

1 **Full title:** Risk factors, temporal dependence, and seasonality of human ESBL-producing *E.*
2 *coli* and *K. pneumoniae* colonisation in Malawi: a longitudinal model-based approach.

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29

30 **Abstract**

31 Background

32 Antimicrobial resistance (AMR) represents an important threat to achieving the sustainable
33 development goals in Sub-Saharan Africa (sSA). sSA is reported to have the highest
34 estimated death rate attributable to AMR, with Extended-Spectrum Beta-Lactamase-
35 producing Enterobacterales, such as *Klebsiella pneumoniae* and *Escherichia coli*,
36 representing the greatest challenge. However, the dynamics of human colonisation with such
37 bacteria in the sSA community setting are not well known. Inadequate water, sanitation and
38 hygiene (WASH) infrastructure and associated behaviours are thought to play an important
39 role in transmission of AMR-bacteria, and an improved understanding of the temporal
40 dynamics of within-household transmission could help inform the design of public health
41 policies that interrupt transmission of AMR-bacteria.

42 Methods and Findings

43 In this 18-month study, individuals from households in diverse areas of Southern Malawi
44 were recruited and human stool samples were longitudinally collected. Using microbiological
45 data and household surveys, we built a multivariable hierarchical harmonic logistic regression
46 model to identify risk factors for colonisation with ESBL-producing *E. coli* and *K.*
47 *pneumoniae*, reflecting household structure and temporal correlation of colonisation status
48 between timepoints.

49 Important risk factors were identified, with men having a lower risk of becoming colonised
50 with ESBL-producing *E. coli* (OR 0.786 CrI[0.678-0.910]) and the use of a tube well or a
51 borehole as a water drinking source highly increasing the risk of becoming colonised (OR
52 1.550 CrI[1.003-2.394]). Coming into contact with standing water also appeared to be
53 negatively associated with colonisation status (OR 0.749 CrI[0.574-0.978]). For ESBL-
54 producing *K. pneumoniae*, having recently taken a course of antibiotics increased the risk of
55 being colonised (OR 1.281 CrI[1.049-1.565]). We also found a negative association between
56 eating from shared plates and colonisation with ESBL-producing *K. pneumoniae* (OR 0.672
57 CrI[0.460- 0.980]). Finally, we detected a temporal correlation range of eight to eleven weeks,
58 providing evidence that within-household transmission occurs within this time frame.

59 Conclusions

60 We suggest that interventions aimed at preventing transmission might have the best impact
61 when targeted at the household-level and focused on a combination of improving WASH
62 infrastructure and modifying associated behaviours. Additionally, we showed that antibiotic
63 use is important when looking at colonisation with ESBL-producing *K. pneumoniae* and
64 therefore infection prevention and control measures and antibiotic use and stewardship
65 training could help in reducing transmission.

66

67 Introduction

68 In 2015, the World Health Organisation (WHO) declared antimicrobial resistance
69 (AMR) as one of the 10 global priority public health threats [1]. The rise of AMR
70 internationally is endangering the achievement of the sustainable development goals,
71 particularly those relating to health, poverty, food security and economic growth [2]. The
72 latest estimates of the global burden of AMR showed that 4.95 million deaths were associated
73 with bacterial AMR in 2019, among which 1.27 million deaths were attributable to bacterial
74 AMR [3]. The death rate attributable to AMR was found to be highest in sub-Saharan Africa
75 (sSA), where the leading pathogens were *Klebsiella pneumoniae*, *Streptococcus pneumoniae*
76 and *Escherichia coli*, however large gaps in data availability were noted in sub-Saharan
77 Africa [3]. Of particular note has been the rapid emergence of Extended-Spectrum beta-
78 Lactamases (ESBL) in gram negative bacteria [4], potentially as a result of increased use of
79 3rd generation cephalosporin antimicrobials in empirical treatment of bacterial infection in
80 hospitals. Whilst studies have shown that the prevalence of infections caused by ESBL-
81 producing Enterobacterales in sSA is high [5,6,7], little is known about asymptomatic
82 colonisation with ESBL-producing Enterobacterales, a key step prior to infection in
83 vulnerable patient groups. Learning more about asymptomatic colonisation in the community
84 is therefore crucial in order to prevent transmission, and as a consequence, reduce drug-
85 resistant infections.

