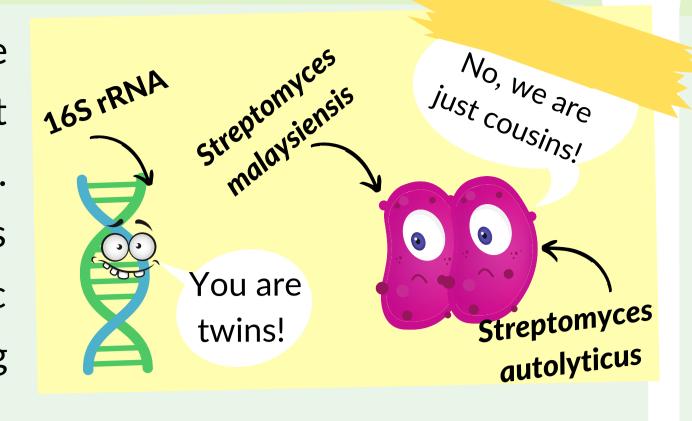
# Improved and Extended Multilocus Sequence Typing (MLST) Scheme for Streptomyces Reveals Complex Taxonomic Structure

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#### Introduction

Streptomyces species produce over 60% of all clinically-approved bioactive compounds[1]. Continuing discoveries of new natural products suggest that 165 rRNA Streptomyces genomes remain a promising source for novel antibiotics[2]. Comparative genomics and pangenomics are powerful tools for inferring genes involved in the synthesis of novel antibiotics from closely related genomic sequences. The contested nature of Streptomyces [3] taxonomy means that relying on existing assigned taxa may be misleading for pangenomics.



With the recent increase in available Streptomyces sequences we can now ask:

- How do STs map onto Streptomyces taxonomy determined from genome sequences?
- What does this tell us about the taxonomic structure of Streptomyces.

# MLST is used for genomic classification by comparing internal loci[4]. The current canonical Streptomyces MLST scheme provided by pubMLST comprises six markers (16S rRNA, atpD, gyrB, recA, rpoB and trpB) and 236 sequence types (STs; only two new STs were reported since 2016)[5].



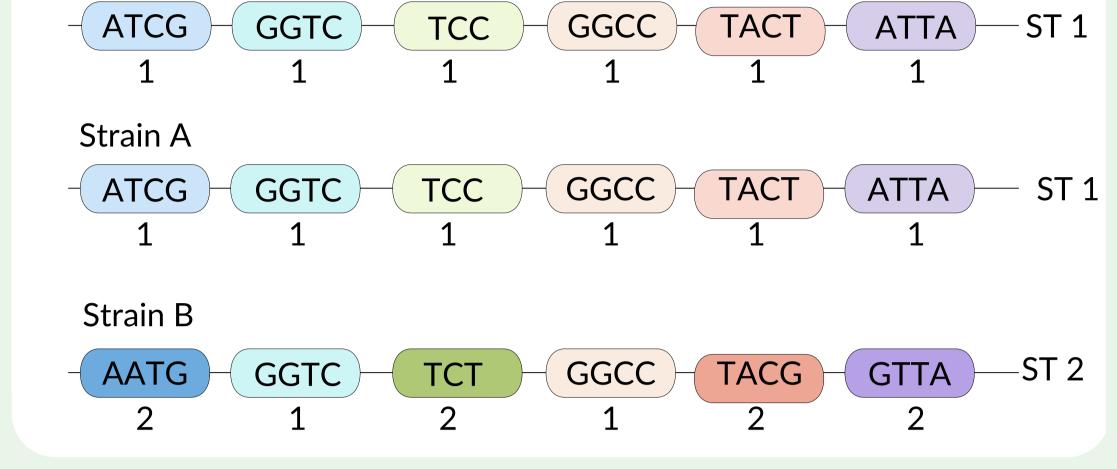


Figure 1. In MLST each marker variant sequence is assigned a unique number. For a single isolate, these numbers are combined to produce a profile, and each unique profile is assigned a ST.

