

A systematic literature review of biocarriers: central elements for biofilm formation, organic and nutrients removal in sequencing batch biofilm reactor

Ahmad Hussaini Jagaba^{a, b, *, 1}, Shamsul Rahman Mohamed Kutty^{a, c}, Azmatullah Noor^{a, 1}, Abdullahi Haruna Birniwa^d, Augustine Chioma Affam^e, Ibrahim Mohammed Lawal^{b, f}, Mubarak Usman Kankia^a, Abdullahi Usman Kilaco^g

^aDepartment of Civil and Environmental Engineering, Universiti Teknologi PETRONAS, Bandar Seri Iskandar, Perak Darul Ridzuan, Malaysia.

^bDepartment of Civil Engineering, Abubakar Tafawa Balewa University, Bauchi, Nigeria.

^cCentre of Urban Resource Sustainability, Institute of Self-Sustainable Building, Universiti Teknologi PETRONAS, 32610, Seri Iskandar, Perak Darul Ridzuan, Malaysia.

^dDepartment of Chemistry, Sule Lamido University, PMB 048 Kafin-Hausa, Nigeria.

^eCivil Engineering Department, School of Engineering and Technology, University College of Technology Sarawak, Persiaran Brooke, 96000 Sibul, Sarawak, Malaysia

^fDepartment of Civil and Environmental Engineering, University of Strathclyde, Glasgow, UK.

^gCivil Engineering Department, University of Hafr Al-Batin, Hafr Al-Batin, Saudi Arabia.

* Corresponding author.

E-mail address: ahjagaba@gmail.com, ahmad_19001511@utp.edu.my, (A.H. Jagaba).

¹ These authors contributed equally to this work.

Abstract

Over the past decades, several materials utilized as biocarriers for immobilization of microorganisms have been gaining popularity showing different degrees of effectiveness. However, information concerning their effects on sequencing batch biofilm reactors (SBBR) performance is still lacking. There is currently no single widely acceptable material documented for proper biofilm formation, as most materials cannot achieve satisfactory level. Problems of biocarrier also exists due to the emergence of newer and more complex pollutants. Therefore, resource-efficient and environmentally friendly biocarriers are a call of the hour. This article thus, presents a systematic literature review of existing research articles on the various advances made between 2005-2021 about biocarrier physical properties, and performances with specific focus on their contributions in biofilm formation, nutrients and organic matter removal. This is to establish their role as central elements in biofilm formation. The review further described the operational challenges, mass transfer considerations, and recommendations made during successful utilization of the biocarriers in SBBR systems. Future research areas that may ultimately lead to large-scale commercial application of this biocarriers and will result in sustainable and environmentally friendly solution to the problems created during wastewater treatment have also been identified. This paper suggests that future researchers investigate the efficiency of composite biocarriers. It is believed that information contained in this review will increase readers fundamental knowledge, guide future researchers and be incorporated into future works on experimentally-based studies on biocarriers in SBBR systems.

Keywords: Biocarriers, Biofilm, Granular Activated Carbon (GAC), Sequencing Batch Reactor, Systematic Literature Review, Polyurethane Foam Cubes

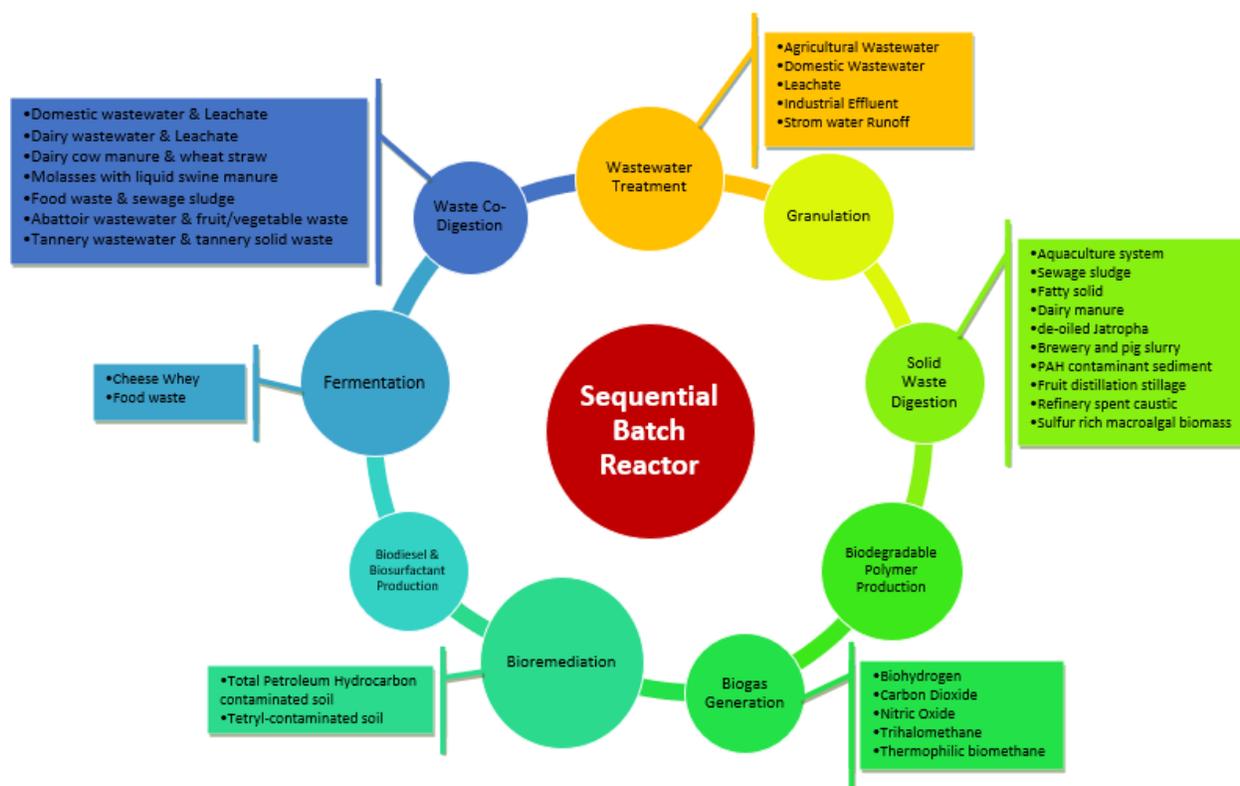
1.0 Introduction

Sequencing Batch Reactor (SBR) systems are non-steady-state, variable-capacity and suspended-growth biological wastewater treatment systems that uses the fill and draw activated sludge (AS) system with clarifier and an intermittent aeration mode where almost all metabolic reactions take place with the segregation of solid-liquid in a unit tank through a timed control sequence [1]. It blends both anaerobic and aerobic stages to successfully achieve nitrification, denitrification and phosphorous removal (SNDPR) concurrently [2, 3]. SBR is used to eliminate high strength organic and inorganic pollutants, nutrients and suspended solids (SS) in a single tank [4, 5]. Thus, it has several other applications as highlighted in Fig. 1.

As a biological treatment system, SBR system has the following benefits: (i) require small footprint, single basin operation with no secondary clarifier required, operated automatically with excellent process control possibilities (ii) Simple and flexible in configuration and operation, low installation and operation cost, high tolerance to various loading shocks with good bulking control (iii) Robustness, higher biomass retention, reduced energy consumption,

54 endurance to toxicity, solid retention time (SRT) decoupling from hydraulic retention time (HRT) and simultaneous
 55 organics, nitrogen and phosphorus removal [6, 7].

56 The use of AS-seeded SBR as a bioremediation technique could shed light on increasing the efficiency of
 57 treatment [8]. However, it has been hindered by the limitations of weak settling features, restricted scale-up and
 58 maintenance, broad footprint requirement, susceptibility to toxic environment and autoxidation, poor extraction
 59 efficiency because of biomass production restriction [9], unstable performance, excess sludge production amounting
 60 to high sludge volume index. Therefore, there is need to modify the SBR system and make it feasible for a broad range
 61 of applications by developing bioreactors with inert biomass immobilization support mediums called the sequencing
 62 Batch Biofilm Reactor (SBBR) [4].



63
 64 **Fig. 1.** Sequential batch reactor application for various treatments.
 65

66 SBBR is a hybrid SBR that suspends AS and connects growth processes into a single biocarrier-filled system
 67 where aerobic and anoxic conditions are given simultaneously in the biofilm's inner and outer layers. SBBR could be
 68 used for effective biological denitrification and dephosphatation as sufficient biomass can be sustained in the reactor
 69 and thus a higher and more stable treatment efficiency achieved, through the use of the biocarrier materials of various
 70 nature and types [1]. Finding an optimal operational strategy is a key point to achieve reactor's stability in SBBR.
 71 Available literatures have studied parameters such as; influent ammonium concentration, temperature, chemical
 72 oxygen demand (COD) load, running pattern, carbon to nitrogen ratio and many others with a conclusion that nutrient
 73 removal efficiency in SBR is always lower than in SBBR [10]. In SBBR systems, specific surface area (SSA) is large
 74 and the main types of microbial aggregates are biofilm and suspended sludge where biomass grows as a biofilm on
 75 small carriers that move freely into the wastewater [11]. The carrier packings serve as filter media to minimize the
 76 concentration of SS in the effluent. SBBR system needs no sedimentation tank [12]. An additional biomass inventory
 77 is been provided through biomass accumulation on biocarrier media in SBBR. Compared to an SBR system with
 78 similar SRT, high biomass concentrations can be attained by an SBBR system, resulting in lower HRT as reported by
 79 earlier studies.

80 Moreover, for the following reasons, SBBR can indeed cope with the heavy hydraulic load: (i) a large number
 81 of microorganisms could prosper because of the ideal microenvironment provided by the biocarrier, (ii) The growth
 82 of microbial attachment can enhance the degradation of TN, COD, TP and $\text{NH}_4^+\text{-N}$ (iii) non/intermittent aeration
 83 process could promote the activity and growth rate of microorganisms in the biofilm [13, 14].

84 The application of biocarriers for biofilm formation in SBBR system during wastewater treatment offers the
 85 advantages of higher nutrient removal efficiency, rich biodiversity, more compact design, improved sludge quality
 86 and biomass retention, energy savings, less footprint, flexible operation and management, higher volumetric loads,
 87 risen process resistance to shock loadings, less sludge and sludge conglomeration, good sedimentation, lower
 88 sensitivity to toxicity and reduced acclimatization time of the system [15]. More so, startup of the SBBR system could
 89 be 2–3 days quicker than the SBR system in attaining steady state [16]. The use of carrier materials to immobilize
 90 cells in sequencing batch reactors eliminates uncertainties about sludge granulation, improve solids retention, suppress
 91 settling step, reduce total cycle time and consequently increase contact for biological reaction [10]. Relatively, the
 92 system experiences fewer bulking problems, minimal clogging in the reactor, large surface area and do not require
 93 sludge recycle devices, while still sustaining great microbial assemblies inside the reactor [17, 18].

94 95 **2.0 Biocarriers and biofilm formation in SBBR**

96 Biofilm reactors are bioreactors that have embedded biocatalysts, either on or attached to the surface of an inert
 97 biocarrier, thus forming agglomerates [19]. Situations where there are high differences in hydraulic loading and where
 98 gently developing species with unique metabolic functions should be preserved from washout find biofilm reactors
 99 very helpful [20]. The bioreactors are capable of providing multiple sub-zones for different kinds of bacteria, thus
 100 preventing the slow-growing nitrifying bacteria from wash out in a fight with heterotrophic bacteria [18]. There have
 101 been studies of concurrent nitrification, denitrification and phosphorus removal in biofilm processes. Diffusion
 102 constraint in the biofilm, however, prevents the removal of biological phosphorus [21]. Irrespective of the kinetics
 103 applied in a biofilm system, hydraulic regime has a major effect on the overall process effectiveness of the procedure.
 104 Gigantic biomass concentration retention inside the reactor raises the resistance of the process and shock load
 105 resistance [22]. The two key parameters that define the efficiency of biofilm reactors are biofilm thickness and density.
 106 The thickness of the biofilm is regarded as the distance perpendicular to the biofilm surface substrata from the biofilm-
 107 bulk fluid interface. With the biofilm thickness, the biofilm roughness, which influences the fluid movement in the
 108 boundary layer, increases. More so, biofilm thickness influences both the useful diffusion coefficients of substrates in
 109 biofilms and the mass shift of substrates to the biofilm from the bulk fluid [23].

110 The diffusivity of oxygen in biofilms for wastewater treatment was studied by Hibiya [24]. The author set
 111 forth an empirical equation (See Eq. 1-3) describing the effective distribution coefficient of oxygen, D_e (m²/s), and
 112 the biofilm thickness,

$$113 \delta \text{ (m): } D_e = (3.5 \times 10^{-19} + 1.5 \times 10^{-21} \delta)^{0.5} \quad (1)$$

114
 115 According to the results, with increasing biofilm thickness, oxygen totally diffused toward the biofilms bottom layer
 116 and the depth of oxygen penetration increased. Substrates diffusion coefficients via biofilms are significantly
 117 influenced by biofilm density, which is the quantity of dry biomass in a given biofilm volume. With declining biofilm
 118 density, diffusion transport increases.

119 D_e depends on biofilm density, X (g/L), as follows:

$$120 D_e = 8.154 \times 10^{-10} \times 10^{-0.00753X} \quad (2)$$

121
 122 Thus, an individual column's biofilm density, X' (g/L), is computed using the following equation:

$$123 X' = \frac{TS}{\delta A_1} (g/TS) \text{ or } X' = \frac{VS}{\delta A_1} (g/VS) \quad (3)$$

124
 125 where VS (g) is the volatile solids, A_1 is the tested columns surface area and TS (g) as the total solids.

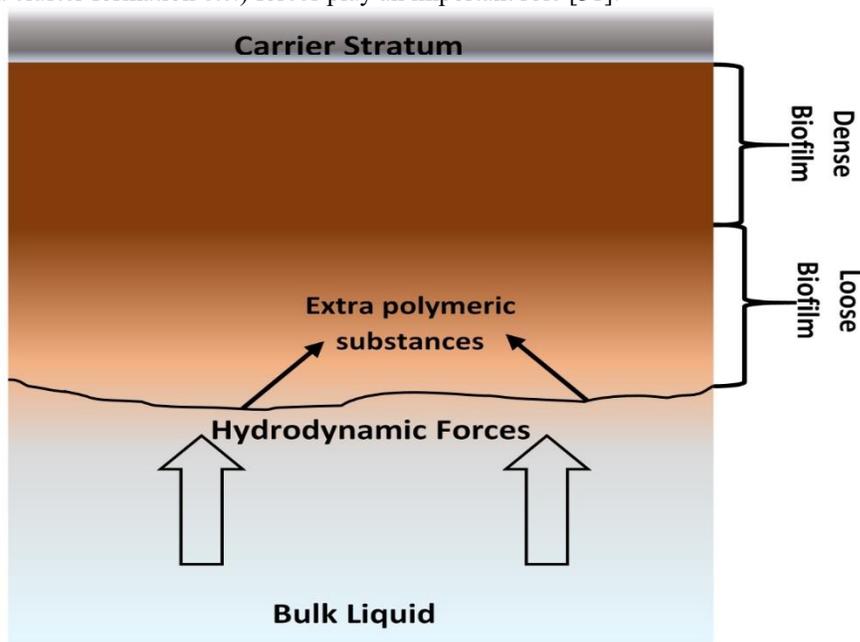
126
 127 The average rates of biofilm density for the three examined columns can be utilized to compute the entire module's
 128 X' (g/L) biofilm density [25].

129 130 **2.1 Biocarriers**

131 A progressively used biological treatment technique is the adhesion of microorganisms on surfaces of
 132 biocarriers [16]. The deciding constituent that fixes the maintenance of the potential microorganism and the reactor
 133 attachment phenomenon is packing. It is an essential factor for preserving SBBR stability and ensuring internal
 134 microbial biomass. [9]. Biocarriers are therefore built to provide the biofilm with a protective surface and optimum
 135 conditions for microorganisms breeding while they are suspended freely in wastewater [26]. In order to render surface
 136 area for bacterial appendage and growth, biocarriers are inserted into traditional AS reactors. To allow slowly-
 137 nitrifying growth and encourage nitrification, biocarriers may also provide greater SRT [27]. Different biocarriers are
 138 favorable to the enrichment of diverse and overflowing microorganisms, therefore, the choice of suitable biocarrier is

139 greatly essential to secure the effectiveness of the reactor. The various key factors to be considered during biocarrier
140 design are specific area, thickness of biofilm, biocarrier material (texture, form, etc.), and filling ratio. Research
141 suggests filling ratio to be less than 70% to enable biocarriers move freely [28]. The ability to monitor surface area
142 loading rates through the fill ratio of biocarrier within the bioreactor, is a major advantage of this process. It enables
143 high-rate treatment in a small footprint, making it a cost-effective alternative for treatment [29]. High surface area
144 biocarriers can produce more areas for the growth and absorption of microorganisms. Biocarrier forms and textures
145 have a similar influence on the weight, behavior and structure of the biofilm [30].

146 In SBBR, microbial attachment and biofilm formation on biocarriers are multiple-step processes. In these
147 processes (see Fig. 2), physicochemical (hydrodynamic, gravity, etc.) and biological (extracellular polymer
148 development, bacteria cluster formation etc.) forces play an important role [31].



149
150 **Fig. 2.** Multiple-step processes for microbial attachment and biofilm formation on biocarriers
151

152 Biocarriers can evenly swim within a moving bed reactor by the turbulent energy supplied by the aerator and
153 mixer [4]. The danger of biomass wash-out can be significantly reduced by immobilizing anammox bacteria as biofilm
154 on the surface of biocarrier. To increase the SSA required for bacterial growth, numerous biocarrier materials have
155 already been used. However, in choosing the most effective one for nutrient removal, DO transport from the bulk
156 solution to the biofilm interior remains the main issue. Daverey et al., [29] in their study also argued that only
157 anammox bacteria could be immobilized by the biocarriers used, while other microbial species (AOB-ammonia
158 oxidizing bacteria and denitrifiers) are also present in completely autotrophic nitrogen removal over nitrite (CANON)
159 and simultaneous nitrification and denitrification (SNAD) systems. This shows that the use of biocarriers involves the
160 biocompatibility of materials with the prevailing microorganism [19].
161

162 **2.2 Biofilm formation**

163 Biofilm is a tight microbial aggregate formed by means of immobilization of bacteria on the surface of biocarrier, as
164 suspended sludge freely disperses in solution (see Fig. 2). Biofilms are complex 3-D non-uniform bio-structures
165 comprising pores, channels, and irregular protuberance which can produce pleasing anoxic opportunities for
166 denitrifying bacteria [31]. They are created when unicellular organisms and associated deposits come together to form
167 a group that is encased in an exopolysaccharide matrix and attached to a solid surface.

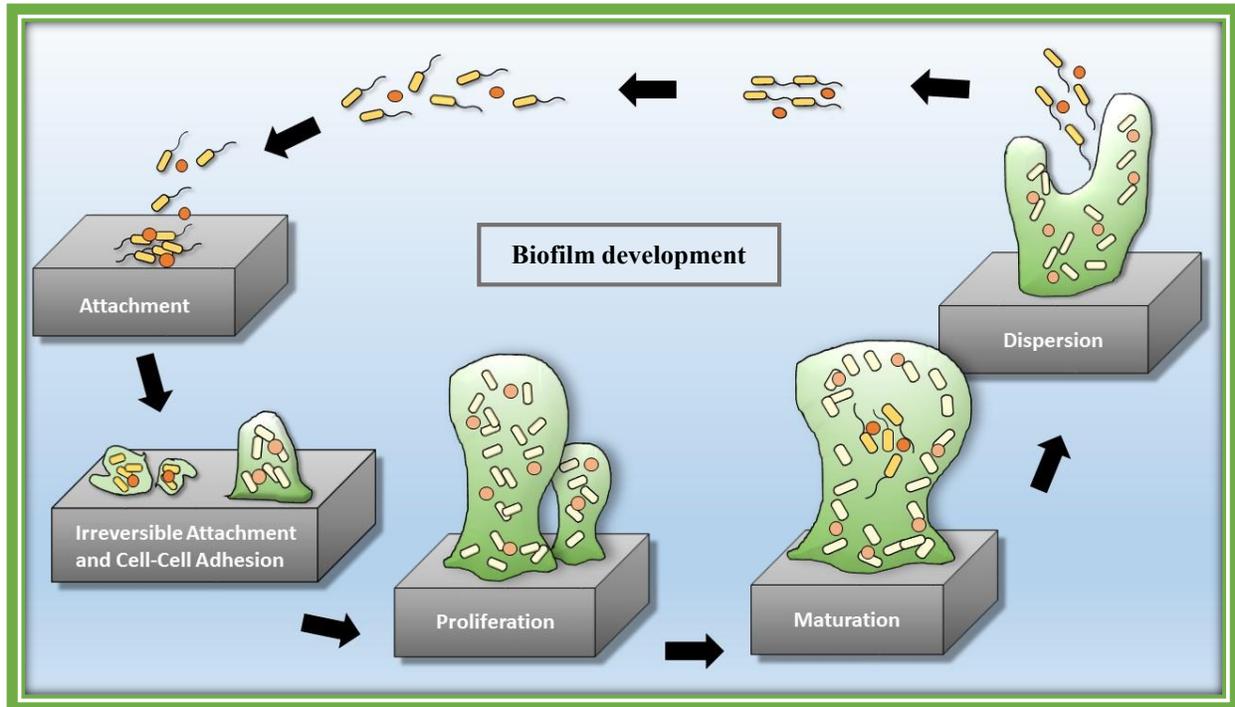


Fig. 3. Stages of biofilm development

168
169
170
171
172
173
174
175
176
177
178
179
180
181
182
183
184
185
186
187
188
189
190
191
192
193
194
195
196

Extracellular polymeric substances (EPS), otherwise referred to as slime, are substances of high molecular weight emitted by bacteria that facilitate bacterial adherence to the support media [26]. They are the main biofilm components that significantly connect cells together, and are recommended to increase nanoparticles resistance from AS and biofilm. Due to the rise in mass transfer resistance, their presence will decrease the diffusion rate of toxic elements into the deeper biofilm layer [4]. Thus, EPS can be described as an organic macromolecule emitted by microorganisms adhering to cell surfaces, enhancing bacterial aggregation and promoting the formation of stable structures in the microbial community to further grow biofilms. It is also a simple way to preserve functional bacteria especially in SBR systems.

Biofilm formation on biocarriers is the basis of a successful start-up that plays a significant role in biofilm reactor efficiency. The difference in growth conditions and substrate loadings can lead to differential biofilm growth in reactor tanks. In summary, biofilms development and growth exhibit four stages, namely: (i) initial attachment and young biofilm formation, (ii) biofilms accumulation, (iii) biofilm sloughing and updating, and (iv) biofilm maturation. The morphological structure of biofilm could cause aerobic, anoxic and anaerobic zones to occur simultaneously in the profile of the biofilm. However, variables such as antibiotics, heavy metals, salinity, and nutrient content may affect its role in nutrients and organic matters removal [32]. EPS is specifically correlated to microbial adhesion, aggregation and biofilm formation in SBBR systems. Therefore, analyzing EPS variations can help in understanding biofilm formation [33].

3.0 Research methodology

A systematic review of literature is an accountable, explicit and comprehensive method of study carried out through a consistent and specified process to promote article selection and the screening of relevant review articles. It is a kind of secondary study with the purpose of identifying, selecting, analyzing and interpreting all existing data related to a specific subject area, research question or interest phenomenon. The design of the systematic review reported in this article began in March, 2020. Fig. 4 (PRISMA) depicts the research method utilized in this study for systematic literature review. This could only be possible by identifying research questions relative to the research objectives.

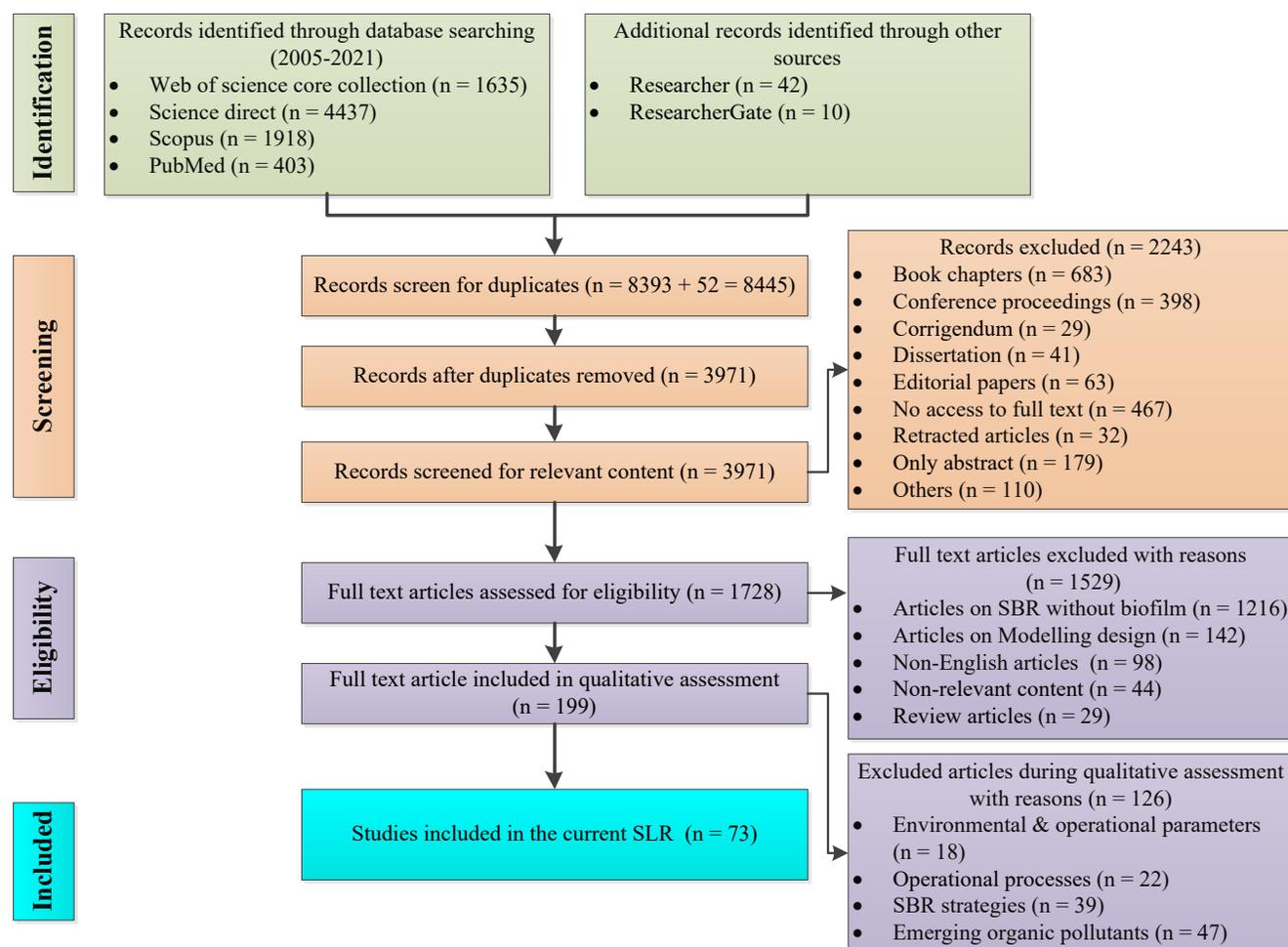


Fig. 4. PRISMA flow chart for the review process

3.1 Research questions

With the expanded utilization of SBBR for waste management resulting to gas generation and pollutant removal in wastewater, it has become necessary to study the function of biocarriers in the biological system. Therefore, the primary aim of this study is to form a perception of biocarrier materials performance in SBBR systems. The study further highlights how SBBR systems using different biocarrier materials become more significant than other biological treatment methods.