86 Prior to 2016, no studies described risk factors for colonisation with ESBL-producing
87 Enterobacterales among healthy individuals in the community in sSA[5]. More recently,
88 recent antibiotic use (in the last weeks to months) was found to be a risk factor in a few
89 community-based studies [8,9,10]. Other risk factors such as older age and previous hospital
90 admission were also identified [8]. A further study found that higher income was associated

91 with a higher prevalence of ESBL colonisation [11]. However, most of these studies focused
92 on a specific subset of the community and not the general population. We cannot, therefore,
93 be confident that the risk factors detected in these specific populations would be the same
94 throughout the general population. This highlights the need for a community-based study
95 within the general population to identify risk factors for human gut mucosal colonisation with
96 ESBL-producing Enterobacterales. Moreover, although it is strongly believed that
97 inadequacies in water, sanitation and hygiene (WASH) infrastructure and associated
98 behaviours play an important role in transmission of AMR-bacteria [12], risk factors related
99 to WASH in this context are still not well described. For example, only one study found that
100 having private indoor access to drinking water was positively associated with ESBL
101 colonisation [13].

102 To be able to reduce transmission of, and colonisation by, AMR-enteric bacteria in
103 East and Southern Africa, we need to explore the dynamics of colonisation in order to tailor
104 appropriate interventions; for example the role of seasonality in colonisation, and how long
105 colonisation lasts after initial acquisition. A first step in understanding the temporal dynamics
106 of within-household transmission is to determine how long a specific household is at risk of
107 colonisation once one member has been colonised, this will help inform the design of public
108 health policies that interrupt transmission of AMR-bacteria. Such interventions will be
109 impactful at interrupting transmission of enteric pathogens more broadly.

110 Here, we describe an analysis of an 18-month longitudinal cohort study using
111 microbiological, household and WASH surveys, where WASH refers to water, sanitation
112 (containment of both human and animal faeces), hygiene and food hygiene. We fit a serially-
113 correlated generalised linear mixed model, exploring household, individual and WASH risk
114 factors for ESBL colonisation in settings with different degrees of urbanisation in Malawi.

115 This allows us to separate the effects of individual- and household-level risk factors from
116 temporal effects we observe in our data.

117 **Methods**

118 The Drivers of antimicrobial Resistance in Uganda and Malawi (DRUM) consortium
119 was an interdisciplinary consortium working across urban, peri-urban and rural communities
120 in Uganda and Malawi (www.drumconsortium.org). Its aim was to study AMR transmission
121 in a One Health setting, sampling areas with different human and animal population densities,
122 and different levels of affluence and infrastructure. DRUM was a repeated-measures study in
123 which individuals, clustered into households, were sampled at four timepoints over 6 months.
124 The detailed protocol is available at [14].

125 This analysis focuses on a subset of the data from the DRUM study, focusing on the
126 3 Malawi study areas: Ndirande, Chileka, and Chikwawa representative of urban, peri-urban,
127 and rural demographics respectively. We model the presence or absence of gut-mucosal
128 colonisation with ESBL *E. coli* and *K. pneumoniae* in individuals, and aim to detect
129 associations with individual-level demographic and health characteristics, household-level
130 WASH indicators, and the social context represented by the study area. In order to capture
131 seasonality and household structure, we used a hierarchical multivariable harmonic
132 regression, with temporal correlation at the household level. As described in the following
133 sections, we first screened our candidate covariates through expert opinion and univariable
134 logistic regression models, before taking these variables forward into our full modelling
135 framework.

136 **Covariate selection**

137 In order to investigate WASH practices at the household-level, household covariates
138 were collected in the following ways: reported variables (i.e. *presence* of a toilet at the
139 household) were based on questions asked to the study participants during the baseline
140 assessment visit, whilst observed variables (i.e. *type* of toilet), were answered by field teams
141 observing the household infrastructure at multiple time points. These variables were screened
142 for importance by a panel of environmental health specialists (Morse, Chidziwisano) with
143 expertise on the risks and critical control points for faecal-oral transmission in Malawi
144 [15,16,17].

145 Individual-level covariates such as age, gender, HIV status and antibiotic use were
146 also selected. Antibiotic use was defined as the reported use of any antibiotics in the last six
147 months (at baseline) and subsequently, for each follow-up visit, as the reported use of
148 antibiotics between that visit and the previous one. We excluded long-term antibiotic therapy
149 such as cotrimoxazole prophylactic therapy (CPT) in order to capture only the immediate
150 change expected to occur following short courses of therapy. Household-level covariates such
151 as household income and size were also included.

152 Stool samples from participants were cultured for growth of ESBL-producing bacteria
153 on ESBL CHROMagar™ chromogenic agar (CHROMagar™, France). Bacterial colonies
154 were classified by colour into categories, and speciation of blue colonies took place to
155 identify ESBL-producing *K. pneumoniae* using polymerase chain reaction (PCR) as
156 previously described in [14]. Throughout this paper, colonisation refers to asymptomatic
157 carriage of either of these pathogens.