**Known Sequence** 

#### Methods

Aim

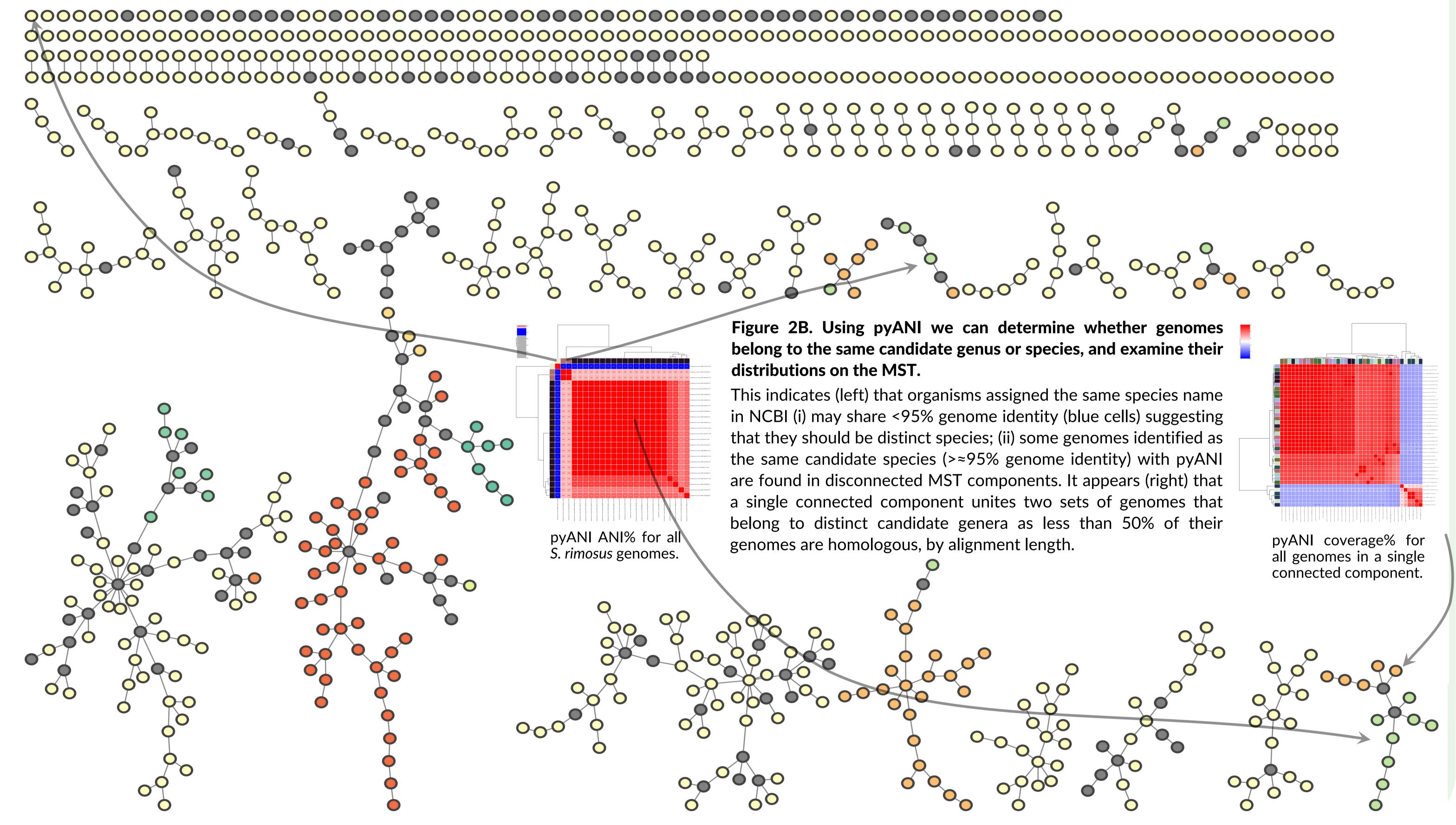
All 2276 available Streptomyces genome sequences were downloaded from NCBI[6] on the 8th July 2021.

673 16S rRNA, 813 atpD, 576 gyrB, 890 recA, 873 rpoB and 784 trpB new allele variants were identified with MLST tool[7].

Streptomyces taxon boundaries were assessed with pyANI [8] (%ID >≈95%, %coverage >≈50%).

### Results

Figure 2A. Minimum spanning tree (MST) with 852 STs and 292 connected components describing all sequenced Streptomyces genomes, and all STs from the pubMLST database. Each node represents a unique ST, and each edge corresponds to traversing from one ST to other by making up to five marker changes. This division of Streptomyces into 292 components that share no marker alleles with each other implies a set of natural divisions between groups of isolates. There are 150 pubMLST STs without any representative genome (grey nodes). Using pyANI it was determined that some connected components describe a single candidate genus (single node colours within a connected component), and some components represent more than one genus (multiple node colours within a connected component).



#### Figure 3: Sequencing a larger number of Streptomyces connected **components.** To investigate whether adding genomes/increasing the sequenced proportion of Streptomyces is likely to connect up the MST (figure 2) into a single connected component, we randomly sampled 10-90% of the available genomes and reconstructed the MST. The distribution of relative connected component sizes was independent of the number of genomes sampled, suggesting scale-free behaviour, that increased ~ 70% sequencing of Streptomyces will not result in a single connected MST, and that this reflects a natural division ~ 40% ~ 30% between groups of organisms. ~ 20% ~ 10%

Size of connected components

60

70

10

### Conclusions

- Multiple different candidate genera can be present in the same connected group of STs.
- Isolates that belong to the same candidate genus or species can be split across disconnected groups of STs.
- Currently assigned species names in NCBI do not reflect genomic difference and may incorrectly group organisms.
- The current set of MLST markers is unlikely to produce a fully-connected MST independent of number of sequenced genomes.





## References

[1] Procopio et al. (2012) Braz J Infect Dis doi:10.1016/j.bjid.2012.08.014 [2] Maiti et al. (2020) Scientific reports doi: 10.1038/s41598-020-66984-w [3] Labeda et al. (2012) Springer doi: 10.1007/s10482-011-9656-0 [4] Maiden et al. (1998) PNAS doi: 10.1073/pnas.95.6.3140 [5] Jolley et al. (2018) Wellcome Open Research doi: 10.12688/wellcomeopenres.14826.1 [6] Sayers et al. (2020) Nucleic Acids Res. doi: 10.1093/nar/gkaa892 [7] https://github.com/tseemann/mlst

[8] Pritchard et al. (2016) Analytical Methods doi:10.1039/c5ay02550h

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