In a typical systematic literature review (SLR), the design of review process is often guided by research questions. Therefore, specifying the questions and developing strategy to answer them is the most important part of any SLR. In this present SLR, the main objective was to answer the questions regarding the performance of biocarrier materials in SBBR systems. Hence, the specific research objectives and the corresponding questions developed for this study as stated in Table 1 below are:

Table 1. SLR objectives and questions

	Objectives	Questions
1	To identify the most common biocarriers applicable in SBBR system and their properties	What are the distinguishing properties between the most common biocarriers used in SBBR?
2	To verify the performance of biocarriers in SBBR during nutrients removal and biofilm formation	How does biocarriers and their properties affect SBBR performance during nutrients removal and biofilm formation?

213
214
215
216
217
218
219
220
221
222
223
224
225
226
227
228
229
230
231
232
233
234
235
236
237
238
239
240
241
242
243
244
245
246
247
248
249
250
251
252
253
254
255
256
257

3.2 Search strategy and criteria

The search process adopted in this review focused on four (4) electronic databases (see Fig. 4) to obtain publications related to the review scope. The databases were selected due to the fact that they are among the most inclusive and exhaustive scientific databases with wide data coverage, encouraging the production process of sound bibliometric investigations. Additional records were found using other sources as “Researcher” and “ResearchGate”.

Pertaining to search criteria, it is worthy to note that, the coverage of this review was from 2005 to September 2020, including the following studies, given that several materials are still being investigated for potential application as biocarriers (e.g waste materials). The collection of data was completed on the 1st September, 2020.

To obtain biocarrier investigations over the scientific community, the next stage of the literature search was to insert two phases in the document search: TITLE-ABS-KEY “Sequencing batch reactor” AND “Biofilm”. The word “biocarrier” was not used because most authors do not use the word in their article titles. Instead, they use “biofilm” since the formation of biofilm is their major target. Finally, in order to obtain specific articles across the scientific community that focused on biofilm formation, which is practically impossible in an SBR system without a biocarrier, authors agreed and adopted the search by “Tittle”. Remarkably, we did not find any SLR article that investigated the impacts of different biocarriers in SBR for wastewater treatment.

3.3 Screening

The screening stage in this study began with duplicates removal. Several duplicates were found because four (4) electronic databases were considered plus other additional two (2) source. Records left after duplicates removal were further screened for relevant content. At this point, the following categories were removed: book chapters, conference proceedings, corrigendum, dissertation, editorial papers, no access to full text, retracted articles, only abstract and others.

3.4 Inclusion and exclusion criteria

The final search was conducted to obtain the most important articles for utilization in the SLR described in this review. Clear contributions, valid research and data were the basis for article selection upon which the study was carried out. The inclusion and exclusion process were intended to select the appropriate studies, extract and synthesize the data needed [34]. In the initial stage, a large number of articles were collected from the chosen databases for analysis. The bias for the articles inclusion or exclusion threshold was carried according to Eq. (4) defined in terms of included articles relevance ratio (R) for a given year (y), the number of matched keywords (k), the total number of proposed keywords (n) and the number of initial papers in a given year (P) according to equation (5).

$$R^y = \frac{\sum_i^n k_i}{P^y} \tag{4}$$

Included and excluded articles used in this research were supported by equation (5) [35, 36].

$$f(k_i) = \begin{cases} \text{included,} & R^y < \frac{k_i}{P^y} \\ \text{excluded,} & \text{(otherwise)} \end{cases} \tag{5}$$

The inclusion guidelines for articles chosen after filtering for the full review due to their quality and well-known relevance are detailed in Fig. 4.

3.4 Taxonomy

Fig. 5 depicts the taxonomy built on the basis of the literature trends; it also represents and differentiates between the various subcategories in the main category in order to prevent overlaps . Within the restrictions of this review, the taxonomy highlights the biocarrier materials used in SBBR systems on the basis of their significance, which it considers to be the key concern in the field of biological wastewater treatment systems.

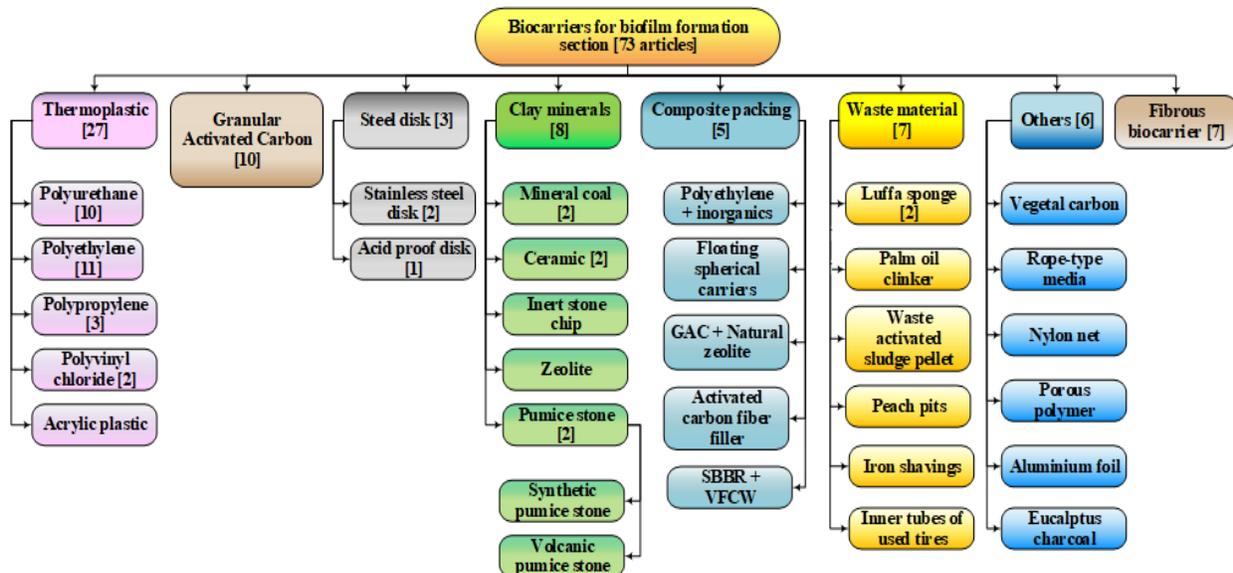


Fig. 5. Taxonomy of research literature on biocarrier materials

258
259
260
261
262
263
264
265
266
267
268
269

3.5 Descriptive data analysis

The scientific interest for the use of both natural and synthetic materials as biocarriers has over the year 2005-2020 maintained similar trend despite technological advancement. But, one thing that would interest you is that, during this period, a wide range of materials have been explored. This was possible as these materials are been subjected to characterization which displays the material property before and after use in SBRR. Fig 6 shows that studies on this specific topic in the year 2012 has eight (8) as the highest number of articles followed by years 2015 and 2019 that both have seven (7) articles. It could also be observed that year 2010 with a single article has the least number of articles.

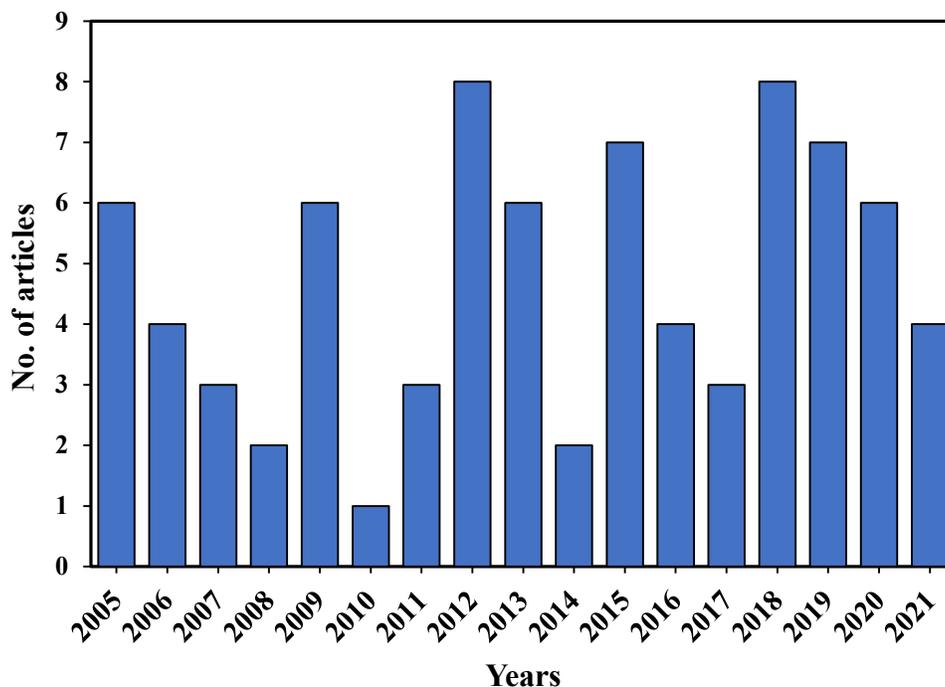


Fig. 6. Evolution of published article on biocarriers per year

270
 271
 272
 273 A total of forty-two (42) different sources of publications were considered in this review. This reveals the
 274 fact that learners and publishers are beginning to proffer significance to the application of different materials as
 275 biocarriers in SBBR systems due to their potential impact on biological treatments. Surprisingly, only five (5) sources
 276 published more than two articles which amounts to a total of twenty-six (26) articles. Of the five (5) sources, the most
 277 prolific journal with 10 papers is “Bioresource Technology” which main goal is the publishing research in biological
 278 wastewater treatment. The second highest is the “International Biodeterioration & Biodegradation” and “Journal of
 279 Environmental Management” with each having five (5) articles. It could also be observed from Fig. 7 that, the journals
 280 “Environmental Technology” and “Water Science & Technology” both had three (3) articles.

281 About Ten (10) journals covering 13.7% of the total seventy-three (73) articles considered in this review had
 282 two (2) articles each. The journals are: Advanced Materials Research, Desalination and Water Treatment, Ecological
 283 Engineering, Journal of Environmental Sciences, Journal of Ecological Engineering, Journal of Environmental
 284 Engineering, Journal of Hazardous Materials, Korean Journal of Chemical Engineering, Water Research, Water Air
 285 Soil Pollution.

286 A total of twenty-seven (27) journals each have one (1) as the least number of published articles on biocarriers
 287 that met the inclusion criteria of this review. They are: Applied Biochemistry and Biotechnology, Asian Journal of
 288 Chemistry, Biochemical Engineering Journal, Biotechnology and Bioengineering, Brazilian Archives of Biology and
 289 Technology, Brazilian Journal of Chemical Engineering, Asia-Pacific Journal of Chemical Engineering, Chemical
 290 Engineering Journal, Chemical Engineering Science, Chemosphere, Chinese Journal of Chemical Engineering,
 291 Desalination, Dyes and Pigments, Environment Protection Engineering, Environmental Progress & Sustainable
 292 Energy, Environmental Science & Technology, Frontiers of Environmental Science & Engineering, International
 293 Journal of Chemical Reactor Engineering, International Journal of Environmental Research, Journal of Bioscience
 294 and Bioengineering, Journal of Environmental Biology, Journal of Environmental Science and Health, Journal of
 295 Industrial and Engineering Chemistry, Journal of Water Process Engineering, Jurnal Teknologi, Process Biochemistry,
 296 Science of the Total Environment.

297 It is vital to note that, when assessing these sources, the majority have a very sectoral reach for engineering
 298 application of biological wastewater treatment. For this reason, journals with a strong environmental scope, as the
 299 “Journal of Environmental Management”, “Bioresource Technology” and “International Biodeterioration &
 300 Biodegradation” carved a widening niche in biocarrier studies.

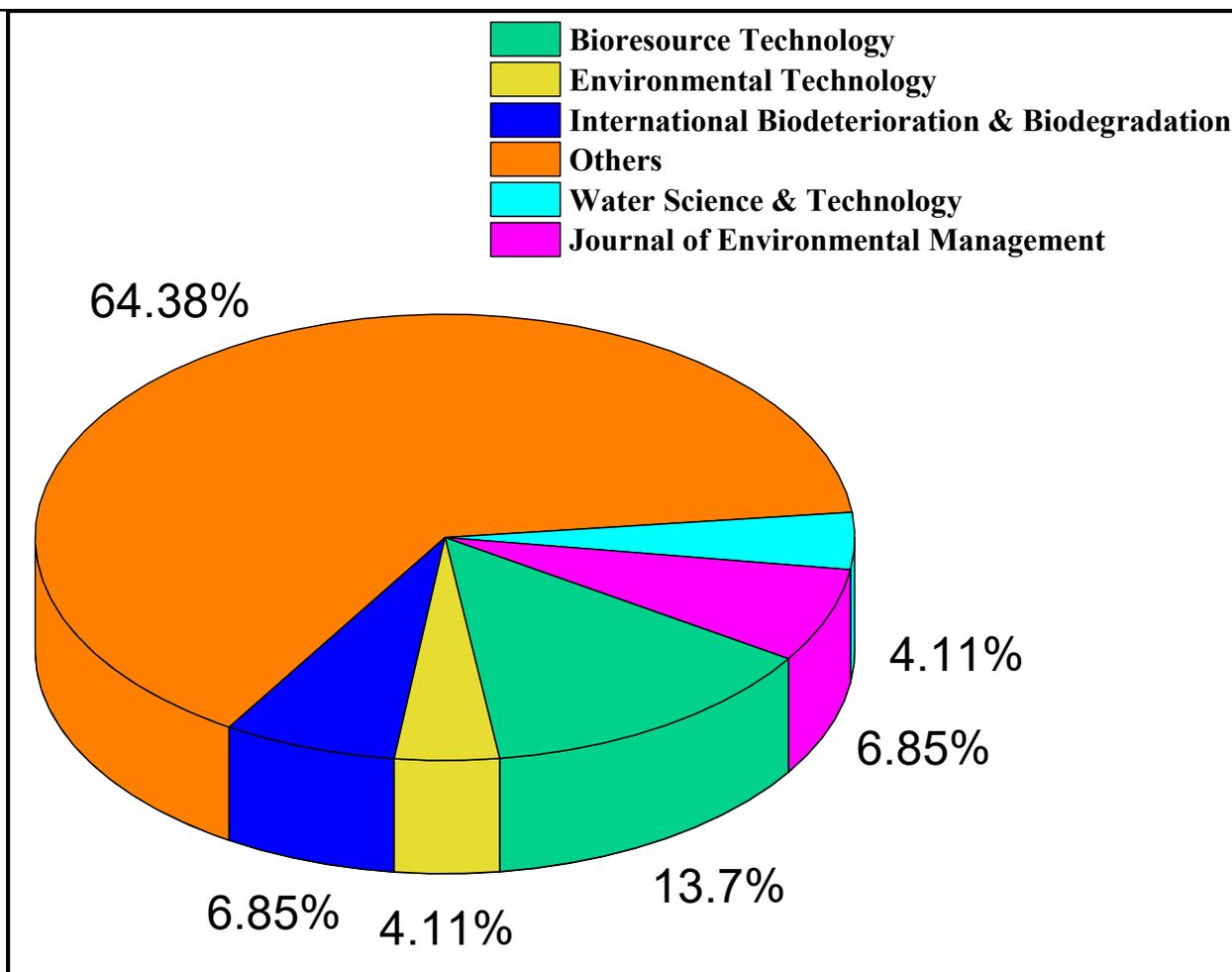


Fig. 7. Percentages of articles published per journal

301
302
303
304
305
306
307
308
309
310
311

Content analysis was utilized to know the geographical locus and sectoral focus of the articles. Although, both were not written in the keywords, the content analysis was conducted to make categorization of the articles in terms of location. It is interesting to observe that different countries are involved in biocarrier material study. Nineteen countries across the globe were found. China, Malaysia and Brazil with twenty-three (23), thirteen (13), and eight (8) articles respectively are the three major countries with the highest published articles considered in this study (see Fig. 8). India, Japan, Taiwan and Thailand published two (2) articles each, Poland Iran published four (4) and five (5) articles respectively. Least of all were countries that published only one article each. They are: Canada, Korea, Mexico, Portugal, Singapore, Spain and Turkey.

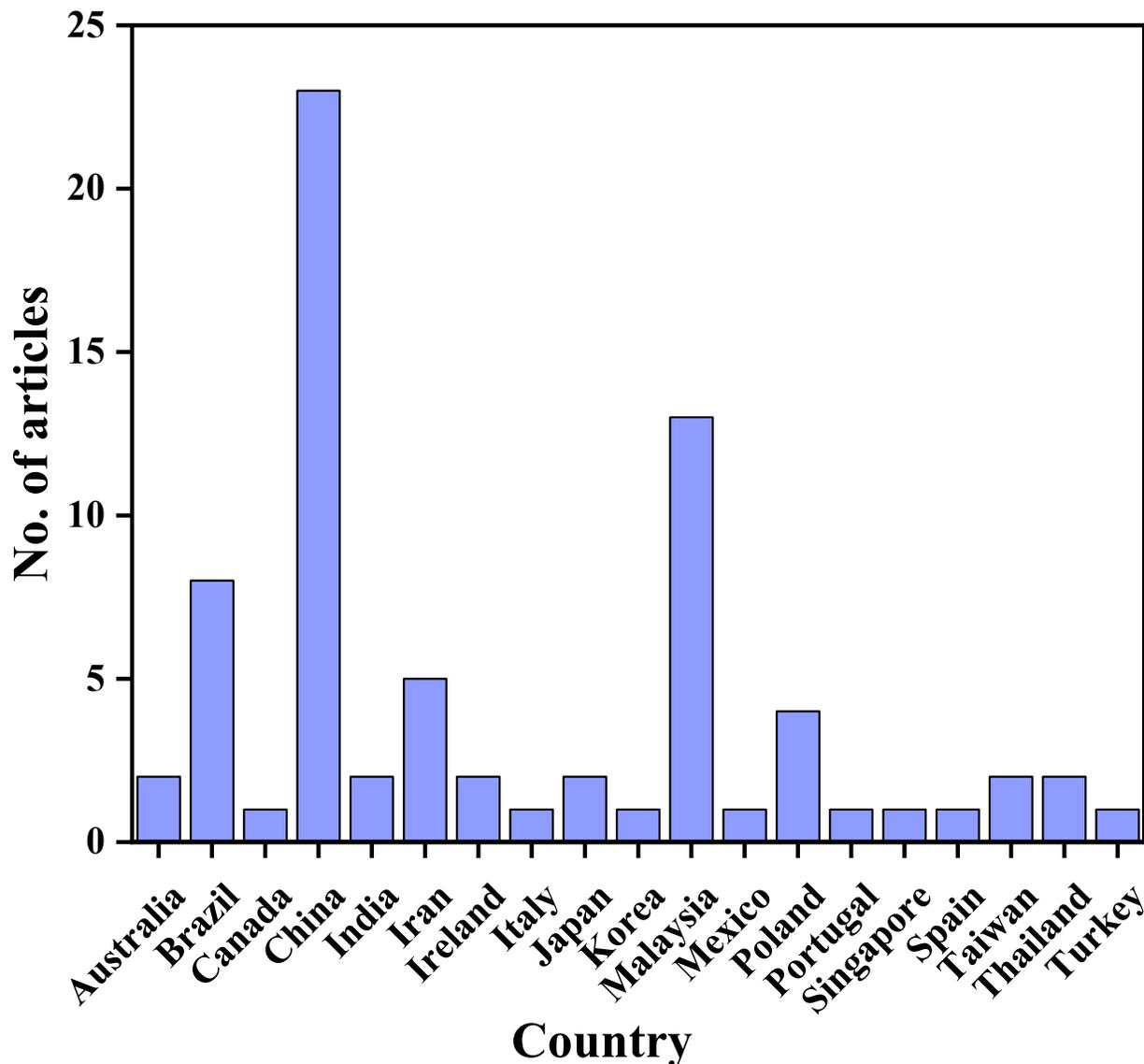


Fig. 8. Biocarrier research studies versus countries

312
313

4.0 Biocarrier properties

Biocarrier material selection is critical to wastewater treatment efficiency. The external and internal layers of the biocarrier are typically dominated by aerobic and anaerobic microbial communities. The total sum of the aerobic and anaerobic layers are used to measure the total biofilm thickness [19]. Several biocarrier elements such as polypropylene (PP), polystyrene, activated carbon, polyethylene (PE), polyurethane (PU) has been used to increase the SSA available for bacterial growth and yield different results. Therefore, it is very important to study and understand the properties of different materials before adopting them as carrier elements in a bioreactor.

The present section deals with the properties of several natural and synthetic biocarriers suitable to be used for biofilm formation in SBBR systems. For the sake of clarity, the comparison among properties of various biocarrier materials (thermoplastic materials, waste materials, minerals, steel disc, GAC) etc are reported and compared against each other in Table 2.

321
322
323
324
325
326
327
328
329

4.1 Thermoplastic materials

4.1.1 Polyurethane (PU)

330 Polyurethanes are formed by reacting a polyol (an alcohol with more than two reactive hydroxyl groups per molecule)
 331 with a diisocyanate or a polymeric isocyanate in the presence of suitable catalysts and additives. Because a variety of
 332 diisocyanates and a wide range of polyols can be used to produce PU, a broad spectrum of materials can be produced
 333 to meet the needs of specific applications. The different PU types existing in literature are: flexible polyurethane foam,
 334 rigid polyurethane foam, coatings, adhesives, sealants and elastomers, thermoplastic polyurethane, reaction injection
 335 molding, binders, waterborne polyurethane dispersions [37]. The cubic shape of thermoplastic PU foam is the most
 336 common type used in SBBR systems. The PU foam with an excellent setting for immobilization of biomass belongs
 337 to the support media type of Linpor. It is marked by great mechanical strength, high porosity, large-specific pore size,
 338 high absorption, corrosion resistant properties and low cost [38]. Combined anoxic, anaerobic and aerobic
 339 environment can be created by PU biocarriers. Biofilm protection from collision and fluid shearing by the formation
 340 of attached-growth biomass, carbon storage, and provision of sheltered anchoring points could be possible by the
 341 porous nature of the PU structure [39]. Apart from biofilm protection, other merits of PU biocarriers are: quick
 342 response to cycle time, short start-up period, changes in volatile solids loading rate and process stability. However,
 343 the porous nature of the biocarriers could lead to low mass transfer efficiency [40]. An SBBR stocked with PU
 344 biocarriers has been confirmed to be an adequate and inhibition-tolerant system for nitrification and denitrification
 345 process that may occur outer and inner layers respectively [41].

346 PU foam mostly described as a sponge media (see Fig. 9) can be available in different forms. These are:
 347 square, rectangular, circular, spherical, cubic etc. they are mostly described as sponge media. To immobilize biomass
 348 in a SBBR system, PU are used as support materials. They are mixed and compressed with a large quantity of sludge.
 349 After keeping in contact for at least 2 hours, the foam matrices could be transferred to a sieve, in which excess sludge
 350 are removed by washing the foam on the screen. The inoculated foam cubes are finally placed into the carrier holder
 351 material allocated inside the reactor encircling the agitator axle to appropriately capture the micro-organisms [42],
 352 until its complete filling. The PU common biocarrier holder materials include: glass baskets, perforated stainless steel
 353 basket (cylindrical), fish wire fixed into a plastic rack etc [33, 43]. In addition, colonization of PU foam cubes with
 354 anaerobic biomass appeared to be supported by the efficient physical retention of influent SS [44].

355 Surface morphology is very essential for PU foam characterization. Therefore, it can be analyzed by a
 356 scanning electron microscope (SEM). The outcome of the SEM does assist in assessing the PU foam cube 's total
 357 internal and external surface areas. PU foam pores are commonly considered to be spherical in appearance with a
 358 consistent diameter and hemispherical pores covering its external surface.

359 Using the assumptions that PU pores are spherical with hemispherical exposed pores. Each pore surface area
 360 (m^2) and volume (m^3) can be expressed by Eqs. (6) and (7), respectively:

361
 362
$$S_i = \pi d^2 \tag{6}$$

 363

364
$$V_i = \frac{\pi d^3}{6} \tag{7}$$

 365

366 where d (m) is the average diameter of the pore. The total surface area can be calculated by dividing Eq. (6) over (7)
 367 to obtain the surface to volume ratio. Rearranging:

368
$$S = \frac{6V}{d} \tag{8}$$

 369

369 where V (m^3) is the total pore volume of a cube of known weight and S (m^2) is the total surface area of a PU foam
 370 cube.

371 To obtain the value of V , foam cubes are being soaked in water to saturation followed by measuring the quantity of
 372 water infused into the pores. By dividing the overall surface area by the known weight of the PU foam cube, the total
 373 surface area per unit weight (m^2/kg) can be determined [39].

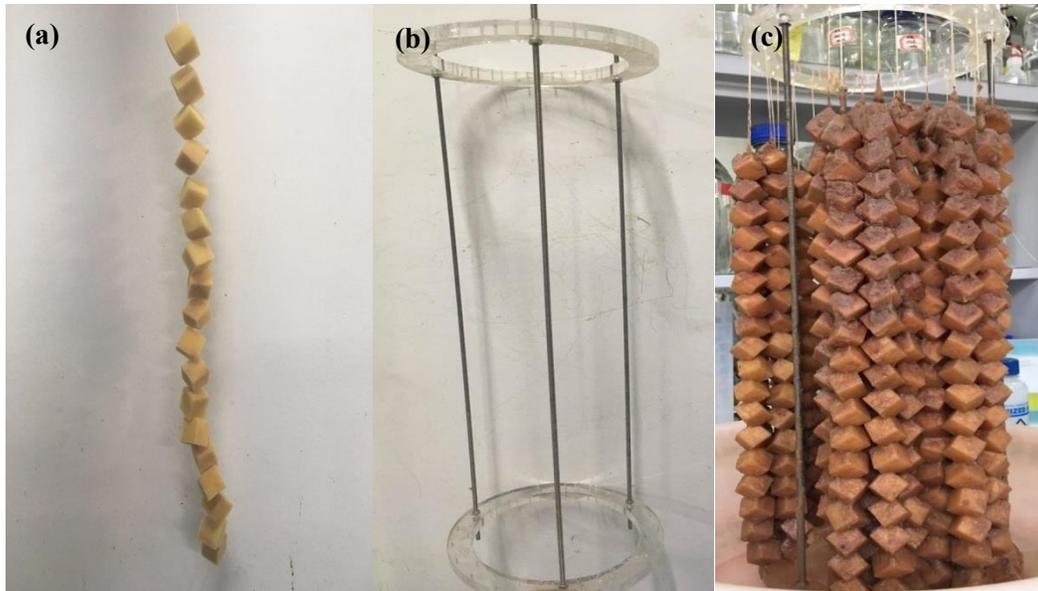


Fig. 9. Structure of immobile carrier used in the experiment: (a) carriers which strung together by using fish wire; (b) a plastic rack; (c) real photo of the immobile carriers (Adapted from [33]).