158 Household, individual and WASH datasets were merged together to form a unique
159 dataset and all variables went through cleaning, removing incomplete and duplicate records
160 assuming missingness at random.

161 Univariable logistic models were then used to investigate the effect of WASH
162 infrastructure and associated behaviours on colonisation with either ESBL-producing *E. coli*
163 or ESBL-producing *K. pneumoniae*. We started by looking at the effect of the study area on
164 colonisation in order to separate its potential effect from the effects of other risk factors and
165 then ran the other univariable models accordingly. This allowed for a refinement of the
166 variables to be included in the full modelling framework, by retaining only the covariates
167 which returned a p-value <0.2. For both the ESBL-producing *E. coli* and the ESBL-
168 producing *K. pneumoniae* models, we pragmatically set this threshold to avoid missing the
169 identification of important covariates.

170

171 **Multilevel harmonic hierarchical regression**

172 The modelling framework is detailed in S1 Appendix. Briefly, we used a hierarchical
173 multivariable harmonic logistic regression, incorporating covariates selected by the
174 univariable analysis. In addition to the covariates, we included study region, annual and bi-
175 annual harmonic terms to capture seasonality, and a serially-correlated household-level
176 random effect to reflect temporal correlation in household-level prevalence between
177 timepoints as well as heterogeneity in prevalence between households over and above that
178 explained by the covariates. The analysis was carried out twice, for each of our *E. coli* and *K.*
179 *pneumoniae* binary colonisation response variables.

180 A Bayesian approach to inference was used to allow for more flexibility in designing
181 our model, using prior information from recent work on the dynamics of gut mucosal
182 information with ESBL-producing Enterobacterales in Malawi [18] and to explore different
183 forms of the temporal correlation structure at the household level. The model was fitted using
184 the standard implementation of the No-U-Turn Sampler (NUTS) in the Stan modelling

185 language [19] in R v4.1.1 through the package RStan [20]. The model was run with 20000
186 iterations for each of three independent Markov chains. Convergence was evaluated by
187 inspection of trace plots and the Gelman-Rubin statistic being close to 1. Posterior estimates
188 of parameters were expressed as medians with 95% credible intervals.

189 Initial model exploration considered adding the serially-correlated random effect at
190 the individual-level, but found no evidence of individual-level temporal correlation. We
191 therefore applied the serial correlation to a household-level random effect, enabling us to
192 quantify both a household-effect (over and above that due to observed covariate information)
193 and to account for possible “household contamination” with an ESBL as a result of a longer-
194 term transmission process operating within the home. To ensure maximum identifiability of
195 the random effects, we performed our analysis only on households with available sample
196 results for all four time points. Throughout this analysis, the harmonic terms were always run
197 as a single term. The variables were standardised, therefore the odds ratio should be
198 interpreted as a change for each increase in standard deviation.

199 Ethical approval was obtained from Liverpool School of Tropical Medicine (LSTM)
200 Research and Ethics Committee, UK (REC, #18-090) and College of Medicine Research and
201 Ethics Committee, Malawi (#P.11/18/2541).

202

203 **Results**

204 **Exploratory analysis**

205 Twenty-six WASH variables were selected by the expert panel for further analysis.
206 Among these variables, three were removed due to a lack of variation. Twenty-seven
207 households were excluded due to missing enrolment or WASH data. Four individuals were

208 removed due to missing data on age and gender, eighteen duplicate records were removed.
209 Out of 2845 human stool samples collected by the field teams over time, forty-four duplicate
210 records were removed. After merging, 224 samples were removed due to missing covariate
211 data, and 84 samples from households ≥ 200 meters outside of the polygon limits, were also
212 removed. This threshold permitted retention of households subsequently chosen by the field
213 teams after refusal from the original sampled household. The combined dataset, which was
214 used for the analysis, contained 2493 samples from 894 individuals in 259 households. The
215 complete list of covariates and outcome variables can be found in S1 Table.

216 The distribution of samples, individuals, and households per polygon are presented in
217 Table 1. Although the number of households and individuals in Ndirande and Chileka was
218 higher than in Chikwawa, the number of available samples was similar with 36% in
219 Chikwawa and Chileka and 28% in Ndirande.

220 **Table 1. Distribution of the number of households, individuals and samples per polygon**

	Ndirande	Chikwawa	Chileka	Total
Households	96	64	99	259
Individuals	285	259	350	894
Samples	709	891	893	2493

221

222 After data cleaning, 233 individuals had one sample, 96 had two samples, 192 had
223 three samples and 373 individuals had all four samples. The 129 households in which
224 individuals had four available samples were used for the hierarchical model.

