 °C for 20 min, indicating that GVL efficiently stabilized as-produced HMF in the
372 reaction system (**Figure. S8**). In summary, the mechanism for converting cellulose to
373 HMF in the biphasic solvent was proposed. Cellulose was first depolymerized to yield
374 glucose assisted by the molten $\text{LiCl}\cdot 4\text{H}_2\text{O}$. After which, $\text{Al}_2(\text{SO}_4)_3$ triggered the
375 isomerization of glucose to fructose, which dehydrated to give HMF. The produced
376 HMF then entered the upper GVL layer and was efficiently dissolved in this layer
377 (**Figure. 4c**). The effect of ratios between $\text{LiCl}\cdot 4\text{H}_2\text{O}$ and GVL on the reaction was also
378 investigated. The sequence of HMF yields in different reaction media is as follows:
379 77.9 mol% (10/20, $\text{LiCl}\cdot 4\text{H}_2\text{O}$ /GVL, g/g) > 74.6 mol% (5/25, $\text{LiCl}\cdot 4\text{H}_2\text{O}$ /GVL, g/g) >
380 66.5 mol% (15/15, $\text{LiCl}\cdot 4\text{H}_2\text{O}$ /GVL, g/g) > 42.8 mol% (20/10, $\text{LiCl}\cdot 4\text{H}_2\text{O}$ /GVL, g/g)
381 (**Figure. 4d**). To understand such differences, molecular dynamics (MD) calculations
382 were conducted to show the free energy of each intermediate for cellulose conversion.
383 Glucose-fructose isomerization and cellulose-glucose depolymerization showed the
384 biggest and the second biggest energy barrier, 11.9 KJ/mol and 9.8 KJ/mol, respectively,
385 indicating these steps were the rate-limiting steps (**Figure. S9**). Further calculation
386 results for the energy barrier values of the rate-limiting steps in reactions with different
387 $\text{LiCl}\cdot 4\text{H}_2\text{O}$ /GVL ratios were as follows: -35.4 KJ/mol (10/20, $\text{LiCl}\cdot 4\text{H}_2\text{O}$ /GVL, g/g) <
388 102.2 KJ/mol (5/25, $\text{LiCl}\cdot 4\text{H}_2\text{O}$ /GVL, g/g) < 119.7 KJ/mol (15/15, $\text{LiCl}\cdot 4\text{H}_2\text{O}$ /GVL,
389 g/g) < 123.8 KJ/mol (20/10, $\text{LiCl}\cdot 4\text{H}_2\text{O}$ /GVL, g/g) (**Figure. 4e**). Thus, the rate-limiting
390 steps showed the lowest energy barrier when the weight ratio between $\text{LiCl}\cdot 4\text{H}_2\text{O}$ and
391 GVL was 10:20. Such a low energy barrier was beneficial for the cellulose-HMF
392 conversion and induced a high conversion yield. Interestingly, the conversion yield was

393 even higher than the value of microcrystal cellulose-to-HMF conversion (70.2 mol%)
 394 at the same condition. This effect was attributed to the lower crystallinity of cellulose
 395 in **Solid-2**, compared to the microcrystal cellulose, which was proved by XRD, FTIR,
 396 and FTIR (**Figure. S10**). The lower crystallinity facilitated accessibility of solvents and
 397 catalyst to the cellulose in **Solid-2**, which promoted the conversion. As a result, energy
 398 consumption analysis suggests that to produce a considerable amount of HMF, the
 399 energy consumed by microcrystal cellulose is 1.16 times (2.117 kWh) that of as-
 400 obtained cellulose (1.822 kWh) (**Figure. S11**).