374
375
376
377

4.1.2 Polyethylene (PE)

378 PE is a lightweight, variable crystalline structure and most commonly manufactured and used durable thermoplastic
379 today. It is a man-made, homo-polymer, linear material mainly used for packaging [31]. It is also applicable to
380 laminates, films, biocarriers, tubes, plastic parts etc. The biocarrier elements made of PE can be shaped like small
381 cylinders with longitudinal fins on the surface and a cross in the inner layer of the cylinder [45]. Suitable and better
382 SSA for biofilm growth could be provided by the inner layer of the cylinder [18]. For these processes, the cumulative
383 SSA for biofilm growth could be provided by the inner layer of the cylinder [18]. For these processes, the cumulative
384 SSA is considerably greater than the efficient SSA. Therefore, the most significant parameter that defines the
385 performance of SBBR is the effective SSA [27]. The PE biocarrier density is less than water. This permits biocarriers
386 to circulate and suspend within the reactor [26]. In order to get more lighting and optical energy for bacterial growth
387 and enrichment, biocarriers can therefore rise to the surface during the non-aeration time [4]. Usually maintained in
388 suspension with an airflow rate, PE biocarriers act as biofilm formation and microbe adhesion vectors [46].

389 The numerous PE biocarriers found in literature are Kaldnes media, PE beads, commercial ring-lace kinds of
390 modified PE materials. The pumps and timing of operating cycles must be controlled by a time cycle controller device
391 using the PE beads fixed by a PE net. Kaldnes media as illustrated in Fig. 10 is one of the most commonly used
392 products as a biocarrier because of its simplicity in operation, high resistance, good total SSA and economic gain [17].
393 The Kaldnes media made of high-density PE (HDPE) exist as K_1 , K_2 , K_3 , K_4 , and K_5 [3, 47]. According to literature,
394 biofilm media microscopy from several full and pilot scale moving bed biofilm plants showed that there was no
395 evidence of biofilm germination on the surface of the smooth plastic elements using various Kaldnes PE media.
396 Erosion incurred by the regular collisions between the pieces could be assumed as the explanation for this.
397 Accordingly, internal surface of the plastic element can be used to compute the biofilm surface area [48, 49].
398

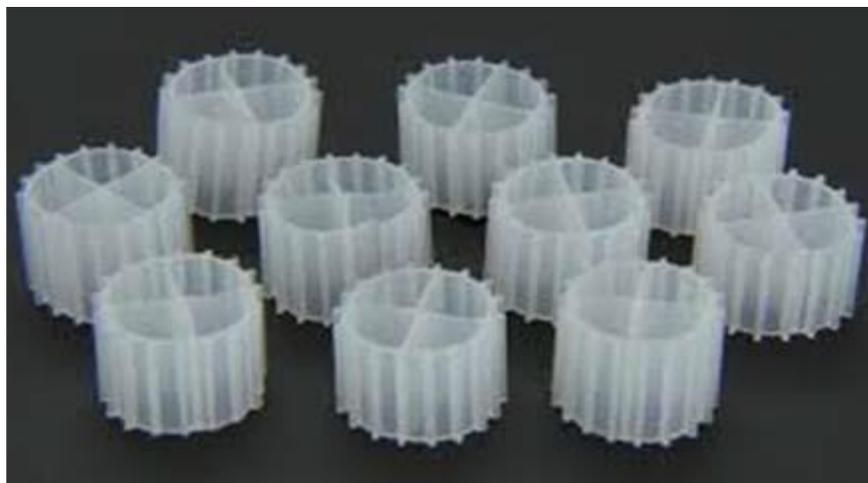
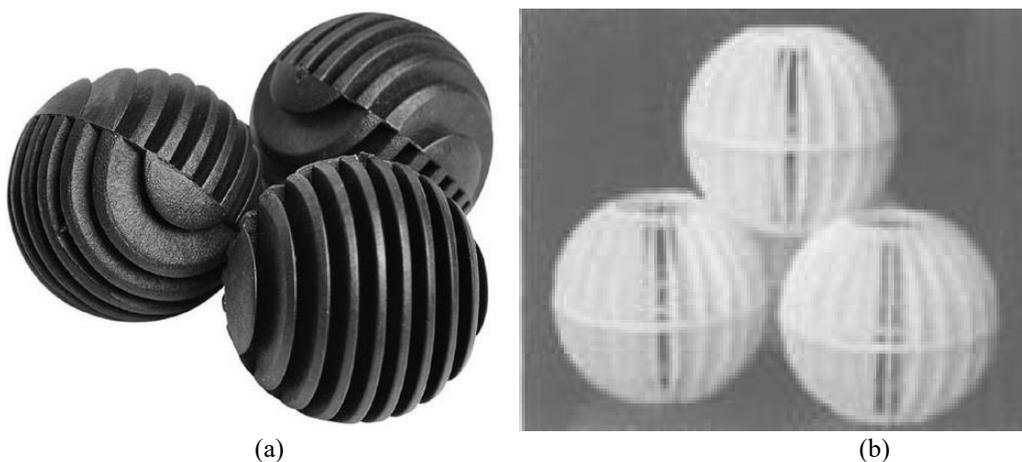


Fig. 10. Kaldnes biofilm carrier (Adapted from [17])

399
400
401
402
403
404
405
406
407
408
409
410
411
412

4.1.3 Polypropylene (PP)

PP is a thermoplastic polymer manufactured through chain-growth polymerization from the monomer propylene with wide variety of applications. It is partially crystalline and non-polar belonging to polyolefins groups. Multifaceted hollow balls, PP packing ring, membrane module containing hydrophobic PP dense hollow fibers, PP BioBall® carriers (see Fig 11a) etc have been used by researchers as carriers for bacterial immobilization [50]. BioBall® as a biocarrier made of PP has a spherical design is utilized by aquarists as mechanical and biological filter mediums [30]. The PP packing ring (see Fig 11b) are cylindrical in shape with a unique arrangement of three-dimensional hollow that could shield the biofilm from fluid shearing and create DO concentration gradients inside the biocarrier, presenting typical circumstances for SND. The PP rings are used to structure the PU foam, thus resulting to good bed porosity. It also has decussation for structural support at the center. Similar to some biocarrier materials, the shape of PP packing rings was regular, with large SSA. A study reported that packing rings shifted during aeration [21, 49].



413
414
415
416
417
418
419
420
421
422
423
424

Fig. 11. Polypropylene (a) BioBall® carriers and (b) white suspended packing rings (Adapted from [21] and [30])

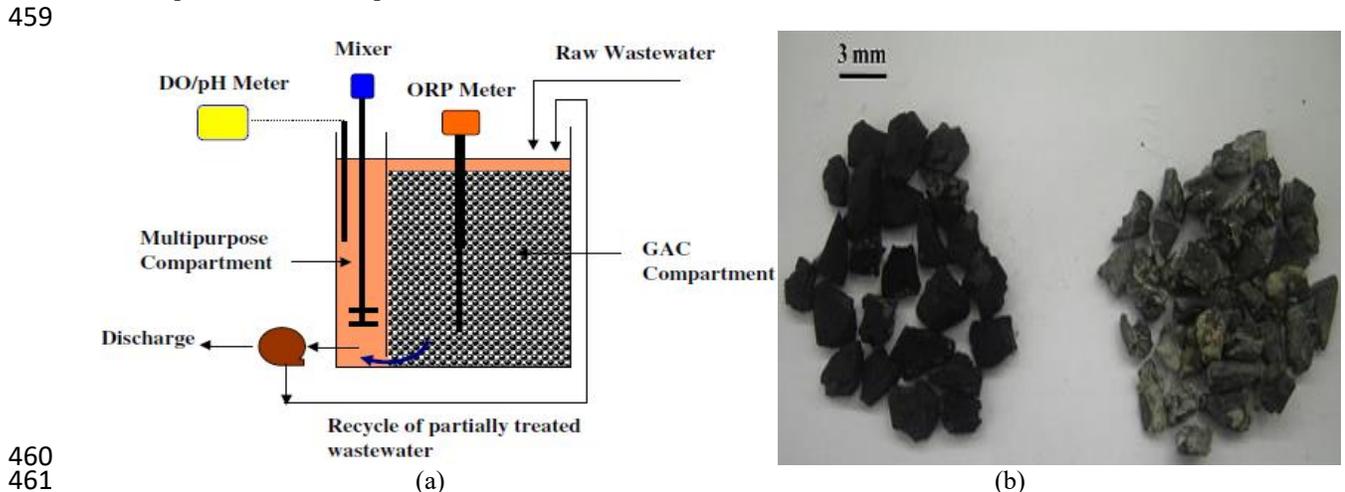
4.1.4 Polyvinyl Chloride (PVC)

PVC is the third most commonly manufactured and high-strength thermoplastic material on earth, which is essentially rigid and flexible in shape. Available in granular or powder form, it is a brittle, white, solid substance. It is being used in a range of applications in the health care, building and construction, automotive, electronics and other industries, in products ranging from blood bags and tubing, piping and siding to windshield system parts, biofilm formation, wire and cable insulation and more, because of its versatile properties such as: low cost, lightweight, simple processability and durability. To achieve an adequate surface area, PVC plastic modules can be assembled manually in a reactor with a stainless-steel frame attached to a pneumatic piston meter placed above the bottom of the reactor to accommodate

425 these modules. This is to provide space for the lower level switches to work freely and isolate the modules' bases from
426 any sludge build-up [51]. Pneumatic piston, timers, and limit switches are used to vertically move the modules in and
427 out of the wastewater in cycles [25].

428
429 **4.1.5 Porous polymers**
430 Porous polymers possess large SSA, high sphericity and structural strength, and a density similar to that of water. The
431 biocarriers are easy to fluidize but not easy to split, and thus ideal for the immobilization of microorganisms. Circular
432 fluidization of biocarriers in SBBR system has numerous benefits, including: (i) the breaking down of air bubbles that
433 improves the efficiency of mass transfer and enhancing interaction between biofilm and contaminants; (ii) reinforce
434 the collisions between biocarriers to renew biofilm immobilization on time and consequently foster biofilm activity;
435 (iii) decreased short circuiting, clogging, and excessive pressure loss occurrences that frequently occur in moving
436 biofilm or fixed bed SBBRs [52].

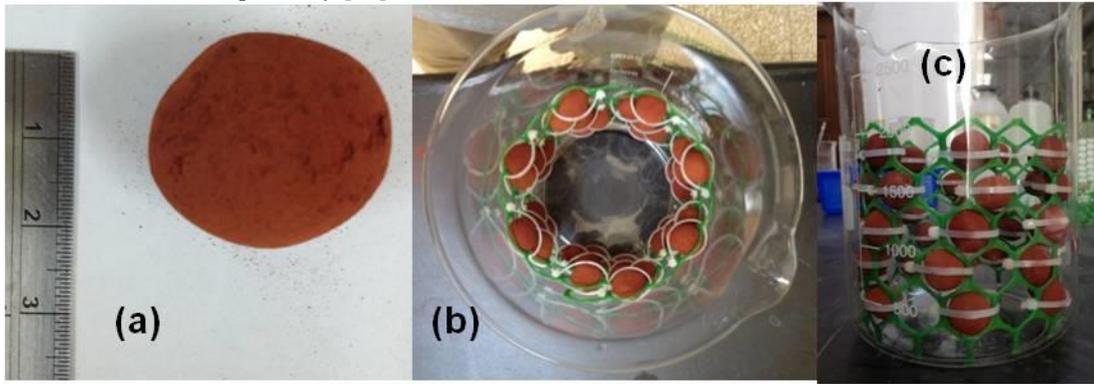
437
438 **4.2 Granular Activated Carbon (GAC)**
439 The filtration by GAC typically used in wastewater treatment was initially intended to eliminate organic matter by
440 adsorption. However, once the active sites are depleted, the GAC will no longer be useful. Therefore, the spent GAC
441 is either dumped or revived as secondary waste. GAC can be re-generated from apricot shells, coconut shells, coconut
442 husks, bamboo, walnut shells etc [53]. Most of these are commercially readily available [54]. Reports have it that, the
443 immobilization of microflora as biofilm on GAC particles (see Fig. 12a and 12b) results in high biomass holding,
444 allowing the process to significantly operate at higher liquid throughputs and OLR [55]. This is because of the
445 numerous advantages possessed by GAC that result in high process stability and increased substrate removal
446 efficiency: high surface area, porosity, large adsorption capacity, rough medium and continuous GAC bio-regeneration
447 [56, 57]. Since GAC's low specific gravity enables fluidization even at less upflow velocity, its capacity for sorption
448 shields the attached biofilm from shock loads [58, 59]. As an adsorptive biocarrier medium, GAC serves as a buffer
449 to decrease toxic chemicals level during process activity [60, 61]. The adsorption capacity of spent GAC may be
450 restored by chemical and thermal recovery on spent GAC to act as a support media for microorganisms' attachment
451 and growth. However, these methods have significant disadvantages, such as spent GAC disposal issues, high energy
452 consumption, carbon loss, loss of adsorption capacity and high cost. In addition, the spent GAC surface could be
453 regenerated for further adsorption by microorganisms that grow on it. Therefore, the use of spent GAC as a support
454 medium for biofilm formation could significantly contribute to minimizing waste generation and as well adding value
455 to the material in question [62]. The method is cheap, simple and ecofriendly. On the contrary, organic matter
456 saturation is an unavoidable limitation of GAC adsorption process. An undrained organic matter GAC is considered
457 poor since there is a short service life of around 6-12 months for GAC. Consequently, the biofilm can recover GAC
458 after a period of net adsorption to maximize its lifetime.



460
461
462 **Fig. 12.** (a) Schematic diagram of GAC-biofilm configured SBCR and (b) image of biofilm GAC acquired from
463 pilot-scale GAC-SBBR along with virgin GAC prior to its addition to the reactor (Adapted from [60] and [55]).

464
465 **4.3 Waste materials**
466 **4.3.1 Waste activated sludge (WAS)**

467 Several materials have been utilized as biocarriers for biofilm formation. Most of them were, however, solely used
468 to immobilize anammox bacteria, whereas AOB and denitrifiers are also present in the SNAD and CANON system
469 [58]. Therefore, new biocarriers that are highly effective in biomass retention within the reactor and are capable of
470 starting the SNAD process with long-term operational stability in a very short period are required. Biomass carriers
471 prepared by using WAS, chemical additive vesicant and red soil (see Fig. 13a, b and c) have been documented to be
472 appropriate. Red soil and chemical additive-vesicant added to the mix are responsible for providing stickiness and
473 hardness to the biocarrier respectively [29].



474
475 **Fig. 13.** Images of biomass carrier showing diameter (a), and arrangement in the reactor: top view (b), side view (c)
476 (Adapted from [29]).
477

478 4.3.2 Inner tube of tires

479 A biocarrier sourced from the inner tube of used tires in the form of cubes can be applied in an SBR system to increase
480 the generated bio-sludge quality and system efficiency. This is feasible as the material possess properties such as:
481 reusability without any regeneration, non-biodegradability and good sedimentation. Unlike biocarriers made from
482 thermoplastic materials, the density of carrier from used tires is greater than the density of water. This attributes to the
483 good settling characteristic of the material [63].
484

485 4.3.3 Palm Oil Clinker (POC)

486 POC is a by-product of the incineration process resulting from the exhaust stream of the palm oil industry plant in a
487 steam boiler producing electricity. It is a black solid waste material seen in Fig. 14, which causes environmental
488 sustainability problems. POC has an irregular shape, rough surface that contain huge number of voids and pores. Its
489 lightweight nature is attributed to the extremely porous nature of its interior portions. With a chemical mixture of
490 Al_2O_3 , Fe_2O_3 , and SiO_2 , POC is regarded a firm and non-biodegradable substance. Because of its easy operational
491 capability, affordability and the pace at which it improves the contact surface area for biomass aggregate, POC is
492 considered acceptable for biofilm formation in SBBR [64].



493
494 **Fig. 14.** Chunks of POC (Adapted from [64])
495

496 4.3.4 Peach pits

497 Thermoplastic materials have been widely studied for utilization in SBR serving as biocarriers for biofilm formation
498 during wastewater treatment. However, they are neither supportive to the community nor cost-effective. In terms of
499 environmental issues, the use of carbonaceous eco-friendly and wood-based biocarriers for proper immobilization of
500 bacterial consortia as biofilm may be a promising choice. Peach pits depicted in Fig. 15 could be used as sustainable
501 biocarriers due to their high porosity, firm structure, cheapness, and abundance. Pits must be removed from the reactor
502 and drained for 1 h at 105°C to compute the mass of biomass attached to the peach pit surface. The dried biocarriers
503 will be weighed to calculate their total mass (M_t) with biofilm. Afterwards, pits will be rinsed and sanitized to assess
504 the weight of biofilm-free (M_c) carriers [9]. Lastly, the volume of biofilm fastened over the surface of 20 peach pits
505 (BS_{20}) can be calculated according to the following equation:

$$BS_{20} = M_t - M_c \quad (9)$$

$$BS = BS_{20} \times \frac{585 L^{-1}}{20} \times FR \quad (10)$$

509
510 When the filling ratio (FR) is 100%, 585 carriers can flow into the unit volume of the reactor. The 585 used in equation
511 10 above is the total number of carriers required in unit volume of a reactor. It depends on the average volume of each
512 carrier to be used [9]. Consequently, employing Eq. 10, it is possible to measure the total quantity of biofilm in the
513 reactor (BS) [65].



514
515 **Fig. 15.** Typical peach pits

516 517 **4.3.5 Luffa Sponge (LS)**

518 The LS is usually organic and biodegradable, naturally soft, yet durable materials. It is an annual vine that is edible
519 when young, and widely used as a bath sponge, with large cylindrical fruits and tendrils. However, young fruits that
520 are < 7 inches long and green can be consumed as fresh cucumber or squash replacement. The fruits fibrous interiors
521 of the LS gourd plant (*Luffa aegyptiaca* Mill) shown in Fig. 16 are the portion used as biocarrier. As a natural resource,
522 it begets no harm or ill effects on microorganisms. LS facilitates denitrification by serving as a carbon source. In fact,
523 it is an adequate habitat for microbes and easy to enrich the biofilm of microorganisms [66]. According to literature,
524 LS is ideal for use in immobilization processes due to the fact that: (i) the natural structure of its micro-fiber offers
525 voids where metabolites and substances are exchanged (ii) it has a broad specific surface with less density than water
526 and is also rich with hydrophilic surface cellulose and lignin [67].



Fig. 16. Structure of luffa cylindrical sponge (Adapted from [67]).

527
528
529
530
531
532
533
534
535
536
537
538
539
540
541
542
543
544
545
546
547
548
549
550
551
552
553
554
555
556
557
558
559

4.3.6 Iron shavings

Iron shavings can be utilized as biocarrier materials. They are commonly produced from mechanical cutting of cast iron ingots with a reduced iron content and SSA than nano-iron particles or iron powder purchased commercially. Because of their comparatively inferior surface area-to-weight ratio, they are ideal for promoting iron-assisted autotrophic denitrification. Iron shavings can be purchased from a machinery factory [68].

4.4 Steel disk

A biocarrier consisting of a bundle of eight fully submerged stainless-steel discs, co-axially joined and rotated by an electric motor in a bioreactor shown in Fig. 17 is said to allow biomass preservation in the form of biofilm coating on the discs [69]. It also conducts wastewater stirring and periodical aeration, discharges the entire amount of wastewater and enables the sloughing of surplus biomass. The disks submersion rate is to 100 % [32, 70].

4.5 Clay minerals

4.5.1 Zeolite

Zeolites are aluminosilicate microporous minerals widely used for concentrated $\text{NH}_4^+\text{-N}$ wastewater treatment as industrial catalysts and adsorbents due to their outstanding selectivity for many cations (see Fig. 18). Due to its superior adsorption characteristic, molecule sieve feature and ion exchange, natural zeolite is one of the most desirable materials for biological and physical wastewater treatment. It has a good selective ability to adsorb $\text{NH}_4^+\text{-N}$ and has been shown to support both aerobic and anaerobic microorganisms [71]. In order to provide a healthy living environment for AOB development, it interestingly serves as an excellent biocarrier suitable for AOB adhesion and enrichment. $\text{NH}_4^+\text{-N}$ concentration in zeolite-filled SBBR can achieve a dynamic equilibrium of $\text{NH}_4^+\text{-N}$ through ion exchange, which may also provide sufficient free ammonia levels in the reactor to maintain nitrification. It is assumed that a suitable FA range could be given and excellent partial nitrification achieved by maintaining a desirable $\text{NH}_4^+\text{-N}$ concentration for Nitrite Oxidizing Bacteria (NOB) inhibition [72].

4.5.2 Inert stone chips

Using an inert stone in an SBR, fixed bed has to be placed up to certain height level with a reasonable void ratio after immobilization of anaerobic mixed consortia. These packing material can be used to encourage the germination of H_2 producing acidogenic mixed microflora [20].



Fig. 17. SBBR disc media (Adapted from [32, 69, 70]).

560
561
562



Fig. 18. Zeolite

563
564
565

566 **4.5.3 Mineral coal (MC)**

567 MC is a hard-combustible sedimentary rock of plant origin, created by natural processes with some quantity of mineral
568 impurities, that appears in the form of beds or layers among other sedimentary rock (see Fig. 19). It is a complex rock
569 consisting of many natural organic and inorganic components, found in underground beds with different
570 physicochemical, technological, and mechanical properties. It is often regarded as a natural composite. By removing
571 the need for an internal system to maintain the bed particles, the replacement of many biocarriers with MC as a denser
572 material will simplify the reactor design. MC is said to be cheaper than several other biocarriers [73].



573 **Fig. 19.** Schematic representation of a typical MC (Adapted from [73]).

574
575
576 **4.5.6 Ceramics**

577 Ceramics can be utilized as vectors for biofilm formation and microbe adhesion due to the porous nature of its surface
578 structure. Since its density is slightly greater than water, the biocarrier could fix well on the supporting sheet. In order
579 to obtain adequate nutrients for growth and reproduction during the aeration cycle, bacteria and algae on the biofilm
580 could have detailed interaction with wastewater. And during the time of non-aeration, the two phases could be
581 completely separated. As biocarriers remain on the supporting layer to obtain optical energy and more illumination
582 that is beneficial for algae growth, AS can easily settle down to the bottom of the reactor [74].

583
584 **4.6 Fibrous carriers**

585 The fibrous packing media shown in Fig. 20 can shield the biofilm and provide bacteria with an ideal environment to
586 flourish. It combines the features of semi-soft and soft biocarriers. The biocarrier offers a robust framework to protect
587 bacterial cultures from the impact of operating excursions [14]. Fiber threads are normally filled into holes and
588 connected to the reactor surface by various materials, such as iron wire, thereby creating a biofilm carrier [75]. In
589 order to shape a huge surface area, filaments are spread and flexible carbon fiber yarns swung under the aeration
590 condition with the water in the reactor so that the mass transfer in the biofilm could be enhanced [76]. In both the
591 aeration and reverse flow region, these biocarriers can be packaged to shield the biofilm from fluid shearing and clash
592 and to absorb enough microorganisms on the biocarrier. Fiber threads have high adhesion efficiency, large SSA, large
593 biomass and evenly dispersed filaments around the block that can efficiently cut gas and increase the oxygen utilization
594 rate, thus providing ideal conditions for SNAD [1, 10, 77]. The powerful adsorption and growth of organisms results
595 from the rough surfaces of the fiber threads. The fiber threads could also be used as filter media to reduce effluents
596 SS concentration, which subsequently decreases bioreactor volume and settling time. The utilization of the flexible
597 fiber as biocarrier offers a high surface area for greater microorganism attachment resulting to increased biomass
598 concentration [78].

599
600 **5.0 Influence of biocarrier on biofilm formation, nutrients and organic matter removal**

601 The literature study reveals in general that several materials: thermoplastic (PU, PE, PP, polyester, polyvinyl chloride
602 polyacrylonitrile), steel disk, minerals (coal, inert stone chips, ceramic), fibrous materials, GAC, waste materials
603 (WAS, inner tube of used tires, peach pits, iron shavings, palm oil clinker, luffa sponge) etc. are used in SBBRs to
604 enhance the systems performance. The differences in their performance as depicted in Table 4 would be thoroughly
605 discussed in this sub-section as it directly affects nutrients removal and biofilm formation.

606

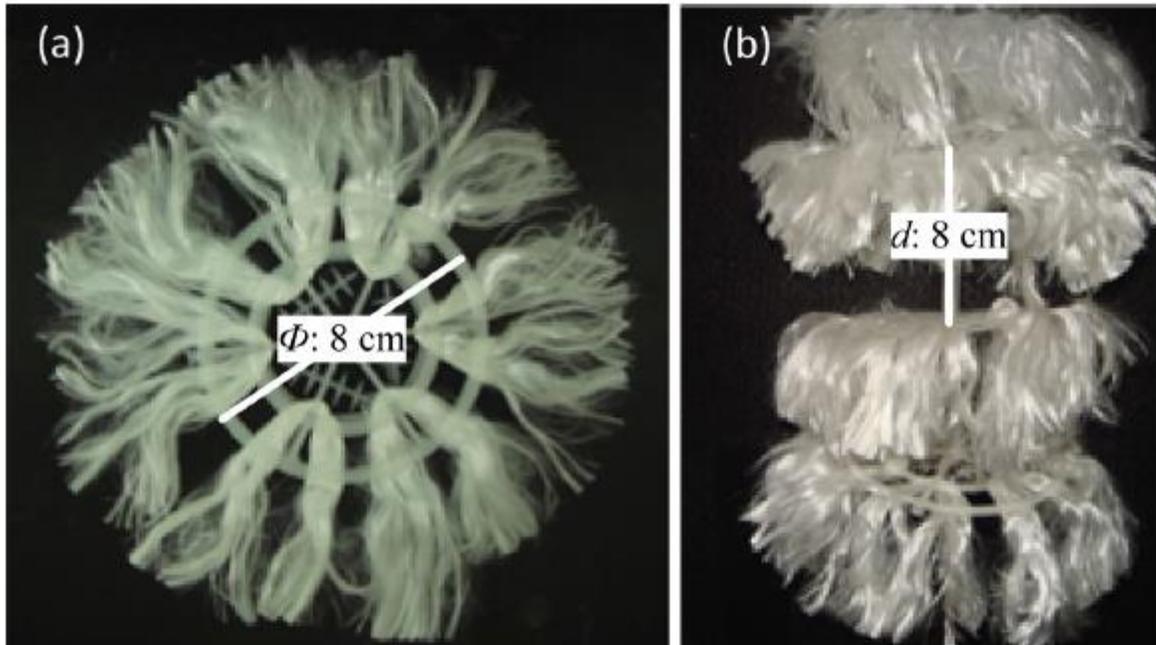


Fig. 20. Pictorial view of fibrous packing media used in SBBR (Adapted from [14]).

607
608
609
610

5.1 Biocarrier influence on nutrients and organic matter removal

5.1.1 Thermoplastic

5.1.1.1 Polyurethane (PU)

614 The improved performance usually experienced for SBBR systems is largely because of the appearance of attached-
615 growth biomass [43, 44, 79, 80]. The application of Linpor plastic PU foam was explored and achieved complete
616 nitrification even at higher organic loads because the average mean cellular retention times (MCRT) was 22.9 days
617 when used as a biocarrier by Valdivia et al., [81]. The greater MCRT value played significant role towards achieving
618 better nitrification by Linpor. Although denitrification was greater at lower organic loads, at higher organic loads $>$
619 $3.0 \text{ gCOD/m}^2 \cdot \text{d}$, better removal of COD and $\text{NH}_4^+ \text{-N}$ could be attained. With values ranging from 15-20 mg/L, Linpor
620 achieved the highest $\text{NO}_x \text{-N}$ yield. After the cycle operation, the nitrate concentration increased and ultimately
621 decreased.