225 **Fig 1. Distribution of samples collected, over time and by study area**

226 **Baseline participant data**

227 Our age distribution data reflect Malawi's population structure (Fig 2). People were
228 considered adults at ≥ 16 years (54%, 483/894) and school age was ≥ 5 to 15 years (27.1%,
229 242/894), whilst 57.1% (510/894) participants were female with slightly varying proportions
230 in each age group (65.2% (315/483) of adults and 46.7% (113/242) of school age children).

231 **Fig 2. Distribution of age and gender for the 894 individuals**

232 At the first visit, 15.2% (129/851) of participants reported having taken at least one
233 course of antibiotics in the preceding six months, while between subsequent visits, 6%
234 (37/616), 9.4% (54/570) and 8.3% (38/456) reported at least one course of antibiotics. This
235 varied by region, with 15.4% (137/891) in Chikwawa, 9.3% (66/709) in Ndirande and 6.2%
236 (55/893) in Chileka.

237 Overall, the prevalence of ESBL-producing *E. coli* in our samples was 37%
238 (922/2493) and the prevalence of ESBL-producing *K. pneumoniae* was 11.9% (296/2493).
239 The prevalence of colonisation with ESBL-producing *E. coli* and ESBL-producing *K.*
240 *pneumoniae* in our samples varied over time (Fig 3). The fluctuations in prevalence for both
241 bacteria appear to follow a similar pattern in time. Some evidence of seasonality can be
242 discerned for both, although more evidently for *E. coli*, with a decrease in prevalence during
243 the dry season (May to October) followed by an increase during the wet season (November to
244 April). This is followed by a sudden drop between April and July 2020, a period for which
245 we have no available data due to Covid-19, and an increase in the last few months followed
246 by a decrease in November 2020.

247 **Fig 3. Prevalence of ESBL-producing *E. coli* and ESBL-producing *K. pneumoniae* per** 248 **month.**

249 **Baseline WASH data**

250 Correlations between household-level WASH variables are depicted on the heatmap
251 in Fig 4. There was a strong positive correlation between multiple animal-exposure related
252 factors, for example bird owners appeared to be more likely to keep animals inside, therefore
253 it was also more likely for these animals to come into contact with food preparation areas.
254 Keeping animals inside also increased the likelihood of visible animal faeces around the
255 household area. In terms of sanitation, the data suggested that with increasing income, there
256 was increasing likelihood the household's water drinking source came from a pipe, rather
257 than a tube well or borehole. Moreover, increasing household income correlated with
258 presence of hand washing facilities and soap presence in the household, and presence of
259 cleaning materials such as toilet paper near the toilet. Some food consumption factors such as
260 eating from shared plates appeared to be negatively correlated with the previously mentioned
261 sanitation factors - the higher the income of the household is, the less chance individuals used
262 shared plates when eating.

263 **Fig 4. Correlation heatmap of household-level covariates**

264

265 **Impact of WASH infrastructure and associated behaviours on ESBL**
266 **colonisation**

267 In order to start exploring the effect of WASH variables on colonisation with either
268 ESBL-producing *E. coli* or ESBL-producing *K. pneumoniae*, and to look at variability
269 between regions, we first ran a generalised linear model including only the study area for
270 both ESBL-producing *E. coli* and ESBL-producing *K. pneumoniae* (Table 2 and 3). We
271 found a significant effect of the study area on ESBL-producing *E. coli*, with a higher risk of
272 being colonised in Ndirande (OR 1.39 CI[1.13-1.70]) compared to Chileka. This was not the
273 case for ESBL-producing *K. pneumoniae*, which showed no significant effect of the study

274 area on the colonisation status. We observed that in the case of *E. coli*, WASH variables that
275 vary depending on the study area the participant was in had a different signal if the study area
276 was included as a variable in the analysis. Consequently, univariable analysis was run
277 differently depending on the bacterial species with the study area included as a covariate
278 when running univariable models for ESBL-producing *E. coli*, but not for ESBL-producing *K.*
279 *pneumoniae*.