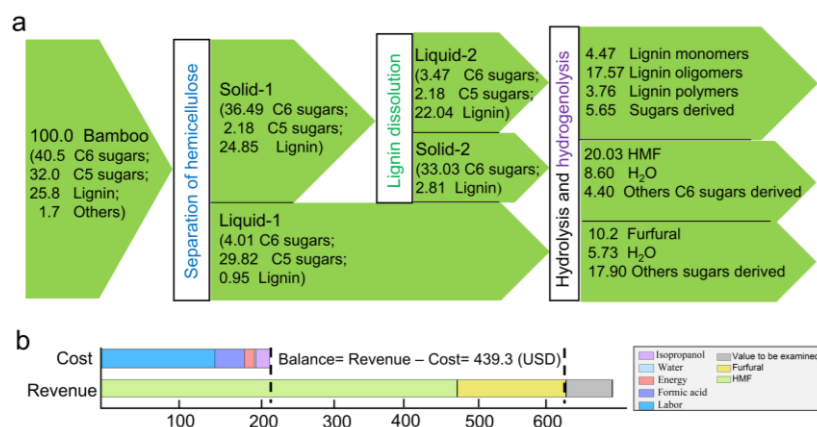


401 **Figure. 4. Cellulose-to-HMF conversion.** a) Schematic illustration of cellulose-to-
 402 HMF conversion. b) Conversion yield of HMF in LiCl·4H₂O/GVL at different times,
 403 reaction conditions: 0.54 g Solid-2, 10.0 g LiCl·4H₂O, 2 Mpa N₂, 20.0 g GVL, and
 404 0.05g Al₂(SO₄)₃ at 170 °C catalyzed by Al₂(SO₄)₃. c) Proposed mechanism for
 405 Cellulose-to-HMF conversion in LiCl·4H₂O/GVL catalyzed by Al₂(SO₄)₃. d) The
 406 activation free energy profile of each reaction intermediate in a biphasic solvent. e)
 407

408 Hydrolysis of cellulose to HMF.

409 **3.4. Comprehensive mass balance of biorefinery strategy**

410 Based on the obtained experimental data, technical-economic was conducted on the
 411 integrated hemicellulose-first biorefinery (**Figure. 5a**). In the model process, 20.03 kg
 412 HMF, 10.2 kg furfural, 25.8 kg photothermal materials, 27.94 kg sugars derived (for
 413 bioethanol) and 14.33 kg H₂O could be obtained from 100 kg lignocellulose. Market
 414 investigation showed that the price of HMF and furfural is 500.75 USD and 153.0 USD,
 415 respectively. The assumptions in the biorefinery are listed in **Table S3**. Labor cost
 416 (143.36 USD) is the biggest contributor to the costs. Due to the loss of formic acid
 417 recovery process, formic acid consumption section is the second highest contributor to
 418 the cost (~39.0 USD). It is worth noting that the value of lignin-derived photothermal
 419 materials is waiting for further evaluation. Overall, our proposed process can generate
 420 revenue of 439.3 USD by processing 100 kg of lignocellulose (**Figure. 5b**).



421

422 **Figure. 5. Mass balance and economics for the bamboo powder biorefinery**
 423 **strategy.** a) bamboo fractionation and full-component conversion process. b) costs and
 424 revenues. Reaction conditions: Separation of hemicellulose (130 °C for 1 h, 2Mpa N₂),
 425 Lignin dissolution (160 °C for 1 h, 2Mpa N₂), Hydrolysis (cellulose: 150 °C for 0.75 h,
 426 hemicellulose: 170 °C for 1. 5 h, 2 Mpa N₂), Hydrogenolysis (210 °C for 4 h, 2Mpa H₂).

427 **4. Conclusion**

428 In summary, we have proposed a sustainable and profitable biorefinery strategy in this

429 paper via a recyclable biphasic solvent ($\text{LiCl}\cdot 4\text{H}_2\text{O}/\text{GVL}$). Hemicellulose, lignin, and
430 cellulose were fractionated from lignocellulose at a high yield. Subsequently, the
431 fractionated hemicellulose and cellulose showed high reactivity and can be efficiently
432 converted to furfural and HMF, respectively. Moreover, depolymerized products from
433 as-separated lignin were fully converted to photothermal materials by coordinating with
434 iron ions for solar-thermal-electricity conversion without any separation and
435 purification, which overcomes the limitations in traditional lignin biorefinery. The
436 rational design enabled a revenue of 439.3 USD by processing 100 kg of lignocellulose.
437 More importantly, all the catalysts and chemicals in the developed biorefinery are
438 commercially available. As a result, our method is applicable and very promising in the
439 practical industry. Generally, this work is expected to give a new strategy and practical
440 solution to produce valuable chemicals and functional materials from lignocellulosic
441 biorefinery

442 **Author contributions**

443 C.L. carried out the lignocellulose fractionation and depolymerization experiment. X.Z.
444 performs photothermal reagent preparation experiment. J.H. and X.Z. perform
445 molecular dynamics calculation experiment. X.Y. and T.C. perform the experiment of
446 the chemical component of lignocellulose. C.L. wrote the manuscript assisted by K.W.,
447 Z.C. and X.M.; K.W., J.J. and A.J.R. conceived the work, and all authors discussed the
448 experimental and analyzed the data. X.M. and A.J.R. efforts were funded by the
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450 **Competing interests**

451 The authors declare no competing interests.

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459 **Data availability:** The authors declare that all of the data that support the findings of
460 this study are available within the article and its Supplementary Information files or
461 from the corresponding author upon reasonable request.

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