622 According to Sarti et al., [38] in their work, the ultimate quantities of COD removal observed were the direct
623 product of methanogenic action. It may also be linked to the physical preservation of SS in the PU foam rather than
624 to the formation of a proper biofilm by microbial degradation. The carbon source and the presence of anoxic zone
625 were responsible for the promotion of SND processes alongside an improved TN removal in an MBSBR-PU biocarrier
626 [66]. Throughout the operational duration, almost complete $\text{NH}_4^+ \text{-N}$ removal with a medium effluent concentration of
627 0.5 mg/L was achieved. This suggests that acclimated PU foam cubes did not perform any significant role in the
628 process of assimilation [41]. The removal of $\text{NH}_4^+ \text{-N}$ may also be credited to the reality that the high pores of the PU
629 biocarriers have been trapped by many microorganisms. However, in a study by [37], the $\text{NH}_4^+ \text{-N}$ removal efficiency
630 has been reported to noticeably decrease as nitrogen loading rate (NLR) increases. Thus, this increase in NLR, NOB
631 inhibition coupled with decrease in DO concentration during the aerobic phase were responsible for Nitrite-N
632 accumulation.

633 In another study, it has been reported that for nearly 8 months, safe and highly effective nitrogen elimination
634 ($> 90 \%$) was attained. Consequently, Zhang et al., [33] stated that the application of porous sponge biocarriers could
635 provide a strong benefit for biomass immobilization and boost the retention within the reactor of anammox bacteria.

636
637

Table 2. Physical properties of biocarriers

Biocarriers	Size	Shape	Density	Surface area	Porosity (%)	Others	Reference
PU	20 x 20 x 20 mm		0.022 g/cm ³	>20,000 m ² /m ³	>97	Pore size (25 ppi)	[40]
	50 mm W	Cubes	23 kg/m ³				[38]
	2 x 2 x 2; 3 x 3 x 3; 4 x 4 x 4 (cm)	Cubes	87.4 ± 5.3 kg/m ³			Average pore diameter (661 ± 53 µm); pore count (42 ± 7 per 25 mm); total surface (177 ± 3 m ² /kg)	[39, 41]
	375 cm ³	Cubes	1 g/cm ³				[33]
	1 cm L		23 kg/m ³		95		[43]
			0.3 – 0.5 g/cm ³	900 m ² /m ³	97	tensile strength, 120 kPa	[66]
	Ø 0.6 cm	Cubes	0.023 g/cm ³	43.8 m ² /g	92	mean pore diameter, 543 µm	[79, 82]
PE	Ø 17 mm; 10 mm H	hollow, star-shaped		407 m ² /m ³	74		[46]
	Ø 8 mm, 16 L; 7 mm H		0.95 g/cm ³	690 m ² /m ³		Dp, 2 mm (Dp = equivalent diameter)	[47]
	Ø 9.1 mm; 7.2 mm L		0.96 g/cm ³	150 m ² /m ³			[17]
	Ø 9 mm; 7 mm H	Cylindrical	145 kg/ m ³	800 m ² /m ³		Bioflow 9	[26]
	1.5 x 1.5 x 1 cm		0.95 g/cm ³	415 m ² /m ³		Weight 0.43 g	[45]
	Ø 19 mm; 14±2 mm H			369.6 m ² /m ³	23	Weight, 87±3 N/m ³	[3, 48]
	Ø 25 mm; 12 mm H	cylindrical	110 kg/ m ³	500 m ² /m ³			[18, 27]
	Ø 25 mm; 10 mm H		0.95 g/cm ³	460 m ² /m ³			[31]
	Ø 10 mm; 7 mm H		0.17 g/cm ³	500 m ² /m ³	44	bed height, 0.17-0.33 m	[81]
	Ø 0.5 cm	cylindrical	0.40 g/cm ³				[82]
	Ø 17 mm; 10 mm H			407 m ² /m ³		<i>DupUM</i>	[49]
Ø 9.5 mm; 10 mm H			640 m ² /m ³		<i>BioloX10</i>	[49]	
PP	1000 mm L	membrane module and a cylindrical plastic shell				thickness of hollow fiber membrane (30–40 µm); intra membrane pressures (0.10MPa and 0.15MPa);	[50]
	Ø 19.0 mm	spherical	0.92 g/cm ³	378.0 m ² /m ³		BioBall®, weight, 1.7 g	[30]

			0.95 g/cm ³				[67]
	Ø 32 mm; 30 mm H			320 m ² /m ³		Bioflow30	[49]
PVC	200 x 200 x 180 mm	Vertical columns		290 m ² /m ³		Biodeks PVC sheets (BIOdek, UK)	[25]
	0.29 x 0.21 x 0.26 m	Vertical columns		290 m ² /m ³		Biodeks media (Munters, UK)	[51]
Acrylic plastic	Ø 5 cm; 1.25 cm H	Cylindrical	0.96 g/cm ³	0.03 m ²		Weight, 2.40 g	[8]
GAC	~1.5 -2 mm		1250 kg/m ³			residue on ignition (600 °C): 5%; loss of drying (120 °C): 10%); Feed rate: 40 g/L	[53, 58]
			560 g/L			GAC exhibits limited service life of ~ 6–12 months.	[54]
	Ø 1.30 mm; 0.25- 2.00 mm		1200 kg/m ³				[57]
	10-30 mm		0.85 g/cm ³	500-1000 m ² /g		specific gravity, 2 g/cm ³	[71]
Inner tube of used tires	2 x 2 x 2 mm ³	Cubes	1.925±0.21 g/cm ³	0.39±0.027 m ²		Volume of 8.0±0.74; weight, 0.0154±0.0022 g	[63]
Iron shavings				0.002634 m ² /g		Weight, 20 g;	[68]
Palm Oil Clinker	25- 35 mm		742-782 kg/m ³			Specific gravity: 1.64- 1.80	[64]
LS		Cylindrical	0.77-0.78 g/cm ³				[67]
		Cylindrical	0.018-0.05 g/cm ³	850 m ² /m ³	85-95	Tensile strength, 11.1 MPa	[66]
Peach Pit			1.134± 0.064 g/cm ³			Weight, 7.025 ± 0.342 g; volume, 6.2 ± 0.258 cm ³	[9]
Steel disk	Ø 0.10 m			25 m ² /m ³		thickness 1x10 ⁻³ m of each disk, distance btw disk, 20 mm/2 cm; rotation speed, 60 rpm	[32, 69, 70]
Zeolite	Ø 80 - 200 mm			412.67 m ² /g		Adsorption capacity of NH ₄ ⁺ -N, 10.41mg/L; Pore volume, 0.073-0.23 cm ³ /g; Pore size, 2.25- 3.82 nm	[72]
Inert stone chips	2.5 cm×1.5 cm				49		[20]

MC	Ø 40 – 80 mm				50	Weight, 500 kg	[73]
	Ø 20 – 40 mm				52	biocarrier occupied reactor volume 7.8 L	[19]
Ceramic	Ø 16 ± 3 mm	Spherical	1.1–1.2 g/cm ³	900 m ² /m ³	20.72–24.12		[74]
	Ø 4 – 8 mm	Sphere	1.4 g/cm ³	480 m ² /m ³	38	bed height, 0.16-0.33 m	[81]
Flexible fiber	Ø 0.07 mm; 75 mm L			2200 m ² /m ³	>90		[78, 83]
Fibrous slag	Ø 3 – 8 mm; 2-3 mm L			30 m ² /m ³			[84]
Plastic ball and GAC	Ø 3 cm, 2-3 mm		1111 kg/m ³			Plastic ball, total volume of 2.2 m ³ Volume occupied by the GAC 0.14 m ³	[55, 56]
Floating spherical carriers (PU with PE shell)	Ø 60 mm	Spherical PE shell	1.21-1.23 kg/m ³	2.15-2.21 m ² /g	85	water holding capacity, 452.38%,	[85]
Suspended carriers (carriers with a cross inside and longitudinal fins outside)	Ø 50 mm; 60 mm H	Spherical PE shell	0.96-0.98 kg/m ³	1.23-1.25 m ² /g		water holding capacity, 5.41%	[85]
WD-F10-4 BioM™ (composite of PE with inorganics)	Ø 10 mm; 10 mm L	Tube chip	0.96-0.98 kg/m ³	900 m ² /m ³		Thickness, 0.4 – 0.6 mm	[65]
Natural zeolite	10-30 mm		2.1 g/cm ³	500-1000 m ² /g	20-100	hardness, 3 N/mm ²	[71]
Rope-type media	220 mm L; 30-40 mm W				95		[86]
Nylon nets				4,140 m ² /m ³			[87]
Porous polymers	Ø 0.32 mm		1,320 kg/m ³	5,357 m ² /m ³		wet stacking density, 1,010 kg/m ³ , pore volume of 0.301 ml/g	[52]
Aluminum foil	Ø 10 mm; 10 mm H			500 m ² /m ³	95	specific gravity, 960 kg/m ³ ;	[88]
Vegetal carbon	Ø 0.5 cm	cube	0.51 g/cm ³		43	mean pore diameter, 1.9 µm	[82]
Eucalyptus charcoal	Ø 0.5 cm	Irregular pellet	0.51 g/mL	3.51 m ² /g	43	mean pore diameter, 1.9 µm	[79]

638 Note: L = Length; B = Breadth; H = Height; W = Width

639 It has been stated that seed sludge is primarily responsible for the improved efficiencies for the removal of COD and
640 nitrogen in MBSBR as it might easily penetrate and adsorb the porous layer of the PU type. Therefore, affecting the
641 production of biofilms in the form of a thin and cream-colored bio-membrane filled with microorganisms [40].

642 Bioparticle size exerts a decisive effect on the output of an anaerobic process under different operational
643 conditions. In to the findings by Pinho et al., [42], authors concluded that, total volatile solids (TVS) of the PU foam
644 biocarrier are not altered by changes in the bioparticle size, showing that the kinetic results were not affected by the
645 amount of biomass contained in the device. In a similar study, also varying bioparticle size, SND processes in the
646 MBSBR system resulted in increased TN removal with decreasing PU foam cube sizes [39]. The comparatively lower
647 TN removal attained when larger PU foam cubes were used could be due to the view that larger PU foam cubes were
648 not completely connected by biomass. This is in line with the classic theory as earlier stated by [42]. The elimination
649 of $\text{NO}_2\text{-N}$ concentration at $\approx 90\%$ was achieved using 8-mL PU foam cubes, followed by 80 and 70% for reactors with
650 27- and 64-mL PU foam cubes respectively. The fact that the existence of a greater volume of PU foam resulted in
651 the lower quantity of suspended-growth biomass involved in the nitrification process can explain the decreasing values
652 of $\text{NH}_4^+\text{-N}$ removal reported with an increase in biocarrier concentrations.

653 Comparing the performance of PU foam with some other biocarriers, it offered the quickest start-up and
654 produced steady effluent COD samples [82]. Considering the above-mentioned significant effects of PU foam in
655 nutrients removal, it is clear that $\text{NH}_4^+\text{-N}$ removal rate decreases with increase in NLR and biocarrier concentration.
656 The review results also revealed that the SBBRs with PU as biocarrier system has good capacity to withstand pollution
657 and hydraulic loading.

658

659 5.1.1.2 Polyethylene (PE)

660 PE media has been suggested as an efficient biocarrier for biodegradation of organic compounds [4, 49, 80,
661 82]. The concentration of COD and the period of settling may be the significant factors in denitrification [31, 45, 46].
662 The improved efficiency of the reactor biocarrier material in the elimination of aromatic compounds following
663 anaerobic decolorization was due to the lower COD concentration reported in the fixed-bed sequencing batch biofilm
664 reactor (FB-SBBR) effluent [16]. Using PE, biofilm biomass decreases as the C/N ratio decreases [26]. With
665 inadequate microbial denitrification activity at a low C/N ratio in the reactor, nitrate concentration remained high in
666 addition to suspended growth due to attached growth on biofilm carriers [17]. The findings showed that at a low C/N
667 ration, biofilm was a more suitable environment for nitrifying bacteria than activated sludge.

668 In an anoxic biofilm denitrification system, highly efficient nitrogen removal was achieved [27]. Due to the
669 endowment of denitrifying bacteria on biofilm, nitrite reduction was also observed. $\text{NO}_2\text{-N}$ concentration was found
670 to gradually decrease to 1.78 mg/L after one month, indicating that the capacity to lower $\text{NO}_3^-\text{-N}$ to $\text{NO}_2\text{-N}$ was
671 strengthened by biofilm growth and denitrifying microorganism enrichment on the biocarriers. In a Partial
672 Nitrification-SBBR (PN-SBBR) cycle, biofilm system was highly adaptable to an unstable influent substrate
673 concentration with high TN removal [18] and as the principal nitrogen species in the effluents after the creation of
674 biofilm, NOB activity was completely inhibited with nitrite. $\text{NH}_4^+\text{-N}$ concentration was rapidly reduced during stable
675 activity because of the dilution process following influent augmentation, and modified during the anoxic
676 denitrification step. Biosorption processes can be related to the decrease in $\text{NH}_4^+\text{-N}$ concentration.

677 Study summarized that ammonium removal efficacy of SBBR was more eminent than that of SBR as the
678 biocarrier made it easy to remove more ammonium from high strength ammonium wastewater. Small ammonium
679 molecules diffused within the formed nitrifying biofilm and a surface biochemical reaction occurred. Nonetheless, the
680 fixed biofilm enabled simultaneous conversion of accumulated nitrite to nitrogen. Fixed biocarriers provided more
681 mass transfer in comparison of moving biofilm carriers or suspended growth systems [3]. According to literature,
682 SND occurs in the depth of SBBR-PE biofilm carriers [47, 48]. The average MCRT for PE (Kaldnes) (10.0 days) was
683 far much less than achieved when PU foam (22.9 days) was used. For $\text{NH}_4^+\text{-N}$ and COD, thin biofilms (Kaldnes)
684 actually do better at lower organic load values (<2.5 gCOD/m²·d), with stable output at medium organic load values
685 [81].

686

687 5.1.1.3 Polypropylene (PP)

688 As with most thermoplastic biocarriers, PP biocarriers just couldn't get the biofilm prepared for COD removal in an
689 SBBR during the early days due to its smooth surface, which made it hard for microbes to bind, and in the meantime,
690 when the water was pumped in and out, the biofilm could easily be washed off [67]. The effluent COD with PP
691 biocarrier was about 39~88 mg/L. High-efficiency phosphorus removal in the built and run moving bed sequencing

692 batch biofilm reactor (MBSBBR) reactor [30] is likely to benefit from a combination of biofilm cultivation on
693 BioBall® carriers and suspended biomass in the reactor. Even though, phosphate accumulating organisms (PAOs) may
694 sometimes occupy the biofilm media. The evolution in the total quantity of attached-growth and suspended biomass
695 involved in the reactor will contribute to the fluctuation in the nitrification rate in the MBSBBR since nitrification
696 rates depends on oxygen penetration and the mixture of heterotrophs and nitrifiers inside the biofilm [21, 50].
697

698 **5.1.1.4 Polyvinyl Chloride (PVC)**

699 SBBR-PVC efficiency was examined by [51] in phosphorus and carbon removal. Sadly enough, clogging
700 took place in the plastic module when COD_T average removal rates were 8.3 g COD_T m²/d and 1.1 kg COD_T m³/d.
701 Analysis performed following the incidence of clogging did not show any substantial reduction in the unit's COD
702 removal capability.
703

704 **5.1.1.5 Acrylic plastic**

705 An acclimatization time that was 2-3 d shorter than that of a traditional SBR system with 5-7% higher COD
706 and BOD₅ removal efficiencies was obtained by the research team [8] using a sequencing batch reactor biofilm
707 (MSBR) system. This could be supported by the fact that, there was an increased quantity of biofilm mass on the
708 acrylic plastic media. Hence, the overall bio-sludge mass of the MSBR system was greater than that of the SBR
709 system.
710

711 **5.1.2 Granular Activated Carbon (GAC)**

712 Employing GAC with additional biomass plays an excellent role in nutrients and heavy metals removal as
713 adsorbent, filtration unit and biofilm attachment in wastewater treatment It demonstrated excellent efficiency in
714 performance and portrays a reliable process of treatment under stern organic load variations [57, 59]. GAC have
715 severally been reported to possess high adsorption capacity [61, 62].

716 To begin with, NH₃-N and COD demonstrated stable removal during the entire study duration by [54] using
717 an multimedia-sequencing batch biofilm reactor (MM-SBBR). High suspended solid and color removal efficiencies
718 were obtained using GAC with physical adsorption as the main colour removal mechanism [53, 60]. Other researchers
719 also claim that, after treatment with the pilot GAC-SBBRR, it was almost hard to foresee 2,4-dichlorophenol (2,4-
720 DCP) residues in the effluent [55, 56]. Results of the study by [58] was in the agreement with several other studies
721 showing high COD removal efficiency. Even at higher OLRs without process inhibition, the GAC-biofilm sequencing
722 batch reactor (SBGR) system retained its efficiency. The subsequent suspended growth system, however, led to
723 process failure. This can be due to the existence in the reactor of high composite chemical wastewater substrate
724 gradients that inhibit native biomass. Considering the above-mentioned significant performance, it can be concluded
725 that GAC biofilm has the potential to be further developed into low cost and environmentally friendlier process as no
726 chemical application is required with zero chemical sludge production.
727

728 **5.1.3 Waste materials**

729 The insertion of the inner tube of used tires could increase the moving biofilm aerobic sequencing batch
730 reactor (MB-aerobic-SBR) system's efficiency even at high organic loading. This has been seen as MB-aerobic-SBR
731 effluent consistency has been more stable at the same operating conditions than aerobic-SBR. According to [63], the
732 BOD₅, TP and TKN removal efficiencies of the MB-aerobic-SBR were 1–2, 10–20 and 2–3% higher than aerobic-
733 SBR, respectively. Given the porosity, hydrophilic and non-toxic characteristics of LS, it could provide great number
734 of microorganisms both inside and outside of it with habitat for their growth and reproduction, MBSBR-LS could
735 efficiently extract organic contaminants [67]. The COD and NH₃-N removal efficiencies are depicted in Table 2. The
736 TN removal efficiency could be enhanced by the denitrification process and subsequently enhance external carbon
737 availability in the biocarrier [66].

738 Iron shavings can be utilized as the principal substrate to support denitrification process in SBR operation.
739 However, it may result to very low denitrification rate. TN removal is closely related to the total iron concentration in
740 reactor. Effluent iron concentration recorded was as low as 2–4 mg/L and the depleted iron were mainly adsorbed
741 over the microorganism's surface, thereby affecting the microbial activity. An effective way to regain microbial
742 activity is the extraction of TSS by half and subsequently adding fresh mixed liquor suspended solid (MLSS) with
743 extremely low iron concentration [68].

744 POC provides low cost and simple operation. It is said to be more efficient as COD managed to achieve the
745 targeted effluent limit of ≈ 20 mg/L for the reactor with POC as a biocarrier. This proved that the application of POC

746 in SBR mode helps in improving the effluent quality in wastewater treatment [64]. Just like the inner tubes of used
747 tires, high OLR increases the biofilm total VSS ratio in a peach pit supported SBBR system [9]. Similarly, as OLR
748 and NLR were increased with decreased HRT in a WAS pellet biocarrier supported system, the reactor performance
749 gradually increased under with stable nutrient removals. These suggests that the biocarriers used are very efficient for
750 biomass retention [29].

751

752 **5.1.4 Steel disk**

753 When a steel disk was used in SBBR system, the improved performance in the absorption of organic substrate leads
754 to a high denitrification efficiency [70]. The thickness of the biofilm varies from 1-20 μm and allowed the organic
755 substrate to be ingested effectively.

756

757 **5.1.5 Clay minerals**

758 **5.1.5.1 Mineral coal (MC)**

759 In an SBBR flow cell experiments, the production of shortcut nitrification-denitrification to achieve efficient
760 nitrogen removal can be credited to the selection of MC as a biocarrier. This is dependent on the need for a lower
761 volume of oxygen and organic sources to support the procedure [19]. For the initial experimental periods, high VSS
762 and TSS removal efficiencies were reported. However, increased concentrations occurred in the subsequent periods,
763 possibly due to biomass detachments [73]. A progressive decline in the efficiency of COD removal was also noted
764 during the experiments.

765

766 **5.1.5.2 Ceramic**

767 The SBBR packed with Liapor ceramic spheres was at its best at less organic load [81]. The biocarrier
768 increased nitrate concentration, which gradually decreased at the end of the run, with effluent COD levels between
769 17-20 mg/L. The highest TN values for backwashing were achieved, suggesting that greater quantities of biofilm were
770 sloughed off during this process. In a related study, the addition of spherical ceramic biocarriers enhanced the removal
771 of TN and TP. The improved efficiency of nutrient removal was mainly because of biofilm bacteria and algae
772 development on the biocarriers. Moreover, the sludge discharge and biocarrier replacement were independent,
773 suggesting that algae and sludge retention time could be segregated. The harvesting of accumulated algae in biocarriers
774 will sustain the quantity and operation of algae biomass, which is good for the long-term stability of the A-SBBR
775 system [74].

776

777 **5.1.5.3 Inert stone chips**

778 At steady state condition, SBBR operated with inert stone chips showed high COD removal [20]. Biofilm floc
779 immobilization on stone chips with an anoxic micro-niche in the inward sections may be due to sulfate conversion in
780 the SBBR. According to the authors, at higher OLR, the reactor consistently showed good performance and stabilized
781 within a short period of time.

782

783 **5.1.5.4 Pumice stone**

784 In an FB-SBBR, volcanic pumice stones were used as biocarriers. the reactor achieved steady state condition
785 with a medium effluent COD concentration of 56.2 mg/L [16]. In a related study, synthetic pumice (SP) showed
786 chemical constituents loss with a COD effluent of 233 ± 52 mg/L during the reactor operational process [82].

787

788 **5.1.5.5 Zeolite**

789 Natural zeolite sequencing batch biofilm reactor (NSBBR) and synthetic zeolite sequencing batch biofilm reactor
790 (SSBBR) zeolite filled SBR were used to assess the impact of NH_4^+ -N adsorption ability on microbial characteristics
791 and nitrogen conversion. Nitrification and ammonium exchange occurred concurrently with significant nitrite
792 production rate differences observed in the two reactors studied by [72]. NOB was successfully inhibited while effluent
793 NAR was average at 95%. This shows that the reactors will both accomplish PN. Zeolite plays the dual functions of
794 biomass carrier and ion exchange for NOB and AOB during adsorption with high selective NH_4^+ -N adsorption
795 potential. This in turn made SSBBR and NSBBR differ in FA concentrations. AOB in the SSBBR desorbed the high
796 NH_4^+ -N concentration, because its NH_4^+ -N adsorption capacity was 2–3 times larger than that of NSBBR.

797

798 **5.1.6 Fibrous biocarriers**

799 Achieving good TSS and COD removal efficiencies, SBBR coupled with the flexible fibre packaging
800 material will successfully yield excellent result. There is an inverse relationship between the efficacy of COD removal

801 with regard to changes in OLR. However, a direct relationship exists in the case of TSS performance [83]. The high
802 initial concentration of COD could lead to the biofilm absorption of aerobic organic matter. Thereby, transporting
803 organic matter into the internal biofilm, to serve as a source of carbon during denitrification [87]. Therefore, SBBR
804 systems developed with fibrous materials as biocarriers are highly efficient and energy saving wastewater treatment
805 technologies. It is apparent that SB-FFBR can withstand OLR to a certain degree and can handle very high
806 contaminated wastewater. However, with an increasing organic loading rate, the COD dissolution rate shows a rapid
807 decrease. The reduction in COD removal ability with an increment in OLR in the SB-FFBR could be due to the
808 decrease in contact time between microorganisms and substrates [78]. Similar to most biocarriers, the crucial
809 determinant for attaining high COD removal efficiency at the most leading OLR was the increased amount of biomass
810 concentration. This was due to the existence of fibrous packing media that provide a more precise surface area for the
811 growth of microorganisms. A research team in their study [75], attained low $\text{NH}_3\text{-N}$ removal efficiency. This result
812 revealed that autotrophic bacteria activity was inhibited by the concentration of carbon and that the efficacy of biofilm-
813 absorbed aerobic organic substances was simpler than the absorption of $\text{NH}_3\text{-N}$.

814 Increasing the biofilm density of a fibrous biocarrier from 15 to 50% resulted in greater biomass volume.
815 This show that the number of microbes has increased, resulting in low $\text{NH}_4^+\text{-N}$, TN and COD effluent concentrations.
816 However, the high biomass inhibited mass transfer as the biofilm density proceeded to rise from 50 to 70% and the
817 microbes were unable to access the organics. Thus, this affects the pace of COD elimination. Due to the large number
818 of microorganisms and consistent influent concentration, the average nitrogen load per biomass was also low, thereby
819 reducing TN removal rate. Hence, these two studies [1, 10], recommended biofilm density of 50%. In an SBBR system
820 made of rope-type media, the cyclic COD profiles showed that most biodegradable organic compounds were absorbed
821 in the anoxic process by facultative bacteria leaving the non-biodegradable component during the rest of the cycle
822 period [86]. Similar to other biocarriers, the increase in COD load seems to have resulted in high heterotrophic bacteria
823 growth rates during the liquid phase of the reactor. As it showed appropriate volumetric removal rates of total inorganic
824 nitrogen, the fixed-bed biofilm sequencing batch reactor (FbSBR) was regarded as an efficient and portable
825 mechanism.

826 827 **5.1.7 Composite packing**

828 In the experiment conducted with a sequencing batch biofilm reactor combined with a vertical flow
829 constructed wetland (SBBR-VFCW), the VFCW performed well in $\text{NH}_4^+\text{-N}$ dissolution. Plants and Clinoptilolite and
830 in the VFCW also demonstrated strong $\text{NH}_4^+\text{-N}$ absorption [14]. The SBBR-VFCW system could occupy less space,
831 work stably, run normally, cope with higher hydraulic loads, withstand impact loads and obtain greater purification.
832 Combination of GAC and natural zeolite as biocarriers in the anaerobic sequencing batch biofilm reactor (ASBBR)
833 improved biogas generation and COD removal rate. Observed effluent COD concentrations was 30-40 mg/L with 278
834 mL/d as the maximum biogas productions. Authors reported that GAC was responsible for the improved COD
835 degradation while zeolite was for gas generation [71]. The high biomass concentration within the moving-bed biofilm
836 sequencing batch reactor (MBBSBR) was due to the efficiency of the composite biocarrier in the aggregation and
837 immobilization of bacteria. This ensured the greater performance of treatment through high loading and allowed shock
838 loading to be strongly tolerated. Although the efficiency of COD declined by 8%, study also reveals that
839 microorganisms have increasingly adapted to the toxic environment after a pretty long operation and have lived on
840 organic compounds found in wastewater [65].