280 **Table 2. Relationship between the study area and the ESBL-producing *E. coli* colonisation**
281 **status**

	Log-odds	P-value	Odds ratio (95% CI)
Intercept	-0.607	<2e-16	0.545 (0.475-0.625)
Living in Chikwawa (vs Chileka)	-0.061	0.540	0.941 (0.774-1.144)
Living in Ndirande (vs Chileka)*	0.326	0.002	1.385 (1.132-1.696)

282 *Significant variables highlighted in bold

283 **Table 3. Relationship between the study area and the ESBL-producing *K. pneumoniae***
284 **colonisation status**

	Log-odds	P-value	Odds ratio (95% CI)
Intercept	-2.105	<2e-16	0.122 (0.099-0.151)
Living in Chikwawa (vs Chileka)	0.196	0.183	1.216 (0.912-1.622)
Living in Ndirande (vs Chileka)	0.098	0.536	1.103 (0.809-1.504)

285

286 **Human gut mucosal colonisation with ESBL-producing *E. coli***

287 Results from the univariable models showed that having a drinking water source
288 coming from a tube well or a borehole, having a drop hole cover on the toilet and animals
289 being able to enter into contact with the food areas all appeared to be highly significant (S2
290 Table). Whilst a positive association with ESBL-producing *E. coli* colonisation status was

291 detected for the drinking water source coming from a tube well or a borehole, the opposite
292 can be said for piped drinking water. This was consistent with the negative correlation
293 between those two water variables noted previously in the correlation heatmap. Animal
294 contact with food areas was positively associated with ESBL-*E. coli* colonisation, whilst
295 having a drop hole cover on the toilet appeared to have a protective effect, being negatively
296 associated with colonisation status. Additionally, variables such as keeping animals inside the
297 house, having a toilet floor surfaced with soil and having clean paper in the toilet were also
298 highly significant. Having clean paper in the toilet was negatively associated with
299 colonisation status while the others showed a positive association. Other variables such as
300 older age, the presence of open defecation in the area, owning cattle, sheep or goats, entering
301 into contact with river water all were significant (<0.05) and were positively associated with
302 the colonisation status. In contrast, male sex, higher income, having a disposal mechanism for
303 animal waste, having a piped water drinking source, storing water in a container with lid and
304 tap were all significantly negatively associated with ESBL-*E. coli* colonisation status (S2
305 Table).

306 We subsequently ran the hierarchical model. We found that men are less at risk of
307 becoming colonised with ESBL-producing *E. coli* (OR 0.786 CrI[0.678-0.910]) and that
308 having a tube well or a borehole as a water drinking source highly increases your risk of
309 becoming colonised (OR 1.550 CrI[1.003-2.394]). Coming into contact with standing water
310 also appeared to be negatively associated with colonisation status (OR 0.749 CrI[0.574-
311 0.978]). Finally, there was an apparent signal of annual seasonality noticeable from the
312 presence of part of the harmonic term (Table 4).

313 Using the covariance structure (S1 Appendix), we found a range of temporal
314 correlation estimated at 77.85 days (CrI [30.85-140.60]), thus samples that have been
315 sampled in the same household more than 77 days apart are effectively uncorrelated.

316 Parameter estimates are shown in S3 Table. The densities of the priors and posteriors of all
 317 three parameters can be found in S1 Fig. Visual inspection of the trace plots in S2 Fig and
 318 calculations of the Gelman-Rubin statistic resulting close to 1 for all parameter estimates
 319 indicates that the model has fitted properly.

320

321 **Table 4. Temporal model results for ESBL-producing *E. coli* colonisation status**

	Log-odds	Odds ratio (95% CrI)
(Intercept)	-0.716	0.489 (0.360-0.663)
Reactive to HIV testing (vs non-reactive)	0.027	1.027 (0.863-1.223)
Unknown HIV status (vs non-reactive)	-0.031	0.969 (0.808-1.163)
Recent use of antibiotics	0.093	1.097 (0.946-1.274)
Age	0.132	1.141 (0.970-1.343)
Being male (vs female)*	-0.241	0.786 (0.678-0.910)
Average household monthly income	0.226	1.254 (0.916-1.715)
Open defecation	0.054	1.055 (0.826-1.349)
Presence of a disposal mechanism for animal waste	0.104	1.110 (0.857-1.437)
Eating from shared plates	-0.245	0.783 (0.598-1.024)
Having a pipe as drinking water source	0.132	1.141 (0.818-1.592)
Having a tube well/ borehole as drinking water source	0.438	1.550 (1.003-2.394)
Use of alternative water for cleaning utensils	0.014	1.014 (0.802-1.283)
Owning cattle, goats or sheep	0.139	1.149 (0.892-1.480)
Keeping animals inside	0.075	1.078 (0.852-1.364)
Contact with river water	0.048	1.049 (0.791-1.391)
Toilet floor material: none (vs concrete/wood)	0.123	1.131 (0.799-1.600)
Toilet floor material: soil (vs concrete/wood)	0.131	1.140 (0.820-1.585)