841 842 **5.1.8 Comparative analysis on nutrients and organic matter removal efficiencies**

843 The experimental results were compared based on treatment efficiency achieved due to the influence of
844 different biocarriers applied. From Table 3 below, it could be observed that PU, GAC, POC, inert stone chips, and
845 biocarriers made of composite packing achieved high and stable COD removal efficiencies. In some cases,
846 even at high OLR values. On the contrary, PP, MC, Fibrous biocarriers had low COD removal rates with
847 some even in a decreasing trend. The utilization of PE, LS, MC, ceramic as biocarriers result to high TN
848 removal. This could be due to biofilm bacteria and algae development on some of the biocarriers which enhance the
849 denitrification process. But when PU was used, low TN removal was recorded due to large PU size. $\text{NH}_4^+\text{-N}$ removal
850 efficiencies in SBBR supported with PU, PE, GAC, Zeolite, and biocarriers made of composite packing for organic
851 and nutrients removal alongside biofilm formation were high due to the exceptional properties possessed
852 by the biocarriers. Fibrous biocarriers had low $\text{NH}_4^+\text{-N}$ removal. Used tires removed TKN, BOD_5 , and TP at high

853 efficiency as SS and TSS were highly removed by GAC and Fibrous biocarriers respectively. PP and ceramics
 854 also removed TP at high efficiency.

855

856 Table 3: Comparative performance of various biocarriers in organic and nutrients removal

Biocarrier	Performance
PU	<ul style="list-style-type: none"> At higher organic loads, better removal of COD and $\text{NH}_4^+\text{-N}$ could be attained. Promotion of SND processes alongside an improved TN removal. However, TN removal decreases with decreasing PU foam cube sizes while $\text{NH}_4^+\text{-N}$ removal decreases with increased PU foam volume The removal of $\text{NH}_4^+\text{-N}$ may also be credited to the reality that the high pores of the PU biocarriers have been trapped by many microorganisms
PE	<ul style="list-style-type: none"> Highly efficient nitrogen removal was achieved. Nitrite reduction was also observed. High TN and $\text{NH}_4^+\text{-N}$ removal
PP	<ul style="list-style-type: none"> They just couldn't get the biofilm prepared for COD removal during the early days due to its smooth surface, which made it hard for microbes to bind. High-efficiency phosphorus removal
GAC	<ul style="list-style-type: none"> Even at higher OLRs, $\text{NH}_3\text{-N}$ and COD demonstrated high and stable removal High SS and color removal efficiencies could be obtained
Waste materials	<ul style="list-style-type: none"> Inner tube of used tires improved BOD_5, TP and TKN removal efficiencies. The TN removal efficiency could be enhanced by the denitrification process and further enhance external carbon availability in the LS. POC is said to be more efficient as COD managed to achieve the targeted effluent limit
MC	<ul style="list-style-type: none"> Achieved efficient nitrogen removal due to the biocarrier need for a lower volume of oxygen and organic sources to support the experiment. A continuous reduction in the efficiency of COD removal.
Ceramic	<ul style="list-style-type: none"> Enhanced removal of TN and TP primarily due to biofilm bacteria and algae development on the biocarriers
Inert stone chips	<ul style="list-style-type: none"> High COD removal even at higher OLR
Zeolite	<ul style="list-style-type: none"> High selective $\text{NH}_4^+\text{-N}$ adsorption potential
Fibrous biocarriers	<ul style="list-style-type: none"> Excellent result for TSS removal COD decreases at high OLR Low $\text{NH}_4^+\text{-N}$ removal
Composite packing	<ul style="list-style-type: none"> High $\text{NH}_4^+\text{-N}$ and COD absorption Improved biogas generation Occupy less space, work stably, cope with higher hydraulic loads, withstand impact loads and obtain greater purification

857

858 5.1.9 Biocarriers significant effect on the removal of hardly biodegradable compounds

859 Most hardly biodegradable compounds affect SBBR system performance at high concentration during initial system
 860 operational stage, irrespective of the biocarrier material type. Their presence reduces microbial activity and affects
 861 microbial community structure, diversity and composition. The protein and polysaccharide contents in the EPS of the
 862 biofilm and suspended sludge increases with an increase in hardly biodegradable compounds concentration. Their
 863 addition in the influent could affect the chemical structure of the LBEPS and TB-EPS.

864 Researchers [55, 56], reported that, after treatment with the pilot GAC-SBBRR, it was almost hard to foresee
 865 2,4-dichlorophenol (2,4-DCP) residues in the effluent. A study [54] using same biocarrier material treating
 866 pentachlorophenol (PCP) also reported good performance efficiency and stable treatment process under stern organic
 867 load fluctuations with about 61% PCP removal. Utilizing a rotating perforated tube as a biocarrier material increases
 868 DCP removal efficiency. This was proven by the large biofilm surface area at high A/Q ratios. Interestingly, increasing
 869 DCP feed concentrations improved DCP removal efficiency up to > 97% [89].

870 According to the findings of this review, it was discovered that two biocarriers were most prominent when
871 handling hardly biodegradable compounds. These are: (i) Fibrous carriers and (ii) combined packing that made of
872 polyester fiber. The fibrous biocarriers have been used by researchers to treat wastewater with significant
873 concentrations of oxytetracycline [90], chlortetracycline [91], sulfadiazine [92], florfenicol (FF) [93], norfloxacin [94]
874 etc. compared to several other biocarriers, the fibrous material has high resistance to toxic effects resulting from the
875 aforementioned compounds. Combined packing made of polyester fiber materials acting as carriers for microorganism
876 attachment have been utilized by several researchers specifically treating wastewater containing different proportions
877 of cerium oxide nanoparticles (CeO₂-NPs) under both short and long-term exposure conditions. It was reported that
878 CeO₂-NPs were adsorbed as aggregates as low concentrations did not affect the biofilms, and the resulting ROS
879 production increase influenced cell growth [95, 96]. The inhibitory effects of CeO₂-NPs with molecular oxygen were
880 reduced after anaerobic exposure [97].-At the end of aerobic exposure, the contribution of EPS to phosphorus removal
881 improved, while the capacity of EPS in phosphorus storage declined after exposure to CeO₂ NPs [98].
882

883

884

Table 4. Efficiency of biocarrier materials for nutrients and organic matter removal in SBBR

System	Reactor Capacity (L)	Biocarrier type	Wastewater type	Loading rate	Nutrients and organic matter removal (%)			Specific area	Filling ratio	Ref.	
					COD	NH ₄ ⁺ -N	TN				
SBBR	31	Linpor (PU)	municipal wastewater	0.5-8.0 gCOD/m ² ·d	93			270 m ² /m ³		[81]	
		Kaldnes (PE)			73.83			500 m ² /m ³			
		Liapor (Ceramic)			92.06			480 m ² /m ³			
ASBBR	1200	MC	industrial wastewater	OLR, 0.6-3.0 kgCOD/cycle; COD _{Total} , 0.98-5.08 g/L; NH ₄ ⁺ -N, 76-515 mg/L	86				0.83	[73]	
FB-SBBR	15	MC	NH ₄ ⁺ -N rich wastewater	0.55±0.10 kgNH ₄ ⁺ -N/m ³ /d			95		0.52	[19]	
AnSBBR	7.2	PU	domestic sewage	COD, 301±103 mg/L	60			43.8 m ² /g	0.51	[82]	
		VC		COD, 347±87 mg/L	40						
		SP		COD, 446±169 mg/L	48						
		PE		COD, 337±82 mg/L	33						
SBBR	35	PU	synthetic wastewater	0.24-1.26 g N/(L·day); COD, 372-382; NH ₄ ⁺ -N, 39.8-209.5 (mg/L)	86.88	74.6	60.5		0.8	[37]	
SBBR	10	PU	domestic wastewater	51–85 mg N/L/d	77.44	99.44	88.2		0.2	[44]	
ASBBR	5.5	PU	synthetic wastewater	COD, 1000 mg/L	78-83					[42]	
MBSBRs	12.8	PU	synthetic wastewater		70		84	177 m ² /kg	0.08	[39]	
MBSBRs	12.8	PU	synthetic wastewater		64		59.5	177 m ² /kg	0.08	[41]	
AnSBBR	10	PU	synthetic wastewater	0.4 and 3.2 COD/SO ₄ ²⁻	67			43.8 m ² /g		[79]	
		EC			81			3.51 m ² /g			
SBBR	10	PU	synthetic wastewater	0.78-1.7 kgN/m ³ /d			93	2395 m ² /m ³	0.15	[33]	
ASBBR	7.2	PU	cane vinasse	OLR, 0.5-5 g/L.d, 9.6 gCOD/L	70					[43]	
MBSBRs	8	PU	synthetic wastewater	NH ₄ ⁺ -N, 50 mg/L			99	71	900 m ² /m ³	0.2	[66]
		LS					99	78	850 m ² /m ³		
SBBR	10	PU	synthetic wastewater		80	100			0.02	[80]	
		PE			76.59	96					
SBBR	28	PE	autotrophic medium				75	407 m ² /m ³	0.47	[49]	
MBSBR	6.5	PU	municipal sewage	COD, 213-286; NH ₄ ⁺ -N, 21.1-28.5 mg/L	92.43	97.89	83.73	>20,000 m ² /m ³	0.3	[40]	
ASBBR	1200	PU	domestic sewage	0.2–1.2 kg COD/m ³ /d	62					[38]	
SBBR	2	PE	diary wastewater	1130-1560 gBOD ₅ /m ³ ·d	81.8	85.1		150 m ² /m ³	0.3	[17]	

SBMBBR	5	PE	synthetic wastewater	COD, 649.44 mg/L	97				0.05-0.1	[4]
ASBBR	3.4	PE	synthetic wastewater		94.8		95.0	500 m ² /m ³	0.4	[27]
SBBR	9	PE	municipal wastewater	500-1000 mg/L COD	90	68.2	60.1	460 m ² /m ³	0.35	[31]
FB-SBBR	14	PE	synthetic wastewater		95					[16]
		Volcanic pumice stone			92.9					
SBBR	30	PE	synthetic wastewater	0.5-2.3 kg of COD/m ³ .d	99	99		690 m ² /m ³		[47]
SBBR	30	PE	synthetic wastewater	COD, 1460±2; NH ₄ ⁺ -N, 265±2 (mg/L)	93.7	99.1	97.5	369.6 m ² /m ³	0.4	[48]
MB-SBBBR	5.5	PE	acid Red 18 (AR18)		83.07			415 m ² /m ³	0.5	[45]
SBBR	20	PE	pharmaceutical wastewater	COD, 10000 - 10050; NH ₄ ⁺ -N, 1875 - 2000 (mg/L)		98.4		480 m ² /m ³	0.23	[3]
SBBR	3.4	PE	ammonia wastewater		99.6	98.2	61.2	500 m ² /m ³	0.4	[18]
SBBR	28	PE	poly-b-hydroxybutyrate			66		407 m ² /m ³	0.47	[46]
IFAS-SBR	800	PE	municipal wastewater	COD, 150-500; NH ₄ ⁺ -N, 50 (mg/L)	98.6	99		800 m ² /m ³	0.5	[26]
SBBR	27.6	PVC	domestic-strength synthetic effluent	8.8 g COD _T m ² /d and 1.2 kg COD _T m ³ /d	94			240 m ² /m ³		[51]
MBSBBR	15	PP	synthetic wastewater	0.84–0.978 g COD/L/d; 0.074–0.106 g N/L/d	97.7		87.8	378 m ² /m ³	0.1	[30]
SBBR	19.8	PP	sewage treatment		83	80			0.5	[67]
		LS			89	90				
SBMABR	2.6	PP	synthetic wastewater		90	96	91			[50]
SBR-BF	1000	PP	urban sewage	NH ₄ ⁺ -N, 35; TP, 13; COD, 400 (mg/L)	95		94		0.3	[21]
GAC-SBBR	2200	GAC	recycled paper wastewater	700-1000 mg COD/L	97.2	99.4				[55]
GAC-SBBR	2200	GAC	recycled paper wastewater	0.7 OLR (kg COD/m ³ .day).	94	98				[56]
MM-SBBR	18	GAC	paper mill wastewater	750-1900 mg COD/L	98	100				[54]
GAC-SBBR	5	GAC	recycled paper mill wastewater	800-1300 mgCOD/L	99	94				[53]
HG-SBR	7	GAC	synthetic wastewater	632 ± 50 mgCOD/L	96					[59]
SBCR	2	GAC	dye-containing wastewater	COD, 250-2500; C.I. Acid Orange 7 (AO7), 125-625 (mg/L)	88					[60]
Expanded bed biofilm reactor	7	GAC	zinc and copper containing wastewaters	COD, 3000; Zn, 200 (mg/L)	90					[57]
Spent GAC–biofilm-SBR	1.8	GAC	methyl orange (MO)-wastewater	COD, 541; MO, 175 (mg/L)	86					[61]
GAC–biofilm SBR	1.8	GAC	diazo dye Reactive Black 5 (RB5)-	COD, 467; RB5, 200 (mg/L)	94					[62]

SBGR	1.7	GAC	chemical wastewater	OLR: 1.7 kg COD/cum-day	78					[58]
SBBR	5	Steel disk	synthetic wastewater	TN, 7-140; NO ₃ -N, 7 mg/L			99			[32]
MB-aerobic-SBR	10	Inner tube of tires	synthetic wastewater	COD, 450±49; TP, 5.0±0.55 (mg/L)	97.5					[63]
SBBR	2	Iron shavings	artificial wastewater	COD, 100±5; NO ₃ ⁻ -N, 41.5±1.5 (mg/L)			90.36	0.002634 m ² /g	0.7	[68]
SBBR	10	POC	synthetic wastewater	COD, 50-100; NH ₄ ⁺ -N, 4-8 (mg/L)	67	90			0.4	[64]
FSBR	12	Peach Pit	synthetic wastewater	OLR, 2–12 kg COD/m ³ /d	98.77					[9]
SBBR	2.5	WAS pellet	synthetic wastewater	NLR, 120; OLR, and 60 (g/m ³ -d)	93	99	95			[29]
SBBR	4	Ceramic	domestic wastewater		90		65.8	900 m ² /m ³	0.2	[74]
SBBR	1.4	Inert stone chips	chemical wastewater	0.92- 4.76 kg COD/cum-day	88.05				0.49	[20]
SBBR	18	Fiber threads	domestic wastewater		90					[75]
SBFFBR	8	Rayon fiber	dairy wastewater	OLR, 0.38-2.74 kg COD m ³ /d	97.2			2200 m ² /m ³	0.9	[83]
SBBR	5.3	Semi-soft fiber	municipal wastewater	COD, 192.82; TN, 57.34; NH ₄ ⁺ -N, 51.37 (mg/L)	85	94.2			0.7	[1]
ASBBR	2.4	semi-soft fiber filler	mustard tuber wastewater	COD, 3600-8200; TN, 560-1100; TP, 8-19 (mg/L)	91.3				0.7	[10]
SB-FFBR	8	Flexible fiber	milk process wastewater	OLR; 0.38–8.19 gCOD/m ³ /d	97.5			2200 m ² /m ³	0.9	[78]
FbSBR	21	Rope-type media	piggery wastewater	COD, 2000; NH ₄ ⁺ -N, 350 (mg/L)	85		80			[86]
SBBR	24	Porous polymer	Livestock and Poultry Breeding Wastewater	COD, 1000–2500; TN, 125–175 NH ₄ ⁺ -N, 85–123 (mg/L)	96.1		92.1	5357 m ² /m ³	0.05	[52]
HSBR	1420	Nylon nets	municipal sewage	0.51 kg COD/m ³ .day and 0.06 kg NH ₄ -N/m ³ .day	92	99		4140 m ² /m ³		[87]
SBBR	0.5	Aluminum foil	synthetic wastewater	NH ₄ ⁺ -N, 100.4 ± 1.5 mg/L		93.7	77.5	500 m ² /m ³		[88]
SBMBfR	2	Fibrous slag	synthetic wastewater				96			[84]
MSBR	25	<i>Acrylic plastic</i>	milk industry wastewater	680 g BOD ₅ /m ³ d	97.9					[8]
SBBR-VFCW	60	PVC, soft PE and porous aggregates	domestic wastewater	COD, 276.20; TP, 11.15; NH ₄ ⁺ -N and TN, 79.25; (mg/L)	97	98.5	91.5			[14]
ASBBR	1	GAC and natural zeolite	synthetic municipal wastewater	COD, 1000 mg/L; OLR, 400 g/m ³ d	98					[71]
FSC-SBBR	10	PU and two fiber balls	synthetic wastewater	COD, 250–380; NH ₄ ⁺ -N, 40–50; TP, 10–15 (mg/L)	93.39	96.66		2.15-2.21 m ² /g	0.6	[85]
MBBSBR	2	PE and inorganics	coking-plant wastewater	OLR, 0.449 kgCOD·m ³ /d	92.9			900 m ² /m ³	0.3	[65]
SBBR	9.7	Activated carbon fiber filler	swine wastewater	COD, 4684; NH ₄ ⁺ -N, 846; TN, 1062; TP, 31.9 (mg/L)	98.2	95.7	95.6			[76]

886 **5.2 Biocarrier influence on biofilm formation**

887 It is clear that biofilm configured systems are best fit for the remedy of wastewater comprising poorly degradable
888 mixtures [58]. Apart from hydrodynamic conditions and substrate loading rates, the properties of different biofilm
889 carriers hold important implications concerning biofilm maturity and in situ sludge minimization. The biocarrier
890 concentration is obtained based on the percentage computed from the proportion of the carrier media's capacity to the
891 reactor effective capacity [80]. In addition, biocarriers set on the water accumulator can not only fasten abundant
892 microorganisms, but maintain the filtration function, letting the solid-liquid separation easier. To predict biomass
893 adhesion and inceptive biofilm development on supports, surface tension parts of all supports and biomass is required
894 to be computed by employing experimental values of contact angles [49]. Filamentous bacteria are important
895 components for biofilm formation. However, different processes such as abrasion, erosion, sloughing and predator
896 grazing can lead to their detachment [25]. The growth of films on the biocarrier cover can be evaluated utilizing a
897 flow cell device [19]. Therefore, this sub-section would focus on the effect of various biocarriers in biofilm formation.
898

899 **5.2.1 Thermoplastic**

900 **5.2.1.1 Polyurethane (PU)**

901 The appearance of PU foam cubes as biocarrier can create an integrated aerobic, anaerobic and anoxic
902 microenvironment, thereby providing ensnared carbon substrate that stimulates the SND process [39]. Aerobic
903 conditions are usually retained on the surfaces of the PU foam cubes as DO inclination befell along the PU foam's
904 interior depth [66, 80]. PU is extremely suitable in anaerobic digestion during biomass immobilization [82]. Therefore,
905 understanding the recurrence and diversity in the microbial morphologies in the PU foam cubes associated biofilm
906 [38].

907 Biofilm formation in an MBSBR was successfully achieved by [40], due to the porous nature of the biocarrier
908 (PU foam) utilized, enabling small molecules such as oxygen to enter through the pores within the biocarrier and
909 making the biofilm have distinct structures and micro-profiles. Water also entered the pores, presenting a way to
910 convey substances inside the biocarrier to support bacterial growth. This could be due to the fact that the thick porous
911 layer was able to effectively trap and capture biomass, and by having a large surface area, the reticulate structure
912 facilitated biofilm accumulation. To further improve and sustain biofilm formation using PU foam, limited aeration
913 was provided to create a favorable condition for biomass holding and a single oxic-anoxic interface for the
914 enhancement of slow-growing anammox bacteria in an SBBR system [44]. Attached-growth biomass was the
915 prevailing sludge morphology formed on the surface of biocarriers as depicted in Fig. 21. AOB and anammox bacteria
916 development was successfully enhanced in the system. This could be attributed to the application of PU as a biocarrier
917 and subsequently great nitrogen removal effectiveness of the reactor. Similar to findings by [37],
918 NOB, *Nitrospira* and *Nitrobacter* gradually increased through the operational period. Even though, the Maximum
919 NOB activity was limited to approximately 1.1 mg N·g/VSS/h [44].

920 An increase in biofilm biomass with concentration varying from 0.30 - 0.55 g TVS/g foam was noticed from
921 the formulation of biopolymers (EPS) when [38] studied the efficiency of an ASBBR in the management of domestic
922 sewage subjugated by anoxygenic phototrophic bacteria. The EPS could be responsible for the enhanced biofilm
923 adhesion to the inorganic matrix. More so, the presence of fluorescent rods, anoxygenic phototrophic bacteria and
924 other archaeal populations with any methanogenic organisms like *Methanosaeta* and *Methanosarcina*-like cells were
925 observed at an oxidation–reduction potential ranging within -280 and -360 mV. From the perceived great frequency
926 of these phototrophic organisms inside the biocarrier interstitial spaces, it can be concluded that PU have cooperated
927 as a second substrate and micro-traps improving the retention of microorganisms during the reactors release stages.

928 Investigating the microbial succession within an ASBBR treating cane vinasse [43], optical microscopy
929 showed high microbial morphological diversity. Rods of multiple morphotypes and filaments commonly associated
930 to anaerobic degradation pathway hydrolysis and fermentation processes have been frequently observed. It was also
931 noted that the mechanically preserved micro-granules inside the PU were neither different when compared to their
932 size nor their microbial morphotypes.
933



Fig. 21. Formation of biofilms surface on biocarriers in the SBBR (left), and biocarriers filled with biomass on (right) (Adapted from [44]).

934

935

936

937

938

939

940

941

942

943

944

945

946

947

948

949

950

951

952

953

954

955

956

957

958

959

960

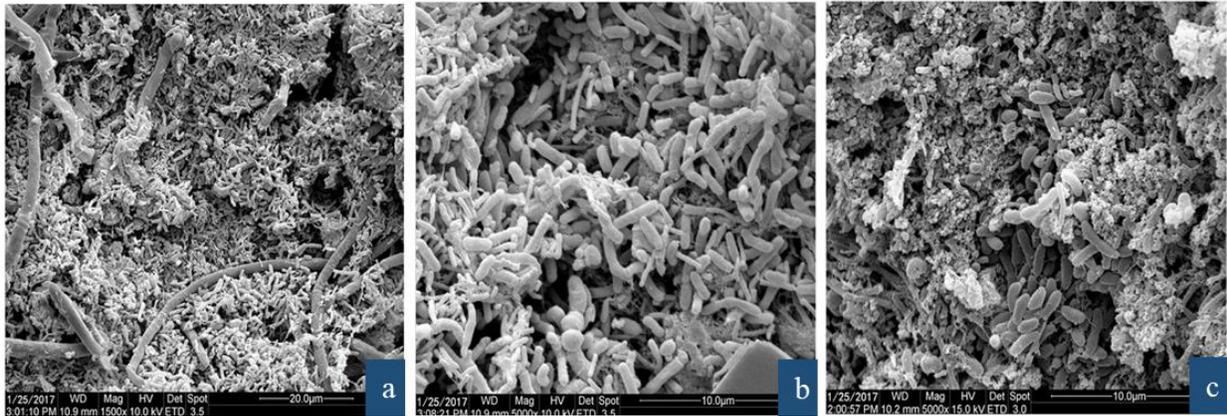
961

962

The hydrodynamic attributes of ASBBR could be the result of these observations, that reduces the impact of substrate delamination and mass transference inside the biofilm. The prevalent archaeal community inside the PU matrices was *Methanosarcina*-like microorganisms at high organic loading ratios beyond 3.3 gCOD/L.d. This has been proposed to be correlated with a higher specific rate of microorganisms' acetate intake. In a related study, when an ASBBR was filled with PU foam for treating sulfate-rich wastewater [79], the key organic matter was transformed by complete- or incomplete-oxidizing sulfate-reducing bacteria (SRB), in supplement to acidogenic microorganisms, without the use of sulphate as the electron donor. SRB was found to represent > 65% of the bacterial community.

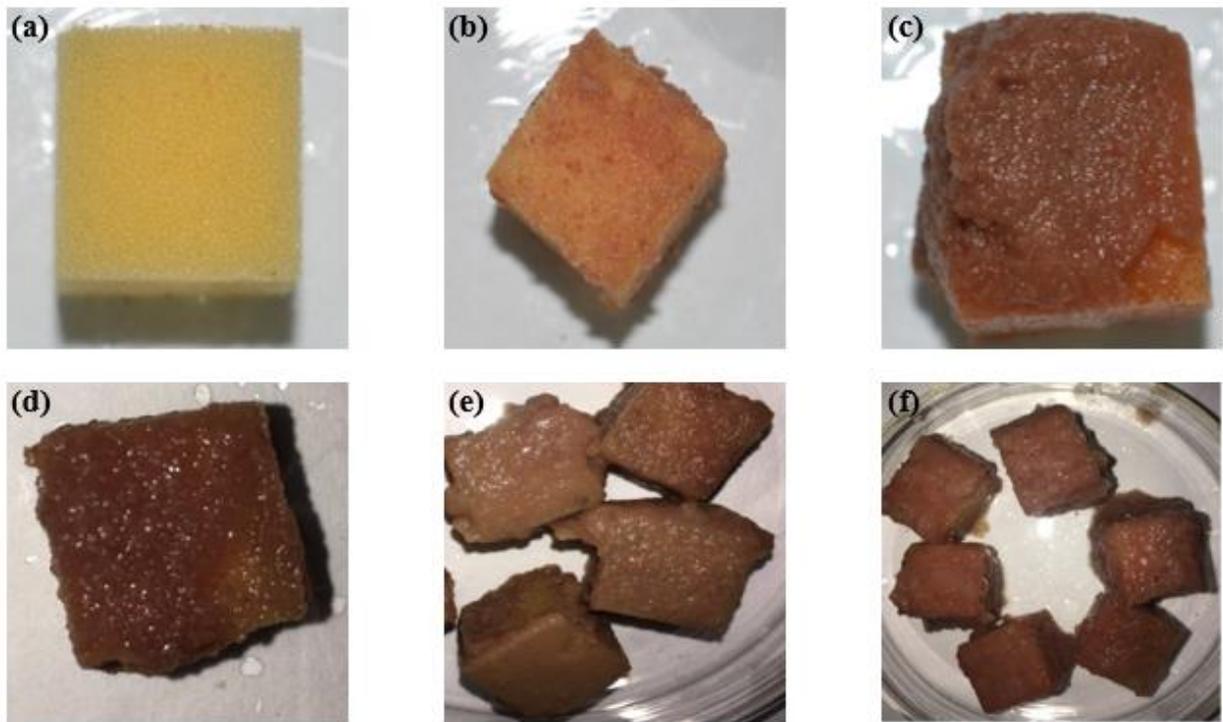
Microbial community similarity in an SBBR operated by [37], decreased with the increase of NLR. The sequencing outcomes confirmed that, with two complicated biofilm systems in the microbial ecology, nitrogen extraction was accomplished by *Nitrobacter sp.*, *Nitrospira sp.*, uncultured AOB, *Pseudomonas fluorescens*, and *Pseudomonas butanovora*. Interestingly, the bacteria quantity on the biocarriers improved following inoculation because of progressive immobilization of huge bacteria over the PU foam.

SEM photographs depicted in Fig. 22 unveiled that the bulk of the spherical and elliptical bacteria were besieged by the EPS and bacillus or filamentous bacteria in the reactor enrichments in period I. However, in periods III and IV, few or even no bacilli and filamentous bacteria were detected, suggesting good bonding between the biocarrier and the bacterial populations. Still on the effect of NLR in a PU foam filled reactor, biofilm system in a denitrifying ammonium oxidation (DEAMOX) process had powerful adaptability to fluctuating loading rate. The huge organic matter contents in SBBR led to the growth of heterotrophic denitrifying bacteria on the exterior of biofilms. The biofilm slowly grew thicker and became red (Fig. 23c, d), symbolizing that anammox bacteria evolved. As the NLR increases, a coating of yellow-gray bacteria, known to be heterotrophic denitrification bacteria, started to cover the biofilm surface, while the interior layer remained red well in the biomass attached on the biocarriers. The biomass densities in biofilms risen from 1000 plus to 3819 mgVSS/L over the whole duration of operation. This symbolizes that a mature and stable biofilm had been developed on biocarriers [33]. The concentration of anammox bacteria in biofilms rose from 6.08×10^9 to 1.00×10^{11} copies/g dry sludge by 2 orders of magnitudes, suggesting that anammox bacteria conformed more adequately to carriers, with the biofilm eventually approaching maturity.



963
964
965

Fig. 22. SEM of biofilm grown on PU foams a, b, and c at different operational periods I, III and IV, respectively (Adapted from [37]).



966
967
968
969

Fig. 23 Biofilm apparent morphology at different periods: (a) day 0; (b) day 9; (c) day 67; (d) day 91; (e) day 138; (f) day 196 (Adapted from [33]).

970
971
972
973
974
975
976
977

The effect of applying different biocarrier size in an SBBR have been extensively studied, because biocarrier size affects mobility of cubes to improve system performance [39, 41, 42, 49]. The effect could be better understood by studying the different mass transfer phenomena occurring in a biofilm reactor, namely: classic theory, intraparticulate convection theory, and convection inside the biofilm. The classic theory postulates that the smaller the particle, the better the external and internal mass transfer conditions. Intraparticulate convection states that the magnitude of this intraporous convection normally increases as biocarrier diameter increases. While, the theory for convection inside biofilm, state that, the biofilm structure may resemble a “tulip” or “mushroom”, whose superior parts can coalesce, originating channels through which wastewater acting as nutrients can flow.

978
979
980

Maximum mass transfer could possibly be achieved when five phenomena (foam packing, liquid-phase mass transfer rate within the frame surrounding the bioparticle, intraparticulate diffusion, intraparticulate convection and internal fluxes in the biofilm) simultaneously occur in SBBR. Because of interior biofilm composition, outer liquid

981 fluxes presumably affect the diffusion and convection that take place inside it; thus, the mass transfer inside the biofilm
982 may be favored by an enhanced participation of convection in the macropores. Findings by [42] and [49] contradicts
983 the classic theory as better mass transfer was experienced at higher particle size. This could be associated with the
984 numerous magnitudes of the detachment forces in the reactors mainly induced by collisions within supports. It was
985 also observed that, higher detachment forces are expected to occur in the reactors operating with smaller size
986 biocarriers as higher quantity of support pieces might have stalled down initial biofilm accumulation.

987

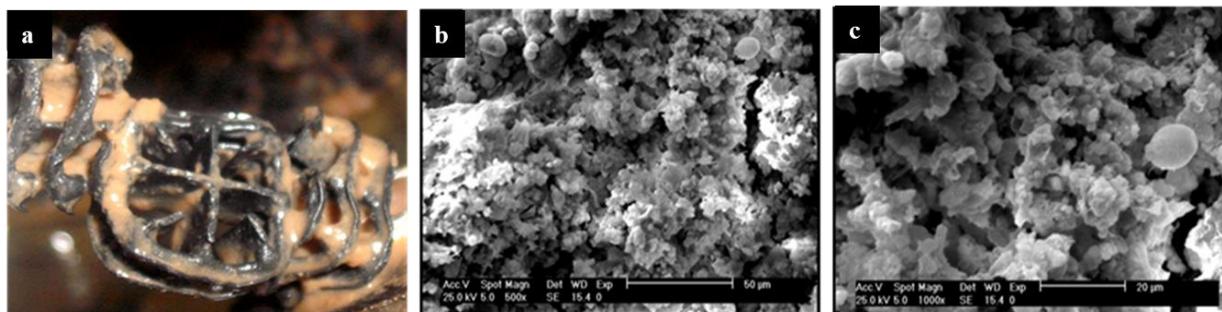
988 **5.2.1.2 Polyethylene (PE)**

989 In an SBBR operated with PE as a biocarrier, biofilm can be distributed between the internal and external surfaces of
990 the PE biocarrier material [46]. This is an advantage that PE biocarriers have over other biocarrier materials [3]. Also,
991 among the several thermoplastic materials used for biofilm formation, PE has proven to be the most suitable bed
992 material for the appendage of methanogenic microorganisms even though it has less satisfactory performance in
993 nutrients removal [48]. Amazingly, PE can be pretreated to enhance its physical properties (surface area, roughness,
994 and porosity). However, PE did not permit any form of microorganism to predominate [17, 27]. In tightly-bound
995 biofilm, biofilm adhesion and compliance were the most substantial, with the weakest in supernatant/surface biofilm.
996 According to [31], supernatant biofilm could be separated by lower centrifugal force (2000 \times g) while the tightly-
997 bound biofilm require higher centrifugal force (10,000 \times g) to separate from biocarriers.

998 According to [82] in their study reported that a huge number of microorganisms is discovered on PE as it did
999 not present adequate characteristics for promoting microbial growth. The relatively low porosity of PE did not permit
1000 for the appendage of an adequately huge concentration of VS on its surface to improve the craved performance. Hence,
1001 fit anaerobic biomass was discovered when testing PE as an immobilization support material. A study by [49] keenly
1002 observed that biofilm build-up was favored on the supports that granted tremendous interior surface area and shielded
1003 biofilms from erosion and abrasion separation mechanisms. The geometry of the collisions among particles and the
1004 hydrodynamic events instituted in the SBBRs appeared to perform a significant role in biofilm development than the
1005 thermodynamic synergy, expressed as free energy of adhesion (ΔG), among the biomass and support material. Other
1006 factors important in the biomass adhesion process, and are not regarded by the thermodynamic approach include:
1007 bacterial molecular structure and growth conditions followed by medium ionic force and composition.

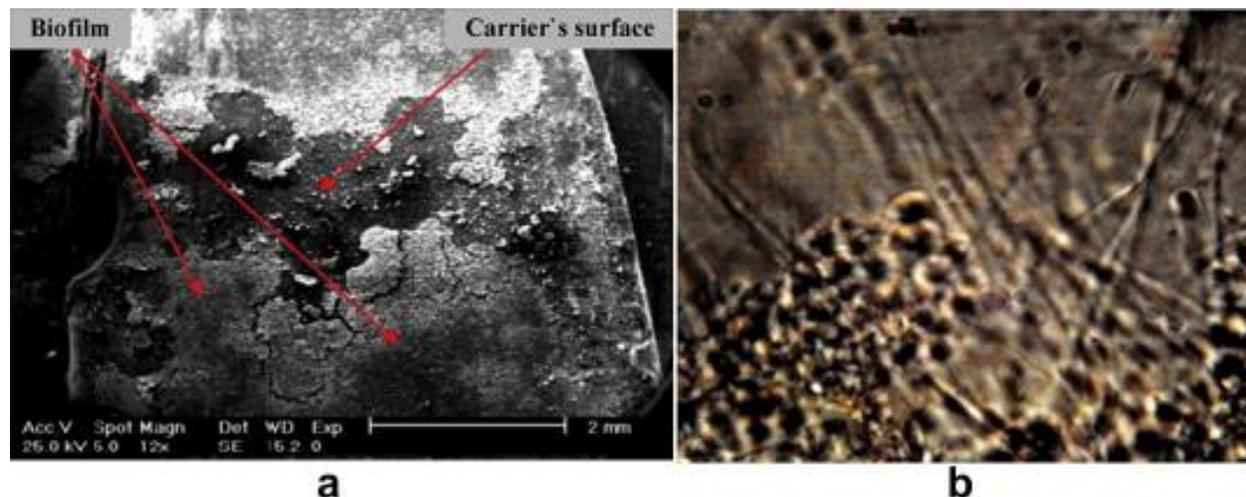
1008 The performance of PE biocarrier was also investigated by [47]. It was discovered that the dense particles
1009 inside the interstitial openings of the PE biocarrier were barely distinguishable from the biofilm fastened to the
1010 biocarrier surface as the huge biomass concentration in the bed manages to gather all the biomass. But, sampling a
1011 small volume of the bed and suspending it in water, it became evident that the reactor is made of two distinct fractions:
1012 the biofilm that lives fastened to the carrier and highly impenetrable aggregates (granules) that mechanically separated
1013 from the carrier. The PE biocarrier also aided biomass granulation through the following: (i) development of a faint
1014 biofilm that wholly masks the carrier; (ii) increment of biofilm thickness; (iii) break-up of the ascribed biofilm with
1015 the liberation of biofilm particles; and (iv) rearrangement of biofilm particles in smooth granules. Biofilm breakup
1016 can be caused by the continual increase in shear force that has significant adverse impact on biomass stableness.
1017 Biofilm is considerably impaired by an increment in shear force, because it develops with a pervious structure and
1018 little adhesion strength (18-22). Thus, resulting to biomass sloughing.

1019 The positive role performed by the active biomass built as a biofilm on the coverings of the PE biocarrier
1020 media which beget a weaker responsiveness to toxicity impacts and other environmental provisions illustrates the
1021 divergent patterns exhibited by the SBBRs [80]. As depicted in (Fig. 24b and 24c), biofilm raised on coverings of the
1022 PE media had a fluffy arrangement. According to the authors [16], the microstructure of biofilm grown on PE when
1023 used as fixed-bed is different from that grown when used as moving carrier. Biofilm fastened to the moving carriers
1024 shows a profoundly permeable arrangement in which great number of filamentous bacteria lives. However, the biofilm
1025 appended to the fixed media had a compact arrangement with no filamentous bacteria noted.



1026
1027 **Fig. 24.** Optical and SEM photographs of biofilm grown on surfaces of PE packing media in FB-SBBR (Adapted
1028 from [16]).
1029

1030
1031 Similar to other studies that uses PE as a biocarrier, notable part of the entire biomass in an MB-SBBRs
1032 belonged to the attached-growth biofilm as the ratio of the biofilm to the suspended biomass concentration was
1033 achieved 0.51- 0.76 [45]. The SEM images of the biofilm raised on the coverings of moving biocarriers in MBSBBRs
1034 was obtained. As shown in Fig. 25a and 25b, after about 50 days, several surfaces of moving biocarriers were shielded
1035 by a brown layer of attached biofilm dominated by filamentous bacteria. It was also noticed that the community of
1036 filamentous bacteria was abundant in the suspended biomass in MBSBBRs.



1037
1038 **Fig. 25.** SEM (a) optical microscopic (b) images of biofilm grown on surfaces of biocarriers in MB-SBBRs
1039 (Adapted from [45]).
1040

1041 Utilizing the Kaldnes ($K_1, K_2, K_3, K_4,$ and K_5) form of PE encouraged nitrite production at the biofilm surface,
1042 inducing a huge concentration slope with an originally low nitrite concentration in the bulk fluid through nitrification.
1043 The expanding biofilm in Kaldnes exhibits features that make dispersion into the deeper layers hard. Thus, The
1044 composition of the biofilm influences the transfer of nutrients to and from the biofilm [81]. Unlike in PU foam cubes,
1045 biomass in the suspended flocs and in biofilm lowered as the organic loading reduces when PE biocarrier was
1046 employed. which demonstrated the consequences of the competition for oxygen and space among heterotrophic and
1047 autotrophic bacteria The percentages of nitrifying associated genes above cumulative genes remained higher in
1048 biofilms as correlated to those in AS flocs, symbolizing biofilm was a major favorable environment for nitrifiers [26].

1049 During the biodegradation of mono azo dye-reactive orange 16 in sequencing batch moving bed biofilm
1050 reactor (SBMBBR), suspended-growth biomass controlled higher than 76% of the biomass system. The smaller
1051 dominance of attached-growth biomass herein could be associated to the: inoculated quantity of AS into SBMBBR;
1052 suspended biomass owns a more porous floc arrangement than attached biofilm. The SBMBBR system was confirmed
1053 to be capable of fighting mono azo dye-reactive orange 16 (RO16) shocks. However, a longer cyclic reaction time,
1054 higher biocarrier filling ratio, and post-treatment are needed to accomplish greater efficiency [4].

1055 In a PN-SBBR system for nitrogen removal via nitrite treating high strength ammonia wastewater [18], the
1056 achievement of PN-biofilm was accomplished by variations in effluent quality and biofilm accumulation. Thin biofilm
1057 was easily attached onto the surface of PE biocarrier and ultimately, the biofilm thickness was commonly established
1058 in the system. Consequently, biofilm ambushed inside the biocarrier progressed to develop until stable operation of
1059 PN-system was achieved. At the end of the investigation, the cumulative quantities of biomass in the attached
1060 biocarrier and suspended phase were 5.35 and 0.5 g/L, respectively, insinuating that the larger share of the biomass
1061 was fastened over the biocarrier in PN-system.
1062

1063 **5.2.1.3 Polypropylene (PP)**

1064 PP has been confirmed to possess a longer life span than other thermoplastic materials used as biocarriers in biological
1065 wastewater treatment [49, 67]. BioBall® renders a more dependable appendage medium than traditional plastic
1066 biocarriers for the maturity of microorganisms. In the study by [30], biofilm was evenly cultured covering the surface
1067 of BioBall® biocarriers, even though the layer was apprehended gradually due to the absence of microorganisms
1068 adhesion. The whole surface of the BioBall® media became shielded with a biofilm layer ≈ 1.2 mm thick.

1069 The rise of biofilm thickness enhances the mass transfer resistance and decreases the internal mass transport
1070 efficiency of biofilm. Thus, backwashing process could be used to effectively control biofilm thickness. When
1071 backwashing process was employed by [50] in sequencing batch membrane-aerated biofilm reactor (SBMABR), the
1072 facultative aerobe and the anaerobic bacteria outside the biofilm did wipe off, while AOB inside the biofilm was
1073 properly stored. The little suspended microbes held in the effluent lead to the somewhat higher COD concentration
1074 discovered in the SBMABR system.

1075 A hybrid system was successfully developed in an sequencing batch reactor-biofilm (SBR-BF) system with
1076 suspended and AS alongside attached biofilm [21]. In addition to rendering safe living circumstances for PAOs,
1077 denitrifying phosphate accumulating organisms (DNPAOs), nitrifying and denitrifying bacteria, the device has also
1078 restrained rivalry among different bacteria for carbon sources. Meanwhile, on suspended carriers, a coating of thin
1079 transparent yellow biofilm was noted.
1080

1081 **5.2.1.4 Polyvinyl Chloride**

1082 It is a known fact that packed biofilms perpetually possess added openings. There is also a tendency for an increment
1083 in porosity with rising biofilm thickness. Authors [25], studied biofilm features and maturity with two tanks in an
1084 alternating pumped sequencing batch biofilm reactor (APSBRR) and confirmed that biofilm density decreases as the
1085 biofilm thickness increases in both tanks. The transformation of the biofilm density could also be due to varying
1086 morphologies and cell lysis. It was noted that the biofilm in Tank 2 was viscous and dense whilst a coagulated biofilm
1087 developed in Tank 1. Investigating the efficiency of a SBBR in organic carbon and phosphorus extraction, biofilm
1088 thickness increased with 96.2% as the medium moisture content of the biofilm. The highest total solids of the biofilm
1089 was 42.2 g/L [51]. Biofilm growth commenced on the module at the start-up of the reactor as the excessive growth
1090 made the module to block following long time work. This could be evaded if a larger amount of water was beyond
1091 the module.
1092

1093 **5.2.1.5 Acrylic plastic**

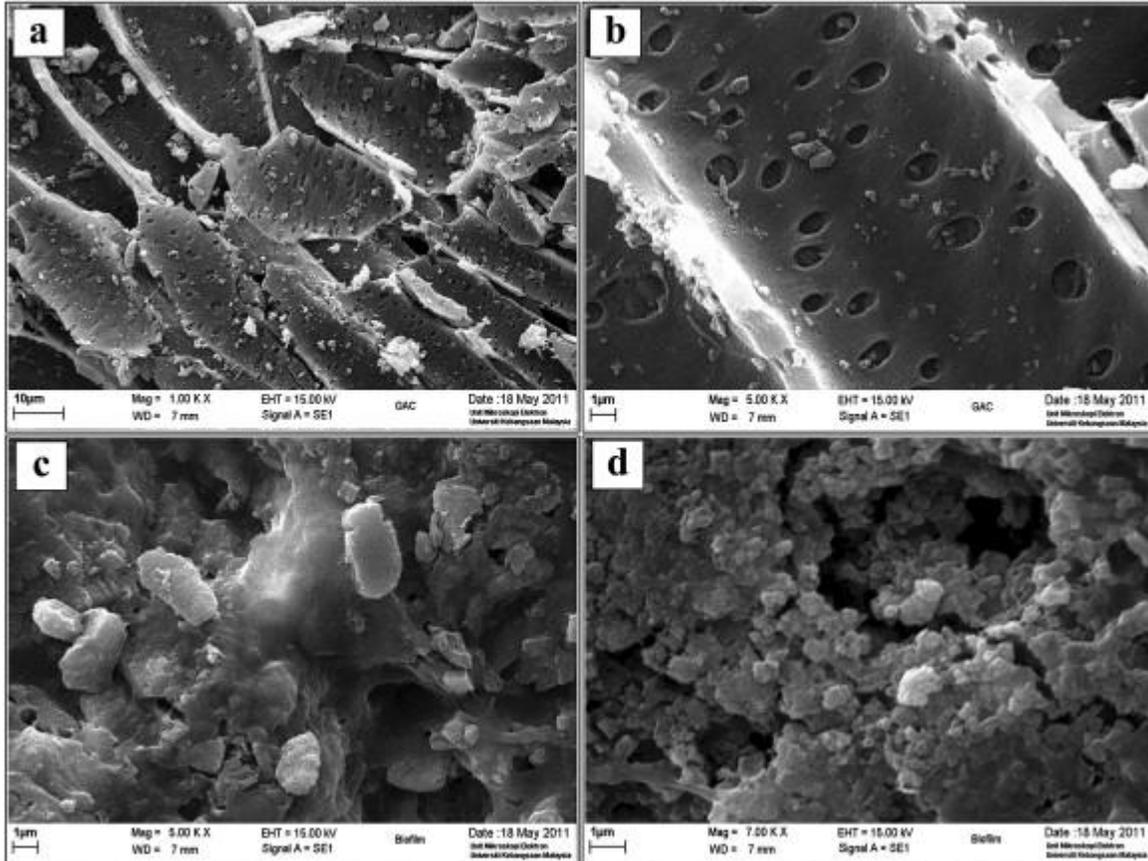
1094 It was discovered that by placing an acrylic plastic media (2.7 m² surface area) at the base of an SBR system
1095 to get an MSBR system, the employment of an attached growth system will raise the quantity of biofilm piece on the
1096 media, increase the efficacy of extraction and persistence of sludge, decrease the quantity of surplus bio-sludge and
1097 also lessen the acclimatization time of the system [8].
1098

1099 **5.2.2 Granular Activated Carbon (GAC)**

1100 In SBBR systems, GAC is being added as a biocarrier to support the production of biofilm as AS harboring
1101 microorganisms are sustained in suspension [53, 54]. The numerous benefits of using GAC as a biocarrier in SBBR
1102 systems are: long preservation of biofilm, suitable for treating poorly degradable compounds, provides less toxicant
1103 concentration in the bulk fluid due to sorption process, decreases biomass susceptibility to toxicants and successful
1104 increase of decolorization rate by increasing co-substrates concentration [58, 60, 62].

1105 According to the findings by [57], the synergy amongst heavy metals and biofilm occurred in the adsorption
1106 of heavy metals upon biofilm, which slowly decreased the aqueous metal concentrations. More so, during the
1107 application of spent GAC by [61], the low reductive condition at the uppermost layer promoted the maturity of aerobic
1108 microorganisms and prompted the mineralization of medium aromatic amines. The GAC particles coated with biofilm

1109 were discovered to present a slight brownish colour [55]. Fig. 26 displays the optical microscopy as observed by SEM
1110 of the GAC molecules. Virgin GAC, with a dimension of 1.3 x 0.3 mm of porous structure, shown in Fig. 26 (a) and
1111 (b) offer niches of shelter from the accompanying shear forces and maintain surface biofilm [56]. It also contributed
1112 to the growth of microorganism and improved the GAC-SBBR's ability to extract COD and Adsorbable Organic
1113 Halides (AOX). The morphology of the biofilm shown in Fig. 26 (c) and (d) was discovered to be complex, with a
1114 staggering dense covering texture developed on the exterior of the GAC with a great interior permeable network.



1115
1116 **Fig. 26.** Microscopic photographs for (a, b) virgin GAC and (c, d) biofilm GAC observed by SEM (Adapted from
1117 [55]).
1118

1119 A Hybrid-Growth Sequencing Batch Reactor (HG-SBR) system worked under immense absolute biomass
1120 concentration emanating from the production of bio-sludge and biofilm mass [59]. A positive relationship exists
1121 amidst MLSS and organic loading in HG-SBR. The increase in the values of MLSS and mixed liquor volatile
1122 suspended solid (MLVSS) was possible due to the growth of the microorganisms attached to the GAC. The co-
1123 existence of biofilm and bio-sludge enhanced the effectiveness of HG-SBR due to immense biomass array.
1124 Observation from the SEM images in Fig. 27 revealed that there stood a contrast amid the raw GAC and biofilm GAC.
1125 Cocci-shaped bacteria with bacterial diameter 0.25-1.0 µm prevalently built on the exterior of GAC biofilm (Fig. 27b)
1126 symbolized the creation of attached germination of microorganisms. A similar structure has also been noted in (Fig.
1127 27c) alongside spherical and irregular shapes because multi culture microorganisms of AS were adopted during
1128 inoculation. The cocci-shape bacteria were nevertheless, the principal shapes that could be recognized.

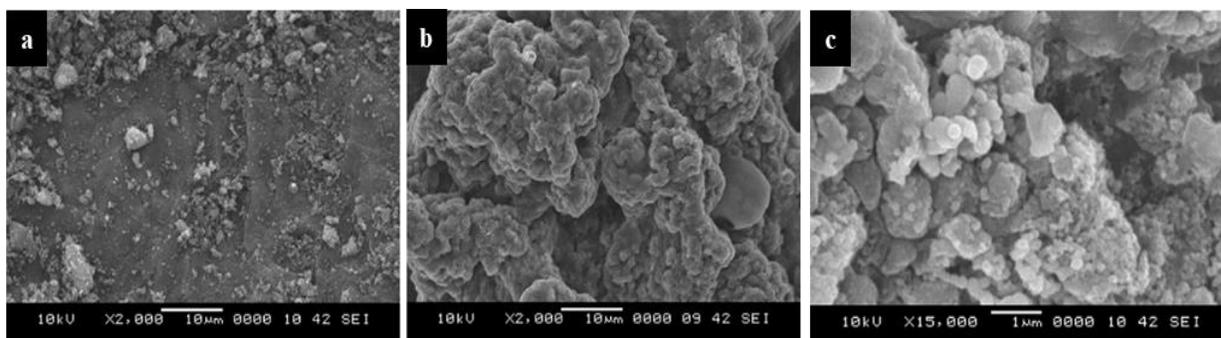


Fig. 27. SEM images for (a) raw GAC (b) GAC biofilm (c) bio-sludge microorganism (Adapted from [59]).

5.2.3 Waste materials

5.2.3.1 Inner tube of tires

The use of inner tube of tire as a biocarrier was explored. Biofilm was completely developed on the exterior of the media of the MB-aerobic-SBR with 1–2 mm thickness. The maximum biofilm volume on the surface was 17.18 ± 0.7 g/m². This is about 30% larger than that of the conventional aerobic-SBR following the equivalent organic loading conditions. The other advantage of the MB-aerobic-SBR was the sustainable stability of effluent quality as early as 2–3 days during operation [63]. However, it must be noted that when utilizing tires inner tube tires, caution need be exercised to hold the metal-bound chemicals normally received in the tires. This can be done by carrying out chemical analysis. also, the leakage of hazardous materials, particularly heavy metals, into the effluent must also be tracked, throughout the operation of the system.

5.2.3.2 Luffa Sponge (LS)

As earlier described in section 4.3.5, LS is a natural biocarrier material used to simply capture a biofilm with fortified microbes throughout the early days of wastewater treatment. It forms biofilm faster than most thermoplastic biocarriers. The fast LS biofilm formation can be associated with the following factors: (i) high SSA which is hydrophilic (ii) enmeshment role of the 3-D fiber structure of the sponge can assemble microbial strains promptly (iii) biological compatibility made microbes develop and breed fast. Investigating the use of LS by [67], it was discovered that, the LS biocarrier had higher density than that of water after the biofilm was developed, since it was packed with large volume of active sludge and biofilm within and on its covering.

In a related study using MBSBR-LS, higher reduction was noted in nitrate-N, which had a lesser relative abundance of *Pseudomonas* (0.3%). The external carbon source produced by the breakdown of luffa, which could have led to the denitrification, might just have accelerated this [66]. Thus, it can be concluded that LS fiber biocarrier had a limited life span and could be progressively deteriorated by microorganisms, without causing secondary pollution.

5.2.3.3 Iron shavings

Unlike the LS biocarrier, biofilm formation was not observed on iron shavings surface during the early days of SBBR operation. In fact, the surface turned black after operating for a long period of time. Surprisingly, after 7 days of service, a thin biofilm started to develop on the reactor's internal surface with gradual increase in thickness. Unfortunately, the biofilm does sweep away as iron delivered by corrosion somewhat suspend in the effluent reactor and slightly absorb to biofilm. When the level of iron in suspended solid was > 70%, the sludge operation was severely suppressed and could be restored by greatly reducing iron concentrate in the reactor [68]. This easily elucidates that iron shavings are not suitable for biofilm formation.

5.2.3.4 Palm oil clinker

An investigation carried out by [64], reported that POC attachment improves the surface contact area with influent wastewater for biomass. Hence, the efficacy of rehabilitation and removal increased. The study, however, note that, a decline of the MLSS content during the sampling times was experienced which could be because of the biomass washout and the stagnation of biomass inside the POC biocarrier.

1172 **5.2.3.5 Peach Pit**

1173 Smooth biofilm without pores and channels were fully developed on the coverings of peach pits used in fixed-bed
1174 sequencing batch reactor (FSBR). This implies that the biocarrier was covered with a suitable biofilm, that elevated
1175 organic matter oxidation and diffusion into the biofilm [9]. At high OLR, the biomass activity in the biofilm secures
1176 great treatment capability and operational steadiness of the FSBR process. This proves that the use of peach pits as a
1177 biocarrier medium can be efficacious in the management of increased nutrient loaded wastewater.

1178
1179 **5.2.3.6 WAS pellet**

1180 The introduction of WAS pellet into the reactor as a biocarrier to encourage microbial development was profoundly
1181 effective in sludge holding. Sludge units proved the co-existence and advancement of anammox bacteria, AOB and
1182 denitrifying bacteria in the reactor and biofilm production on to the biocarriers. The cumulative volume of biomass
1183 appended to the biocarriers signifies about 73% of the suspended biomass. This implies that the principal part of the
1184 biomass was fastened to the biocarrier as diverse classes of microorganisms immobilized and grew on to the biocarrier
1185 to produce biofilm [29]. Biocarrier saturation with fastened biomass and raw biomass continuously developed in the
1186 suspended stage proved that MLSS and MLVSS have substantially increased.