Presence of drop hole cover on the toilet	-0.202	0.817 (0.626-1.067)
Presence of newspaper/paper in the toilet	-0.155	0.856 (0.670-1.094)
Frequency of soap presence in handwashing facilities	-0.000	1.000 (0.640-1.563)
Storing water covered	-0.179	0.836 (0.537-1.302)
Storing water in a container with lid/tap	-0.034	0.967 (0.754-1.240)
Contact between animal and food areas	0.218	1.244 (0.983-1.573)
Presence of standing water around the household	-0.289	0.749 (0.574-0.978)
Number of days since the first sample	0.167	1.182 (0.869-1.608)
Harmonic term (sinday)	-0.466	0.628 (0.453-0.869)
Harmonic term (cosday)	0.371	1.449 (0.958-2.191)
Harmonic term (sinday2)	-0.084	0.919 (0.644-1.314)
Harmonic term (cosday2)	0.018	1.018 (0.711-1.457)
Living in Chikwawa (vs Chileka)	-0.276	0.759 (0.527-1.093)
Living in Ndirande (vs Chileka)	0.386	1.471 (0.980-2.207)

322 *Significant variables highlighted in bold

323

324 **Human gut mucosal colonisation with ESBL-producing *K. pneumoniae***

325 In the case of ESBL-producing *K. pneumoniae*, univariable results showed that the
326 household size was the only highly significant variable ($p < 0.01$) except for the harmonic
327 terms, revealing that with increasing household size, there was greater the risk of being
328 colonised with ESBL-*K. pneumoniae*. Variables such as eating street food, eating from
329 shared plates, owning birds and coming into contact with drains were also significant. Eating
330 street food and eating from shared plates surprisingly appeared to have a protective effect on
331 the ESBL-producing *K. pneumoniae* colonisation status. Owning birds and coming into
332 contact with drains were both positively associated with colonisation status (S4 Table).

333 The hierarchical model for ESBL *K. pneumoniae* found that having previously used
 334 antibiotics (in the last six months or in-between visits) increased the risk of being colonised
 335 with ESBL-producing *K. pneumoniae* (OR 1.281 CrI[1.049-1.565]). We also saw a negative
 336 association between eating from shared plates and colonisation (OR 0.672 CrI[0.460- 0.980]).
 337 Finally, there was a signal of annual seasonality noticeable from the presence of part of the
 338 harmonic term, similar to the one we found for ESBL *E. coli*. These results are presented in
 339 Table 5.

340 **Table 5. Temporal model results for ESBL-producing *K. pneumoniae* colonisation status**

	Log-odds	Odds ratio (95% CrI)
(Intercept)	-3.432	0.032 (0.017-0.060)
Recent use of antibiotics*	0.248	1.281 (1.049-1.565)
Number of people living in the household	0.298	1.347 (0.932-1.947)
Presence of a toilet in the household	-0.136	0.873 (0.468-1.628)
Eating street food	0.091	1.095 (0.797-1.505)
Eating from shared plates	-0.398	0.672 (0.460-0.980)
Having a pipe as drinking water source	-0.253	0.776 (0.506-1.190)
Having a communal tap as drinking water source	-0.423	0.655 (0.408-1.051)
Use of alternative water for cleaning utensils	-0.241	0.786 (0.563-1.097)
Owning birds	0.015	1.015 (0.706-1.459)
Owning dogs or cats	0.024	1.024 (0.735-1.427)
Owning pigs	0.157	1.170 (0.853-1.604)
Contact with drains	0.219	1.245 (0.906-1.710)
Toilet type: other (vs no toilet)	0.203	1.225 (0.752-1.996)
Toilet type: pit latrine (vs no toilet)	0.204	1.226 (0.550-2.734)
Toilet type: shared toilet (vs no toilet)	-0.395	0.674 (0.395-1.148)
Visible human faeces around the household	0.224	1.251 (0.881-1.777)

Storing water uncovered	-0.326	0.722 (0.478-1.089)
Number of days since the first sample	-0.066	0.936 (0.587-1.493)
Harmonic term (sinday)	-0.753	0.471 (0.289-0.767)
Harmonic term (cosday)	0.448	1.565 (0.883-2.774)
Harmonic term (sinday2)	-0.304	0.738 (0.434-1.255)
Harmonic term (cosday2)	0.483	1.621 (0.961-2.736)
Living in Chikwawa (vs Chileka)	-0.038	0.963 (0.606-1.529)
Living in Ndirande (vs Chileka)	0.274	1.315 (0.812-2.130)

341 *Significant variables highlighted in bold

342

343 We found a range of temporal correlation for ESBL-*K. pneumoniae* colonisation
344 estimated at 54.29 days (CrI [12.91-130.43]), thus samples that have been sampled in the
345 same household more than 54 days apart are effectively uncorrelated. Parameter estimates are
346 shown in S5 Table. The densities of the priors and posteriors of all three parameters can be
347 found in S3 Fig. Convergence was verified by looking at the trace plots in S4 Fig and we
348 confirmed that the Gelman-Rubin statistic was close to 1 for all parameter estimates.

349

350 Discussion

351 This study identified varying prevalence of ESBL colonisation over time for both
352 ESBL-producing *E. coli* and ESBL-producing *K. pneumoniae*. A decrease in prevalence was
353 observed during the dry season, followed by an increase during the wet season, and this
354 apparent seasonality was confirmed by the model results. Potential explanations for this
355 variation include the accumulation of mud and floodwater due to the heavy rain in the wet
356 season, which might lead to more contact between individuals and contaminated soil or water.

357 Additionally, the increase in time spent indoors when heavy rain occurs might lead to higher
358 within-household transmission.

359 The correlation heatmap (Fig 4) suggested that the socioeconomic status of the
360 household greatly influences the WASH situation of the household, and that higher income
361 allows for a better access to cleaner water and easier availability of sanitation and hygiene
362 products. In particular, there was a positive association between using a tube well or a
363 borehole as a drinking water source and being ESBL-colonised, which is confirmed in the
364 temporal model for ESBL-producing *E. coli*. Being female was also identified as a risk factor
365 for ESBL-producing *E. coli*, which could be explained by the fact that traditionally women
366 are more likely to perform domestic duties -- such as laundry, housework, and childcare --
367 which would place them at higher risk of being in contact with the faecally-contaminated
368 environment. However, no direct association was found between income and gut mucosal
369 colonisation in the model. Further work is needed to understand the association between
370 income and gut mucosal colonisation, but it is noteworthy that the vast majority of
371 households in the study were below the World Bank defined threshold of absolute poverty
372 (<\$1.90/day per individual) and that income alone is a poor indicator of wealth.

373 Other variables identified by the univariable models as conferring a highly significant
374 increased risk of being colonised with ESBL-producing *E. coli* included the study area and
375 permitting animals inside the home, and allowing them to contact food preparation areas.
376 This is common practice in low- and middle-income countries (LMICs), where animal
377 husbandry is frequently a primary source of income [21]. However, this practice increases the
378 risk of faecal contamination of the soil by enteric pathogens like *E. coli* [22] and therefore
379 puts household members, especially young children, at higher risk for exposure to faecal
380 pathogens and enteric infections [23]. We also found that having access to cleaning materials
381 such as paper in the toilet and a drop hole cover on the toilet were both negatively associated

382 with ESBL-*E. coli* colonisation. Such infrastructure is used to prevent flies from accessing
383 faecal matter, thus this association is consistent with studies which have shown the role of
384 flies in transporting and transmitting *E. coli* [24,25].

385 Antibiotic use was identified as a risk factor for carriage of ESBL-producing *K.*
386 *pneumoniae*, which is consistent with previous studies in sub-Saharan Africa [5,8,9,10]. The
387 highest level of antibiotic use reported in time was at the baseline visit, with lower rates
388 reported at subsequent visits, likely due to shorter intervals between subsequent visits
389 compared to the initial six month question. The reason for the regional variation is uncertain.
390 Chikwawa being the rural area of the study [14], participants may have encountered more
391 often organisations such as non-governmental organisations that might have been able to
392 offer them treatment or antibiotics, or participants might have had increased access to
393 antibiotics due to the greater presence of animal farming [26]. This emphasizes the
394 importance of antimicrobial exposure in driving ESBL colonisation, thus highlighting the
395 need for a more responsible antibiotic consumption.

396 Given our model and dataset, eating from shared plates rather than from separate
397 plates as well as the presence of standing water around the household appeared to have a
398 protective effect. This is a surprising result, since plate-sharing, which is common in LMICs
399 [27] and has been associated with other enteric pathogens in other settings [28], would seem
400 to promote transmission between individuals. Similarly, since wastewater is known to play a
401 role in the transmission of AMR [12], the presence of standing water around the household
402 would also seem to promote transmission. We caution that these results might represent a
403 Type I error (in a Bayesian context), and that further research into WASH behavioural
404 patterns and interactions with other explanatory variables would be necessary to confirm or
405 refute our findings.