1187
1188 **5.2.4 Steel disk**

1189 Research team by [69] in their study explored the application of steel disc in an AnSBBR. They discovered that organic
1190 substrate and nitrates were just accessible in the exterior layers of the disk due to the significant thickness of the
1191 biofilm. Bacterial cells presumably decayed in deeper and older layers of the biofilm harboring no substrates, thus the
1192 discharged organic compounds could be provided to exterior layers comprising nitrates.

1193 In another article published by same authors [32], it was reported that biomass was mainly generated in the
1194 structure of a biofilm attached to fully submerged discs. The morphological structure of the biofilm reveals that biofilm
1195 thickness varied from 0.5-20 mm, because of the various impacts of abrasive forces. Despite portraying clear
1196 stratification, the film color varied from milk-like to dark brown. Bacteria of the biofilm in the submerged disk [70],
1197 manifested an improved intracellular polyphosphates growth. Also, the arrangement of the biofilm stimulated
1198 anaerobic-aerobic as well as anoxic-anaerobic and anaerobic conditions in a SBBR.

1199
1200 **5.2.5 Clay minerals**

1201 **5.2.5.1 Mineral coal (MC)**

1202 MC can be recognized as an effective inert supporter for biomass appendage, particularly for SRB and *methanogenic*
1203 *archaeae*. It is a general observation that operational control for any situation can be realized following a concise time
1204 span following the advances in the operational circumstances, thus symbolizing the great capability of the MC to
1205 preserve biomass.

1206 The experimental results for FB-SBBR during wastewater treatment with a high ammonia concentration
1207 demonstrated the potential of MC as a biocarrier. Authors [19], made this conclusion because of the high cumulative
1208 biofilm thickness (1954 μm) recorded, enabling the nitrification and denitrification to simultaneously befall. A study
1209 by [73] concluded that MC can be utilized for the simultaneous extraction of sulfate and organic matter provided
1210 ethanol is adopted as electron donor. However, at elevated doses, the development of TDS and COD has been noticed
1211 with increasing concentrations. This could be due to the immense concentrations of undissociated H_2S produced
1212 through the sulfate reduction process signifying that SRB were able to bind to the MC.

1213
1214 **5.2.5.2 Inert stone chips**

1215 Treating chemical wastewater by inert stone chips coupled SBBR, authors [20] revealed that , stone chips coated with
1216 biofilm show sparse greenish brown color. Microscopic studies of the self-immobilized biofilm apprehended further
1217 revealed: (i) protozoa, fungi and bacteria emergence (ii) that the morphology of the biofilm possess an uneven surface
1218 texture with excellent interior permeable channels and (iii) heterogeneous with distinct biomass packs intermittently
1219 installed covering the surface.

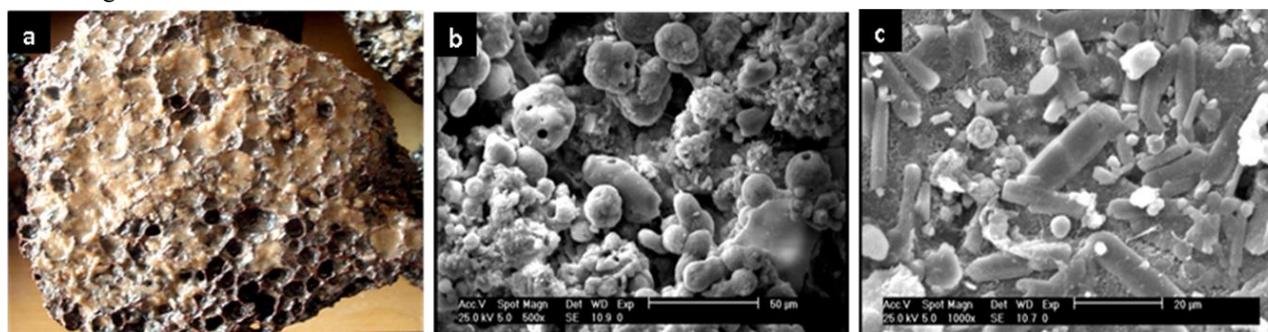
1220
1221 **5.2.5.3 Zeolite**

1222 Notwithstanding relating to varying alkalinity ratio, samples from natural zeolite (NZ) and synthetic zeolite (SZ) were
1223 alike. However, the population arrangement formed varied between the variable materials. The analysis proved that
1224 these modifications of the population arrangement were correlated with the ammonium adsorption potential of zeolite.

1225 The variability could be caused by the distinctive adsorption ability of the materials, which regulate different FA in
1226 solution and farther affect the nitrifying microorganism movements [72]. The most prevalent genus in the NSSBR and
1227 SSBRR was *Betaproteobacteria* with the average abundances of 43.19% and 54.72%, respectively.

1228 1229 **5.2.5.4 Pumice stone**

1230 In SBBR systems, SP presents well-matched anaerobic biomass, high substrate utilization rate and apparent first-order
1231 coefficient constant. However, it is not advisable to be utilized as a support material because of chemical structural
1232 damages. Authors [82], argued that, they realized a sudden boost of 82% in alkalinity when SP was used for
1233 immobilization. This increase could be clarified by the reaction of the SP with the substrate, which provided for
1234 carbonate synergies among them. Contrary to the behavior of SP, the prevalent bacterial strains in the biofilm
1235 cultivated on the scales of the volcanic pumice stones is spherical and rod-shaped bacteria, when used as a biocarrier
1236 in FB-SBBR1 [16]. The optical and SEM images of the biofilm formed on the packing material interfaces are presented
1237 in (Fig. 28a and 28b) where almost all the pores of the pumice stones used in FB-SBBR were masked by a panel of
1238 attached-growth biofilm.



1239
1240 **Fig. 28.** SEM and Optical photographs of the biofilm grown on surfaces of volcanic pumice stones in FB-SBBR
1241 (Adapted from [16]).
1242

1243 **5.2.6 Fibrous biocarriers**
1244 Fiber materials either flexible or semi-flexible serving as biocarriers possess huge SSA for microorganism appendage.
1245 Hence, bioreactors embedded with the fibers are able to treat high strength industrial wastewater because of the huge
1246 substrate and stock mass transfer rendered resulting from the fiber high SSA. They allow SBBR systems to withstand
1247 huge organic load and hydraulic shock loading because of synergistic associations between various microbial species
1248 developed in the biofilm [1, 10, 75].

1249 A research group by [83] used flexible fibers in a sequencing batch flexible fibre biofilm reactor (SBFFBR)
1250 system for the wastewater treatment. They recorded an enhanced quantity of biofilm volume in the reactor leading to
1251 the elevated biomass concentration in the reactor. The main reason for the elevated biomass concentration in the
1252 reactor discharge phase may be the higher SSA of the fiber (2200 m²/m³). Another possible reason could be, the
1253 attachment of bulk biomass to the fiber biocarrier. According to findings as depicted in Fig. 29a, by same authors in
1254 a following paper after a decade [78], biofilms covered the surface of individual fibres and filled the space between
1255 closely opposed fibres. Clumps of bacteria surrounding and closely adherent to the individual fibre are observable in
1256 Fig. 29b. In the outer surface of the biofilm, a variety of bacterial morphologies; rod, coccal, spiral, and
1257 helical morphologies with a mixture of straight- and comma-shape rods were observed as the most abundant organisms
1258 (Fig. 29c and d). Identified by their tail-like structure, flagella were seen in (Fig. 29c) as the availability of DO attracted
1259 grazing nematode species (Fig. 29d).

1260 An investigation focused on the viability of simultaneous nutrients removal by inserting a fibrous gas
1261 permeable membrane layer into a traditional SBRR. Authors [84] discovered that the technique permits long-term
1262 preservation of AOBs, allowing for the uninterrupted production of nitrite and nitrate that can be used by DNPAOs
1263 as electron acceptors soon after penetration from the biofilm into the bulk. However, the optimization of the membrane
1264 fibrous support, its cleaning, and the detection of DNPAOs linger as a problem.
1265

1266

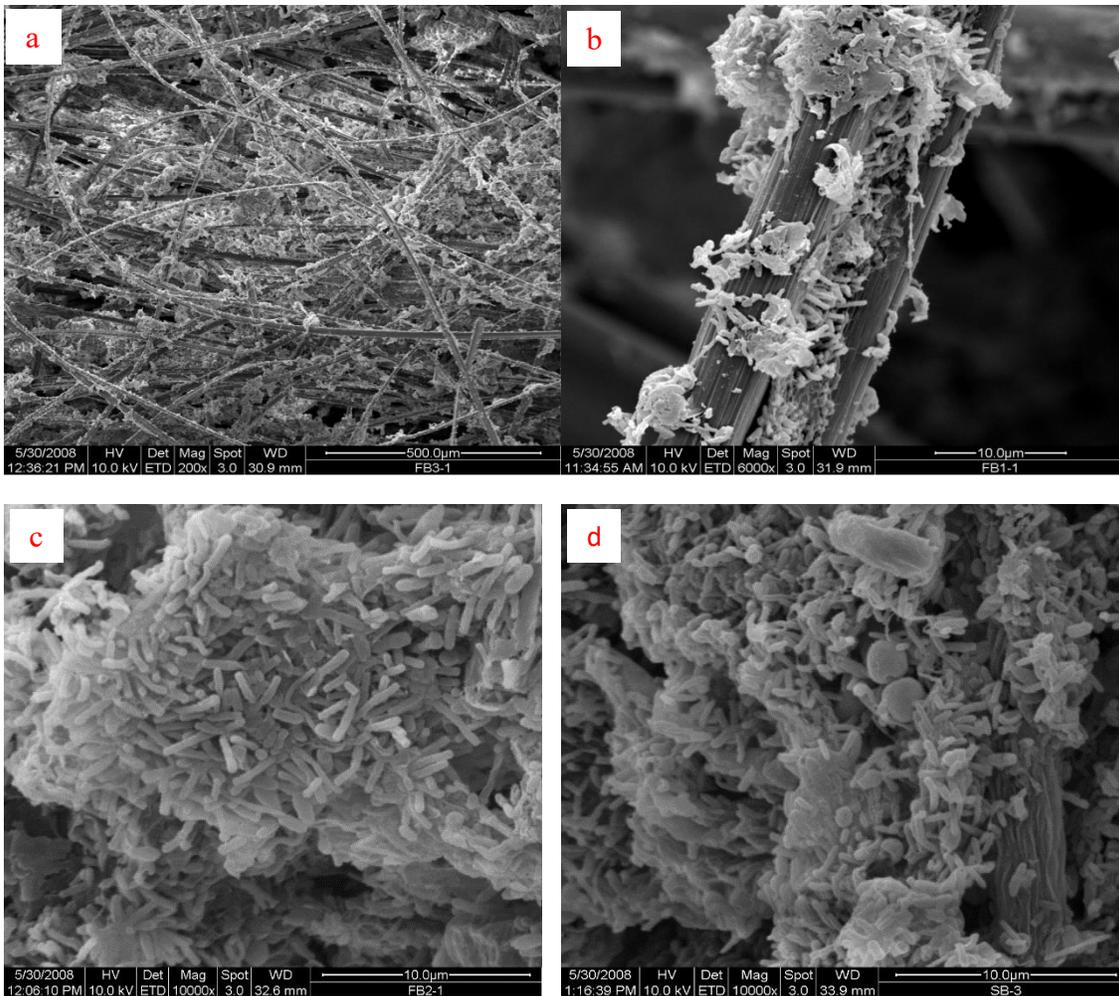


Fig. 29. SEM images of biofilm on flexible fibre with different magnifications of (a) 200x (b) 6000X (c and d) 10000x (Adapted from [78]).

1267

1268

1269

1270

1271

5.2.7 Others

1272

The analytical section “Others” collectively gathers materials which are not explicitly possible for insertion in a particular group. It consists of several articles reviewing various content sources.

1273

1274

1275

5.2.7.1 Vegetal carbon (VC)

1276

Similar to SP as earlier discussed in section 5.1.5.4, VC is not rated fit support for application in anaerobic digestion, since it allow for the appendage of unwanted organisms that struggle with methanogenic organisms for acetate substrate [82].

1277

1278

1279

1280

5.2.7.2 Rope-type media

1281

In an FbSBR, high cover and filamentous-type of BC^{PLUS} media has the influence of preserving a high quantity of biomass on its matrix rapidly ($\approx 7.7 \text{ g TS/m}^2$ media surface) with simultaneous nitrogen and organic removal. According to the authors [86], microorganisms were just found on the mold of media as biocatalyst in the reactor through the perpetual running phase.

1282

1283

1284

1285

1286

5.2.7.3 Nylon nets

1287

When NYLON nets were used as biocarriers, suspended and fixed biomass was made up of 20% NOB and 50% AOB. Optical microscopy of the reactor content showed the presence of metazoans and protozoans, suggesting a balanced treatment condition. A thick biofilm was found to be the fixed biomass and large numbers of *Rotatoria sp.*, *Rotifers*

1288

1289

1290 and *Zooglea sp* were seen to be included [87]. Based on the aforementioned facts, it is highly suggested for utilization
1291 as a biocarrier material.

1292

1293 **5.2.7.4 Eucalyptus charcoal**

1294 Methanogenic action appears to become the deciding factor to explain the greater effectiveness of the reactor packed
1295 with charcoal for organic matter extraction. At a COD/SO₄²⁻ ratio of 0.4, when the reactor was loaded with charcoal,
1296 only incomplete-oxidizing SRB metabolized basic organic matter with acetate, which is only transformed by
1297 sulfidogenic bacteria. This is the sole experimental situation under which SRB operation was the exclusive and
1298 dominant mechanism [79].

1299

1300 **5.2.7.5 Porous polymer**

1301 Porous polymers as biocarriers are fit for microorganism's immobilization. After the start-up of SBBR [52], biofilm
1302 was seen to have successfully covered the biocarrier. Bacteria in the outside layer of the biofilm were principally
1303 aerobic species. Organic matter adsorbed to the biofilm acted as electron donators. Aeration loop positioned in the
1304 intermediate cylinder of the novel SBBR, provided sufficient DO. Thus, nitrification governed the outside layer of the
1305 biofilm in this cylinder.

1306

1307 **5.2.7.6 Aluminum foil**

1308 Study by [88], showed that speedy initiation of SNAP and anammox process in biofilm reactor using aluminum foil
1309 without anammox sludge being inoculated is attainable. In their research, not only was biofilm produced to retain
1310 biomass and enrich AnAOB, tremendous parameters were properly monitored to suppress NOB proliferation, which
1311 helped establish an appropriate condition for AnAOB development, resulting to SNAP process rapid initiation. The
1312 microbial analysis inferred that microbial abundance of biomass improved while the microbial diversity decreased.
1313 *Candidatus Brocadia* and *Nitrosomonas* were enriched as the main AnAOB and AOB, respectively. *Nitrospira* was
1314 the main NOB and its relative bounty was reduced.

1315

1316 **5.2.8 Composite packing**

1317 **5.2.8.1 Composite of PE and inorganics (WD-F10-4 BioM™)**

1318 The self-floating biocarrier (WD-F10-4 BioM™) was competent to develop active biofilm in the system operated by
1319 [65]. The biomass concentration quantified at the completion of the experiment amount to 1,301 mg/L of biomass
1320 appended on the biocarriers and 3,721 mg/L for the suspended biomass, indicating that the biofilm has higher activity
1321 than sludge.

1322

1323 **5.2.8.2 GAC and natural zeolite**

1324 The effect of adding composite biocarriers (GAC and Natural zeolite) were studied. Even though, GAC and zeolite
1325 utilized in the research were alike in particle size, density and SSA. SEM photographs unveiled that, the GAC was
1326 extra permeable, had a buffeting surface with non-uniform structure than zeolite (Fig. 30). This could be the major
1327 reason why higher quantity of microbes were fastened to GAC than zeolite. Microbes willingly appended more to the
1328 fissures and uneven surfaces of the GAC biocarrier. However, the positive charge on zeolite aided the adherence of
1329 negatively charged microbes on to the covering of zeolite. Elongated rod shaped cells were fixed on to the exterior of
1330 both GAC and zeolite [71].

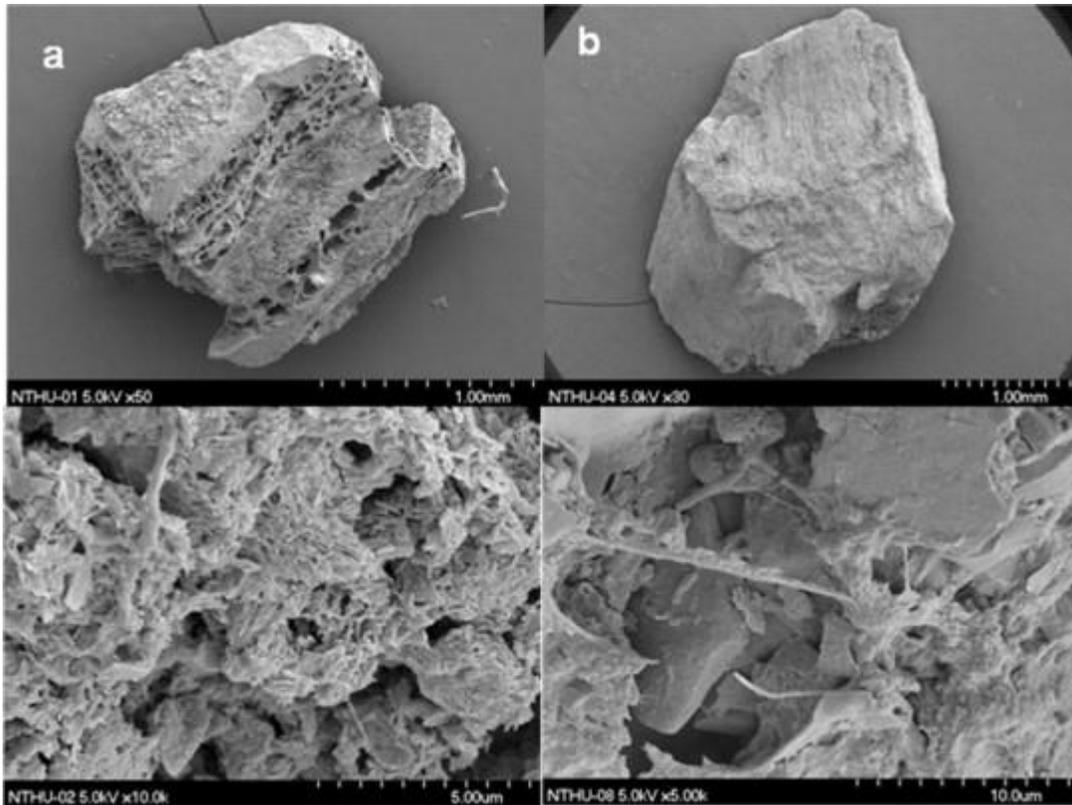


Fig 30. SEM photograph of (a) GAC external surface (top) and within (bottom) and (b) zeolite external surface (top) and within (bottom) (Adapted from [71]).

1331
1332
1333
1334
1335
1336
1337
1338
1339
1340
1341
1342
1343
1344
1345
1346
1347
1348
1349
1350
1351
1352
1353
1354
1355
1356

5.2.8.3 Floating spherical carriers (FSC)

The Floating spherical carriers (FSC) composite carrier created fluid isolation and was able to retrieve suspended matter that led to the liberation of dissolved organic matter by biofilm generation and biofilm lysis. The energy-uncoupling metabolism, slow-growth enrichment, sludge decay, hydrolytic and fermentation bacteria all led to the decline of in situ sludge in the floating spherical carriers-sequencing batch biofilm reactor (FSC-SBBRR). The FSC-SBBR generated 0.16 g MLSS/g COD, nearly 27.27% less sludge [85]. *Azospira* and *Mizugakiibacter* anaerobic effervescence bacterium were found to govern the FSC organisms, symbolising sludge decay as the principal mechanism on biofilm. Going by these findings, it can thus, be concluded that floating spherical carriers can adequately block swung sludge, enhancing biofilm production capability significantly.

5.2.8.4 Activated carbon fiber filler

SEM results indicates that a thick biofilm coated the filler surface, creating an anaerobic-aerobic microenvironment that facilitated nutrient removal. Dense biological membranes were also formed on the exterior of carbon fiber substances [76]. The SEM photographs of the biofilm further designate that the adhesion force of the interface is high with a real elastic force. Fig. 31B presents substantial amounts of cells in the structure of biofilms, sludge, and flocs. Fig. 31C and D, reveals that the exterior of the filler has a complicated structure mirroring the EPS molecules, which is liable for that in Fig. 31B. While Fig. 31D, presented a vast quantity of cocci, *Bacillus*, multi-streptococci and filamentous bacteria embedded in EPS framework, developing a thick biological network. This clearly indicates that, the activated carbon fiber filler demonstrated strong output in biofilm formation and microorganism immobilization. The major two parameters affecting biofilm growth are substrate loading rates and hydrodynamic conditions [25]. Consequently, various processes, including abrasion, erosion, sloughing and predator grazing, are said to be responsible for biofilm detachment.

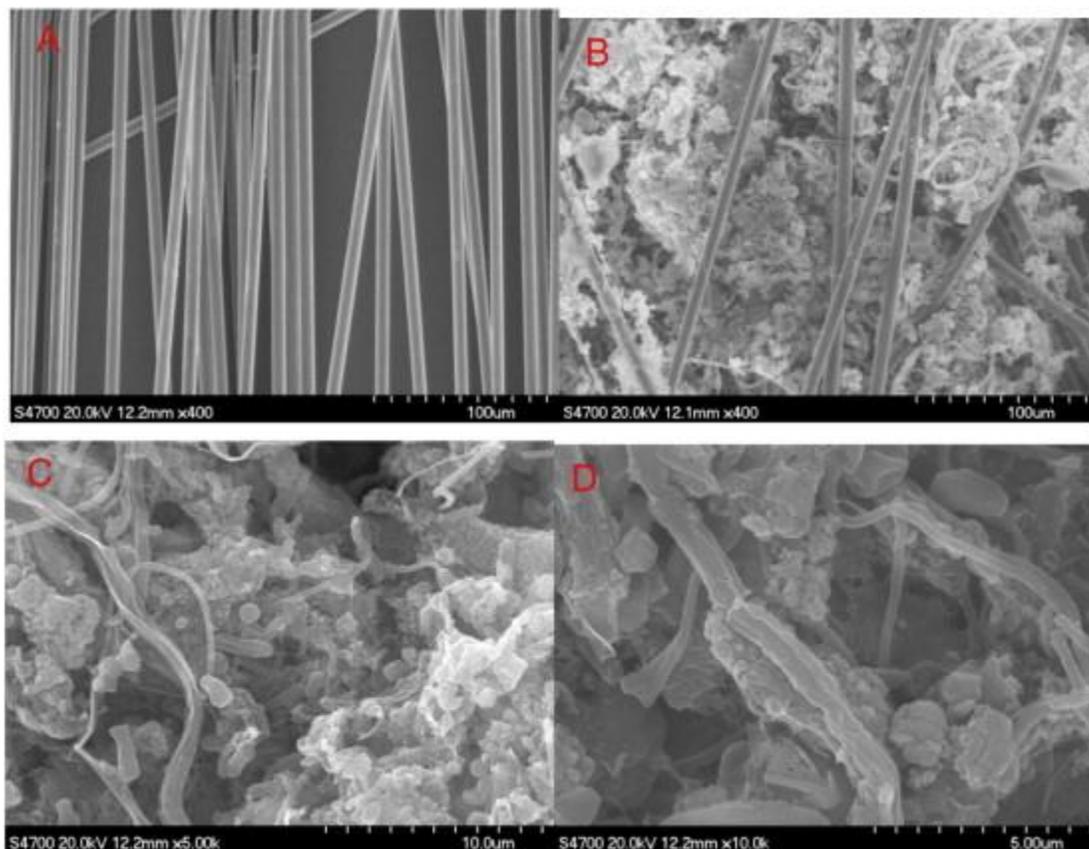


Fig. 31. SEM images of activated carbon fiber filler (A) and biofilm formed on the filler (B, C, and D) (Adapted from [76])

1357
1358
1359
1360
1361
1362
1363
1364
1365
1366
1367
1368
1369
1370
1371
1372
1373
1374
1375
1376
1377
1378
1379
1380
1381
1382
1383
1384

6.0 Conclusion

Numerous materials have been utilized as biocarriers in SBBR systems for simultaneous organic matter removal and biofilm formation during wastewater treatment, depending upon their availability and other specifications. Thus, showing different degrees of effectiveness. However, their extensive implementation is hindered by material property and relatively high operational and maintenance costs. This leads to the search for sustainable, environmentally friendly and economical compact biocarriers, which could retain sufficient treatment efficiency. Thus, this article presented a systematic review of existing research articles to advance our understanding of the properties and performance of various biocarriers with specific focus on their contributions in nutrient removal and biofilm formation. It further identified and discussed the limitations of these biocarriers, made comparative analyses and recommendations for better understanding.

The study, however, note the following:

- Biocompatibility with the predominant microorganism is very vital in selecting a biocarrier material.
- Waste materials offer a cost-effective, environment-friendly, and sustainable alternative as biocarriers for application in SBBR systems.
- Given the inherent disadvantages of synthetic biocarriers, it is recommended that natural materials with an additional incentive of reduced costs be used as an alternative biocarrier materials in SBBR for wastewater treatment.
- The advantage PE biocarriers have over other biocarrier materials is that biofilm can be distributed between its internal and external surfaces. Interestingly, PE can be pretreated to improve its physical properties. However, it does not encourage the predominance of any type of microorganism. Thermodynamic characterization indicated that PE had more favorable surface properties for the initial adhesion of biomass than PP. LS forms biofilm faster than most thermoplastic biocarriers.
- When PU foam was compared with some other biocarriers, it presented the fastest start-up and yielded constant effluent COD samples.