406 For ESBL-producing *K. pneumoniae*, at the univariable level, household size was the
407 only highly significant risk factor, highlighting the importance of the household in driving
408 ESBL transmission. Other variables identified as conferring a significant increased risk of
409 being colonised with ESBL-producing *K. pneumoniae* included owning birds, which are
410 known to be responsible for faecal contamination of the household environment in LMICs
411 [29], and entering into contact with drains, highlighting again the importance of interactions
412 between animals, humans and the environment.

413 The temporal models for both bacterial species detected a temporal correlation range
414 of seven to ten weeks. In other words, two samples taken within that time frame are more
415 likely to both be colonised than if spread apart in time any further. Though our method is
416 designed to only detect association between ESBL prevalence in subsequent follow-ups, this
417 does suggest that within-household transmission occurs within this time frame. Subsequent
418 causal inference studies would, however, be required to confirm this.

419 This study had some limitations. The volume of information available from various
420 questionnaires was considerable and for that reason, we had to pre-select variables based on
421 their perceived importance by environmental health experts. Although we found temporal
422 correlation at household-level, we could not find any at individual-level, which suggested that
423 an individual's samples could be seen as independent from each other. Potential explanations
424 for this lack of temporal correlation at the individual level include the use of stool samples
425 over rectal swabs, which may have been better for screening. Additionally, the laboratory
426 protocol for testing was qualitative, discriminating only between presence or absence of
427 ESBLs without quantification. Further work is needed to consider the impact of
428 microbiological methods on informing these models (i.e. time in enrichment broth and/or
429 quantification by minimal probable number estimates). Whole genome sequencing will allow
430 for a more precise investigation of the samples to get a better understanding of the linkage

431 between sequence types. The COVID-19 pandemic also played a role in derailing the
432 microbiological sampling for our study and potentially impacting our results. The pandemic
433 caused the sampling and microbiological testing to be suspended between April and July
434 2020, which caused some delay in our data collection.

435 Our study suggests that WASH factors and environmental hygiene are key drivers of
436 AMR-transmission in Malawi, consistent with findings in other African settings [30]. Our
437 results also point towards acquisition of ESBL-producing *E. coli* through contaminated water
438 and/or inappropriate WASH infrastructure. Additionally, seasonality and gender also suggest
439 the importance of environmental hygiene and practices in driving ESBL-producing *E. coli*
440 transmission. This underlines the need for improved access to clean water and suggests that
441 associating WASH behavioural practice with better WASH conditions would be instrumental
442 in decreasing transmission. However, for ESBL-producing *K. pneumoniae*, previous
443 antibiotic use was identified as a risk factor, therefore emphasizing the importance of
444 antimicrobial exposure in driving ESBL-producing *K. pneumoniae* transmission and the need
445 for improved infection prevention and control (IPC) measures and antibiotic usage and
446 stewardship training. A better understanding of how the WASH conditions of the different
447 communities impacts ESBL colonisation and transmission will inform public health
448 responses to the challenge presented by AMR and enable design of effective intervention
449 strategies in Southern and Eastern Africa.

450

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459

460 **Data availability statement**

461 The authors confirm that the data supporting the findings of this study are available within its
462 supplementary materials.

463

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587

588 **Supplementary information**

589 **S1 Appendix. Modelling framework.**

590 **S1 Dataset. Dataset used for the modelling.**

591 **S1 Table. Covariates and outcome variables.**

592 **S2 Table. Univariable analysis results between ESBL-producing *E. coli* colonisation
593 status and each variable accounting for the study area.**

594 **S3 Table. Estimates for ϕ , σ and τ in the ESBL-*E. coli* temporal model.**

595 **S4 Table. Univariable analysis results between ESBL-producing *K. pneumoniae***
596 **colonisation status and each variable.**

597 **S5 Table. Estimates for ϕ , σ and τ in the ESBL-*K. pneumoniae* temporal model.**

598 **S1 Fig. Prior and posterior density of ϕ , σ and τ (left to right, without warm-up) for**
599 **the ESBL *E. coli* temporal model.**

600 **S2 Fig. Trace plots of ϕ , σ and τ (left to right, without warm-up) for the temporal**
601 **model for ESBL *E. coli*.**

602 **S3 Fig. Prior and posterior density of ϕ , σ and τ (left to right, without warm-up) for**
603 **the ESBL *K. pneumoniae* temporal model.**

604 **S4 Fig. Trace plots of ϕ , σ and τ (left to right, without warm-up) for the temporal**
605 **model for ESBL *K. pneumoniae*.**

606