- 1385
- 1386
- 1387
- 1388
- 1389
- 1390
- 1391
- 1392
- SBBR systems developed with fibrous materials as biocarriers are highly efficient and energy saving wastewater treatment technologies.
 - Composite packing: could occupy less space, run normally, operate in a stable manner, resist impact load, deal with higher hydraulic load and achieve better purification.
 - PP biocarriers possess a longer life span than other thermoplastic materials providing better attachment than classical plastic biocarriers. Similarly, LS forms biofilm faster than most thermoplastic biocarriers while iron shavings are not suitable for biofilm formation

1393 7.0 Future directions

1394 In the recent years, biofilm technology is increasingly becoming largely employed for wastewater treatment under
1395 various loading and operational conditions, for effective organic and nutrient removal. Therefore, continued efforts
1396 are required in developing suitable biocarriers to address biofilm control issues. Researchers continue to concentrate
1397 on the improvement of biofilm technologies with special focus on biocarrier material. However, design and
1398 implementation of these technologies are hindered by the need for more appropriate biocarriers to account for the
1399 complexities of the heterogeneous communities that support substrates degradation. Hence, the following are
1400 suggested:

- 1401 i. **Developing new and/or modification of existing biocarriers:** In any functional biocarrier, biofilm bioactivity
1402 must be significantly higher than suspended biomass. Therefore, searching for a novel biocarrier that would give
1403 rise to higher biofilm activity is important. While searching for the new biocarriers, researchers may consider
1404 trying the following: (i) modifying the existing ones could also be an option such as, doping fibers on existing
1405 biocarrier materials or specially manufactured membranes for application in SBBR. (ii) application of plastic
1406 waste or fibers extracted from cellulose of by-products of major agro-based materials as biocarriers. This will
1407 also go a long way towards utilizing waste.
- 1408 ii. **Composite biocarriers:** Biocarriers despite their demonstrated effectiveness in treating wastewaters, have their
1409 own limitations and are yet to be investigated for large-scale applications. Therefore, future efforts may be
1410 focused on improving such biocarriers to reduce space and energy consumption as well as gas and odor
1411 emissions. Future researchers should further investigate the efficiency of composite biocarriers instead of stand-
1412 alone biocarriers. A thorough understanding of stand-alone biocarriers and its modification as a composite has
1413 a vast potential to be the future trend for use in wastewater treatment. More studies are required to be conducted
1414 in making composite biocarriers e.g. coating Banana trunk fiber with chitosan composite after durability and
1415 surface integrity test have been investigated.
- 1416 iii. **Biocarrier coating:** Commercial plastic media has limited surface area, and this has limitation to its use when
1417 high biofilm growth rate is required. The development of activated carbon on the surface of plastic biocarriers
1418 for biofilm system in order to provide a large surface area for microbes to attach would result in reduced retention
1419 time. Hence, making the treatment plant more compact and effective. It also retains suspended microorganisms
1420 within the bioreactor, thereby significantly reducing plant footprint and produces less sludge. More so, knowing
1421 fully that nanoporous carbons (NPCs) possesses highly porous structure and enlarged surface area, their
1422 utilization combined with plastic biocarriers would be remarkably valuable in adsorption and separation
1423 processes for pollutant degradation because of its high adsorptive performance. According to findings of this
1424 review, only GAC re-generated from coconut shells/husk has been used as a biocarrier in SBBR. Therefore,
1425 future researchers should conduct more studies using biomass materials like apricot shells, walnut shells,
1426 bamboo etc.
- 1427 iv. **3D printing:** The optimization of biocarriers application in SBBR systems in order to increase efficiency and
1428 reduce operating costs has not been deeply explored. Therefore, more research should be done to vehemently
1429 optimize various biocarrier materials for application in SBBR systems during the treatment of various
1430 wastewater types. During this review, it has been observed that most researchers focuses on the optimization of
1431 the environmental and operational parameters such as Temperature, salinity, HRT, SRT, cycle time etc, thus,
1432 forgetting to optimize biocarrier properties such as density, thickness, specific area, size, shapes, etc. The 3D
1433 printing technique using most recent and sophisticated machines would significantly help in optimizing the
1434 aforementioned biocarrier properties at minimal cost and in less time.

1435
1436

1437 **Acknowledgement**

1438 The study enjoyed the support of Universiti Teknologi PETRONAS (UTP), Malaysia.

1439 **Declaration of competing interest**

1440 The authors declare that they have no known competing financial interests or personal relationships that could have
1441 appeared to influence the work reported in this paper.

1442

1443 **Abbreviation**

1444	AS:	Activated Sludge
1445	AOB:	Ammonia Oxidizing Bacteria
1446	AOX:	Adsorbable Organic Halides
1447	APSBRR:	Alternating Pumped Sequencing Batch Biofilm Reactor
1448	ASBRR :	Anaerobic Sequencing Batch Biofilm Reactor
1449	CANON:	Completely Autotrophic Nitrogen Removal Over Nitrite
1450	COD:	Chemical Oxygen Demand
1451	DEAMOX:	Denitrifying Ammonium Oxidation
1452	DNPAOs:	Denitrifying Phosphate Accumulating Organisms
1453	EPS:	Extracellular polymeric substances
1454	FB-SBRR:	Fixed-Bed Sequencing Batch Biofilm Reactor
1455	FSBR:	Fixed-Bed Sequencing Batch Reactor
1456	FSC-SBRR:	Floating Spherical Carriers-Sequencing Batch Biofilm Reactor
1457	FbSBR:	Fixed-bed Biofilm Sequencing Batch Reactor
1458	HG-SBR:	Hybrid-Growth Sequencing Batch Reactor
1459	HRT:	Hydraulic Retention Time
1460	HSBR:	Hybrid Sequencing Batch Reactor
1461	IFAS-SBR:	Integrated fixed-film Activated Sludge-Sequencing Batch Reactor
1462	GAC:	Granular Activated Carbon
1463	GAC-SBRR:	Granular Activated Carbon Sequencing Batch Biofilm Reactor
1464	LS:	Luffa Sponge
1465	MB-aerobic-SBR:	Moving biofilm aerobic Sequencing Batch Reactor
1466	MBBSBR:	Moving-Bed Biofilm Sequencing Batch Reactor
1467	MBSBRR:	Moving Bed Sequencing Batch Biofilm Reactor
1468	MC:	Mineral Coal
1469	MCRT:	Mean Cellular Retention Times
1470	MLSS:	Mixed Liquor Suspended Solid
1471	MLVSS:	Mixed Liquor Volatile Suspended Solid
1472	MM-SBRR:	Multimedia-Sequencing Batch Biofilm Reactor
1473	MSBR:	Sequencing Batch Reactor Biofilm
1474	NLR:	Nitrogen Loading Rate
1475	NOB:	Nitrite Oxidizing Bacteria
1476	NSBRR:	Natural Zeolite Sequencing Batch Biofilm Reactor
1477	PAOs:	Phosphate Accumulating Organisms
1478	POC:	Palm Oil Clinker
1479	PN:	Partial Nitrification
1480	PU:	Polyurethane
1481	PE:	Polyethylene
1482	PEHD:	Polyethylene high-density
1483	PP:	Polypropylene
1484	PVC:	Polyvinyl Chloride
1485	SBR:	Sequencing Batch Reactor
1486	SBRR:	Sequencing Batch Biofilm Reactor
1487	SBRR-VFCW:	Sequencing Batch Biofilm Reactor combined with a Vertical Flow Constructed Wetland
1488	SBCR:	GAC-Biofilm Configured Sequencing Batch Reactor
1489	SBFFBR:	Sequencing Batch Flexible Fibre Biofilm Reactor
1490	SBMBfR:	Sequencing Batch Membrane Biofilm Reactor
1491	SBGR:	GAC-Biofilm Sequencing Batch Reactor

1492	SBMBBR:	Sequencing Batch Moving Bed Biofilm Reactor
1493	SBMABR:	Sequencing Batch Membrane-Aerated Biofilm Reactor
1494	SBR-BF:	Sequencing Batch Reactor-Biofilm
1495	SC-SBBR:	Suspended Carriers-Sequencing Batch Biofilm Reactor
1496	SEM:	Scanning Electron Microscope
1497	SLR:	Systematic Literature Review
1498	SNAD:	Simultaneous Nitrification and Denitrification
1499	SNDPR:	Simultaneous Nitrification and Denitrification and Phosphorous Removal
1500	SRB:	Sulphate-Reducing Bacteria
1501	SRT:	Solid Retention Time
1502	SSA:	Specific Surface Area
1503	SS:	Suspended Solids
1504	SSBBR:	Synthetic Zeolite Sequencing Batch Biofilm Reactor
1505	SP:	Synthetic Pumice
1506	TN:	Total Nitrogen
1507	TP:	Total Phosphorus
1508	TVS:	Total Volatile Solids
1509	WAS:	Waste Activated Sludge
1510	VC:	Vegetal Carbon

1511

1512 **References**

- 1513 1. Xiang, Y., et al., *Effect of Biofilm Density on Nitrous Oxide Emissions and Treatment Efficiency on*
1514 *Sequencing Batch Biofilm Reactor*. Water Air and Soil Pollution, 2016. **227**(9).
- 1515 2. Zou, J.T., et al., *Enhancing nitrogen removal from low carbon to nitrogen ratio wastewater by*
1516 *using a novel sequencing batch biofilm reactor*. Journal of Environmental Sciences, 2016. **50**: p.
1517 32-37.
- 1518 3. Hajsardar, M., et al., *NITROGEN REMOVAL FROM AMMONIUM-RICH PHARMACEUTICAL*
1519 *WASTEWATER. A COMPARISON BETWEEN SEQUENCING BATCH REACTOR (SBR) AND*
1520 *SEQUENCING BATCH BIOFILM REACTOR (SBBR)*. Environment Protection Engineering, 2018.
1521 **44**(3): p. 95-115.
- 1522 4. Ong, C., K. Lee, and Y.J.J.o.W.P.E. Chang, *Biodegradation of mono azo dye-Reactive Orange 16 by*
1523 *acclimatizing biomass systems under an integrated anoxic-aerobic REACT sequencing batch*
1524 *moving bed biofilm reactor*. 2020. **36**: p. 101268.
- 1525 5. Ng, J., et al. *Organic and nutrient removal for domestic wastewater treatment using bench-scale*
1526 *sequencing batch reactor*. in *AIP Conference Proceedings*. 2021. AIP Publishing LLC.
- 1527 6. Mojiri, A., et al., *Pollutants removal from synthetic wastewater by the combined electrochemical,*
1528 *adsorption and sequencing batch reactor (SBR)*. Ecotoxicology and Environmental Safety, 2018.
1529 **161**: p. 137-144.
- 1530 7. Jagaba, A., et al., *Sequencing batch reactor technology for landfill leachate treatment: A state-of-*
1531 *the-art review*. 2021. **282**: p. 111946.
- 1532 8. Sirianuntapiboon, S., N. Jeeyachok, and R. Larplai, *Sequencing batch reactor biofilm system for*
1533 *treatment of milk industry wastewater*. Journal of Environmental Management, 2005. **76**(2): p.
1534 177-183.
- 1535 9. Soltani, R.D.C., et al., *Organic matter removal under high loads in a fixed-bed sequencing batch*
1536 *reactor with peach pit as carrier*. Environmental Progress & Sustainable Energy, 2013. **32**(3): p.
1537 681-687.
- 1538 10. Chai, H.X. and W. Kang, *Influence of Biofilm Density on Anaerobic Sequencing Batch Biofilm*
1539 *Reactor Treating Mustard Tuber Wastewater*. Applied Biochemistry and Biotechnology, 2012.
1540 **168**(6): p. 1664-1671.
- 1541 11. Arabgol, R., et al., *The impact of biofilm thickness-restraint and carrier type on attached growth*
1542 *system performance, solids characteristics and settleability*. 2020. **6**(10): p. 2843-2855.

- 1543 12. Shore, J.L., et al., *Application of a moving bed biofilm reactor for tertiary ammonia treatment in*
1544 *high temperature industrial wastewater*. 2012. **112**: p. 51-60.
- 1545 13. Guo, X., et al., *Performance and bacterial community of moving bed biofilm reactors with various*
1546 *biocarriers treating primary wastewater effluent with a low organic strength and low C/N ratio*.
1547 2019. **287**: p. 121424.
- 1548 14. Guo, Y.M., et al., *An integrated treatment of domestic wastewater using sequencing batch*
1549 *biofilm reactor combined with vertical flow constructed wetland and its artificial neural network*
1550 *simulation study*. Ecological Engineering, 2014. **64**: p. 18-26.
- 1551 15. Bakar, S.N.H.A., et al., *Performance of a laboratory-scale moving bed biofilm reactor (MBBR) and*
1552 *its microbial diversity in palm oil mill effluent (POME) treatment*. 2020. **142**: p. 325-335.
- 1553 16. Koupaie, E.H., M.R.A. Moghaddam, and S.H. Hashemi, *Evaluation of integrated*
1554 *anaerobic/aerobic fixed-bed sequencing batch biofilm reactor for decolorization and*
1555 *biodegradation of azo dye Acid Red 18: Comparison of using two types of packing media*.
1556 Bioresource Technology, 2013. **127**: p. 415-421.
- 1557 17. Ozturk, A., A. Aygun, and B. Nas, *Application of sequencing batch biofilm reactor (SBBR) in dairy*
1558 *wastewater treatment*. Korean Journal of Chemical Engineering, 2019. **36**(2): p. 248-254.
- 1559 18. Wei, D., et al., *Nitrogen removal via nitrite in a partial nitrification sequencing batch biofilm*
1560 *reactor treating high strength ammonia wastewater and its greenhouse gas emission*.
1561 Bioresource Technology, 2017. **230**: p. 49-55.
- 1562 19. Sarti, A., et al., *A new device to select carriers for biomass immobilization and application in an*
1563 *aerobic/anaerobic fixed-bed sequencing batch biofilm reactor for nitrogen removal*. Water
1564 Science and Technology, 2016. **74**(11): p. 2666-2674.
- 1565 20. Mohan, S.V., N.C. Rao, and P.N. Sarma, *Low-biodegradable composite chemical wastewater*
1566 *treatment by biofilm configured sequencing batch reactor (SBBR)*. Journal of Hazardous
1567 Materials, 2007. **144**(1-2): p. 108-117.
- 1568 21. Yin, J., et al., *Simultaneous biological nitrogen and phosphorus removal with a sequencing batch*
1569 *reactor-biofilm system*. International Biodeterioration & Biodegradation, 2015. **103**: p. 221-226.
- 1570 22. Mohan, S.V., et al., *Influence of recirculation on the performance of anaerobic sequencing batch*
1571 *biofilm reactor (AnSBBR) treating hypersaline composite chemical wastewater*. 2007. **98**(7): p.
1572 1373-1379.
- 1573 23. Krsmanovic, M., et al., *Hydrodynamics and surface properties influence biofilm proliferation*.
1574 2020: p. 102336.
- 1575 24. Hibiya, K., et al., *Simple prediction of oxygen penetration depth in biofilms for wastewater*
1576 *treatment*. 2004. **19**(1): p. 61-68.
- 1577 25. Zhan, X.M., M. Rodgers, and E. O'Reilly, *Biofilm growth and characteristics in an alternating*
1578 *pumped sequencing batch biofilm reactor (APSBBR)*. Water Research, 2006. **40**(4): p. 817-825.
- 1579 26. Shao, Y.X., et al., *Wastewater ammonia removal using an integrated fixed-film activated sludge-*
1580 *sequencing batch biofilm reactor (IFAS-SBR): Comparison of suspended flocs and attached*
1581 *biofilm*. International Biodeterioration & Biodegradation, 2017. **116**: p. 38-47.
- 1582 27. Ding, X.W., et al., *Biological denitrification in an anoxic sequencing batch biofilm reactor:*
1583 *Performance evaluation, nitrous oxide emission and microbial community*. Bioresource
1584 Technology, 2019. **285**.
- 1585 28. Zhao, Y., et al., *Influence of carrier filling ratio on the advanced nitrogen removal from*
1586 *wastewater treatment plant effluent by Denitrifying MBBR*. 2019. **16**(18): p. 3244.
- 1587 29. Daverey, A., et al., *Start-up of simultaneous partial nitrification, anammox and denitrification*
1588 *(SNAD) process in sequencing batch biofilm reactor using novel biomass carriers*. Bioresource
1589 Technology, 2015. **190**: p. 480-486.

- 1590 30. Maslon, A. and J.A. Tomaszek, *A study on the use of the BioBall (R) as a biofilm carrier in a*
1591 *sequencing batch reactor*. *Bioresource Technology*, 2015. **196**: p. 577-585.
- 1592 31. Wang, J.F., et al., *Estimation of spatial distribution of quorum sensing signaling in sequencing*
1593 *batch biofilm reactor (SBBR) biofilms*. *Science of the Total Environment*, 2018. **612**: p. 405-414.
- 1594 32. Mielcarek, A., et al., *Effect of the C:N:P ratio on the denitrifying dephosphatation in a sequencing*
1595 *batch biofilm reactor (SBBR)*. *Journal of Environmental Sciences*, 2015. **38**: p. 119-125.
- 1596 33. Zhang, H.Y., et al., *Mechanisms and characteristics of biofilm formation via novel DEAMOX*
1597 *system based on sequencing biofilm batch reactor*. *Journal of Bioscience and Bioengineering*,
1598 2019. **127**(2): p. 206-212.
- 1599 34. Adam, A.A., et al., *State of the Art and New Directions on Electrospun Lignin/Cellulose*
1600 *Nanofibers for Supercapacitor Application: A Systematic Literature Review*. 2020. **12**(12): p.
1601 2884.
- 1602 35. Yashni, G., et al., *Conventional and advanced treatment technologies for palm oil mill effluents: a*
1603 *systematic literature review*. 2020: p. 1-19.
- 1604 36. Al-Gheethi, A., et al., *Novel Coronavirus (2019-nCoV) outbreak; a systematic review for published*
1605 *papers*. 2020.
- 1606 37. Tan, C., et al., *Effects of tourmaline on nitrogen removal performance and biofilm structures in*
1607 *the sequencing batch biofilm reactor*. *Journal of Environmental Sciences*, 2018. **67**: p. 127-135.
- 1608 38. Sarti, A., et al., *The performance of an anaerobic sequencing batch biofilm reactor treating*
1609 *domestic sewage colonized by anoxygenic phototrophic bacteria*. *Chemosphere*, 2006. **62**(9): p.
1610 1437-1443.
- 1611 39. Lim, J.W., et al., *Nitrogen removal in moving bed sequencing batch reactor using polyurethane*
1612 *foam cubes of various sizes as carrier materials*. *Bioresource Technology*, 2011. **102**(21): p. 9876-
1613 9883.
- 1614 40. Cao, Y.F., et al., *The effect of dissolved oxygen concentration (DO) on oxygen diffusion and*
1615 *bacterial community structure in moving bed sequencing batch reactor (MBSBR)*. *Water*
1616 *Research*, 2017. **108**: p. 86-94.
- 1617 41. Lim, J.W., et al., *Simultaneous 4-chlorophenol and nitrogen removal in moving bed sequencing*
1618 *batch reactors packed with polyurethane foam cubes of various sizes*. *Bioresource Technology*,
1619 2013. **129**: p. 485-494.
- 1620 42. Pinho, S.C., et al., *Influence of bioparticle size on the degradation of partially soluble wastewater*
1621 *in an anaerobic sequencing batch biofilm reactor (ASBBR)*. *Process Biochemistry*, 2005. **40**(10): p.
1622 3206-3212.
- 1623 43. Ribas, M.M.F., et al., *Microbial Succession Within an Anaerobic Sequencing Batch Biofilm*
1624 *Reactor (ASBBR) Treating Cane Vinasse at 55 degrees C*. *Brazilian Archives of Biology and*
1625 *Technology*, 2009. **52**(4): p. 1027-1036.
- 1626 44. Zhang, J.H., et al., *Feasibility of in situ enriching anammox bacteria in a sequencing batch biofilm*
1627 *reactor (SBBR) for enhancing nitrogen removal of real domestic wastewater*. *Chemical*
1628 *Engineering Journal*, 2018. **352**: p. 847-854.
- 1629 45. Koupaie, E.H., M.R.A. Moghaddam, and S.H. Hashemi, *Investigation of decolorization kinetics*
1630 *and biodegradation of azo dye Acid Red 18 using sequential process of anaerobic sequencing*
1631 *batch reactor/moving bed sequencing batch biofilm reactor*. *International Biodeterioration &*
1632 *Biodegradation*, 2012. **71**: p. 43-49.
- 1633 46. Nogueira, R., et al., *Synthesis and degradation of poly-beta-hydroxybutyrate in a sequencing*
1634 *batch biofilm reactor*. *Bioresource Technology*, 2009. **100**(7): p. 2106-2110.
- 1635 47. Di Iaconi, C., et al., *Hydraulic shear stress calculation in a sequencing batch biofilm reactor with*
1636 *granular biomass*. *Environmental Science & Technology*, 2005. **39**(3): p. 889-894.

- 1637 48. Hajsardar, M., et al., *Improving Wastewater Nitrogen Removal and Reducing Effluent NO_x-N by*
1638 *an Oxygen-Limited Process Consisting of a Sequencing Batch Reactor and a Sequencing Batch*
1639 *Biofilm Reactor*. International Journal of Chemical Reactor Engineering, 2019. **17**(7).
- 1640 49. Matos, M., et al., *Sequencing batch biofilm reactor: from support design to reactor operation*.
1641 Environmental Technology, 2011. **32**(10): p. 1121-1129.
- 1642 50. Sun, L.Q., et al., *Enhanced biological nitrogen and phosphorus removal using sequencing batch*
1643 *membrane-aerated biofilm reactor*. Chemical Engineering Science, 2015. **135**: p. 559-565.
- 1644 51. Prendergast, J., M. Rodgers, and M.G. Healy, *The efficiency of a sequencing batch biofilm reactor*
1645 *in organic carbon and phosphorus removal*. Journal of Environmental Science and Health Part a-
1646 Toxic/Hazardous Substances & Environmental Engineering, 2005. **40**(8): p. 1619-1626.
- 1647 52. Xiao, H., et al., *Nitrogen removal from livestock and poultry breeding wastewaters using a novel*
1648 *sequencing batch biofilm reactor*. Water Science and Technology, 2010. **62**(11): p. 2599-2606.
- 1649 53. Muhamad, M.H., S.R.S. Abdullah, and H. Abu Hasan, *Efficiency of Attached-Growth Sequencing*
1650 *Batch Reactor in the Treatment of Recycled Paper Mill Wastewater*. Jurnal Teknologi, 2015.
1651 **74**(3): p. 89-94.
- 1652 54. Muhamad, M.H., et al., *Effects of pentachlorophenol load on PCP, COD and NH₃-N removal in*
1653 *lab-scale multimedia-sequencing batch biofilm reactor treating recycled paper mill wastewater*.
1654 Journal of Environmental Biology, 2019. **40**(3): p. 556-562.
- 1655 55. Muhamad, M.H., et al., *Application of response surface methodology (RSM) for optimisation of*
1656 *COD, NH₃-N and 2,4-DCP removal from recycled paper wastewater in a pilot-scale granular*
1657 *activated carbon sequencing batch biofilm reactor (GAC-SBBR)*. Journal of Environmental
1658 Management, 2013. **121**: p. 179-190.
- 1659 56. Muhamad, M.H., et al., *Performance evaluation of a granular activated carbon-sequencing batch*
1660 *biofilm reactor pilot plant system used in treating real wastewater from recycled paper industry*.
1661 Environmental Technology, 2012. **33**(8): p. 915-926.
- 1662 57. Abd-Rahman, R., et al., *Heavy Metal Biosorption Efficiencies of Expanded Bed Biofilm Reactor*
1663 *and Sequencing Batch Biofilm Reactor*. Asian Journal of Chemistry, 2013. **25**(13): p. 7193-7198.
- 1664 58. Rao, N.C., et al., *Treatment of composite chemical wastewater by aerobic GAC-biofilm*
1665 *sequencing batch reactor (SBGR)*. Journal of Hazardous Materials, 2005. **124**(1-3): p. 59-67.
- 1666 59. Yusoff, N., et al., *Evaluation of biodegradation process: Comparative study between suspended*
1667 *and hybrid microorganism growth system in sequencing batch reactor (SBR) for removal of*
1668 *phenol*. Biochemical Engineering Journal, 2016. **115**: p. 14-22.
- 1669 60. Ong, S.A., et al., *Granular activated carbon-biofilm configured sequencing batch reactor*
1670 *treatment of CI Acid Orange 7*. Dyes and Pigments, 2008. **76**(1): p. 142-146.
- 1671 61. Ong, S.A., et al., *Mineralization of Methyl Orange-containing wastewater by integrated*
1672 *anaerobic and aerobic processes using spent granular activated carbon-biofilm under sequencing*
1673 *batch reactor operation*. Desalination and Water Treatment, 2013. **51**(13-15): p. 2813-2819.
- 1674 62. Ong, S.A., et al., *Performance and Kinetic Study on Bioremediation of Diazo Dye (Reactive Black*
1675 *5) in Wastewater Using Spent GAC-Biofilm Sequencing Batch Reactor*. Water Air and Soil
1676 Pollution, 2012. **223**(4): p. 1615-1623.
- 1677 63. Sirianuntapiboon, S. and S. Yommee, *Application of a new type of moving bio-film in aerobic*
1678 *sequencing batch reactor (aerobic-SBR)*. Journal of Environmental Management, 2006. **78**(2): p.
1679 149-156.
- 1680 64. Kuan, Y.Z., S.R.M. Kutty, and A.A.S. Ghaleb, *Kinetics Coefficient of Palm Oil Clinker Media for an*
1681 *Attached Growth Media in Sequencing Batch Reactor Mode*. Journal of Ecological Engineering,
1682 2019. **20**(9): p. 18-27.
- 1683 65. Jing, J.Y., et al., *Removal of COD from coking-plant wastewater in the moving-bed biofilm*
1684 *sequencing batch reactor*. Korean Journal of Chemical Engineering, 2009. **26**(2): p. 564-568.

- 1685 66. Li, W.J., et al., *Nitrogen Removal in Moving Bed Sequencing Batch Reactors with Polyurethane*
1686 *Foam Cube and Luffa Sponge Carrier Materials*. Journal of Environmental Engineering, 2019.
1687 **145**(6).
- 1688 67. Wang, Y.Z., et al., *Comparison of Luffa Cylindrical Sponge and Plastic Sponge as Carriers in*
1689 *Sequencing Batch Biofilm Reactor for Sewage Treatment*, in *Advances in Environmental Science*
1690 *and Engineering, Pts 1-6*, R. Iranpour, et al., Editors. 2012. p. 2431-2438.
- 1691 68. Wang, Z., H.W. Wang, and L.M. Ma, *Iron shavings supported biological denitrification in*
1692 *sequencing batch reactor*. Desalination and Water Treatment, 2012. **49**(1-3): p. 95-105.
- 1693 69. Mielcarek, A., et al., *Denitrification aided by waste beer in anaerobic sequencing batch biofilm*
1694 *reactor (AnSBBR)*. Ecological Engineering, 2016. **95**: p. 384-389.
- 1695 70. Mielcarek, A., et al., *Effect of Acetic Acid on Denitrification and Dephosphatation Process*
1696 *Efficiencies in Sequencing Batch Biofilm Reactor*. Journal of Ecological Engineering, 2018. **19**(4):
1697 p. 176-180.
- 1698 71. Dutta, K., et al., *Effect of carriers on the performance of anaerobic sequencing batch biofilm*
1699 *reactor treating synthetic municipal wastewater*. International Biodeterioration &
1700 Biodegradation, 2014. **95**: p. 84-88.
- 1701 72. Chen, J., et al., *Response of nitrification performance and microbial community structure in*
1702 *sequencing biofilm batch reactors filled with different zeolite and alkalinity ratio*. Bioresource
1703 Technology, 2019. **273**: p. 487-495.
- 1704 73. Sarti, A., et al., *The treatment of sulfate-rich wastewater using an anaerobic sequencing batch*
1705 *biofilm pilot-scale reactor*. Desalination, 2009. **249**(1): p. 241-246.
- 1706 74. Tang, C.C., et al., *Enhanced nitrogen and phosphorus removal from domestic wastewater via*
1707 *algae-assisted sequencing batch biofilm reactor*. Bioresource Technology, 2018. **250**: p. 185-190.
- 1708 75. Jin, Y.X. and J. Yao, *Biological Nutrient Removal from Simulated Domestic Wastewater in*
1709 *Sequencing Batch Biofilm Reactor*, in *Advances in Environmental Science and Engineering, Pts 1-*
1710 *6*, R. Iranpour, et al., Editors. 2012. p. 2406-2409.
- 1711 76. Hai, R.T., et al., *Simultaneous removal of nitrogen and phosphorus from swine wastewater in a*
1712 *sequencing batch biofilm reactor*. Chinese Journal of Chemical Engineering, 2015. **23**(1): p. 303-
1713 308.
- 1714 77. Hu, J., et al., *Use of a sequencing batch reactor for nitrogen and phosphorus removal from*
1715 *municipal wastewater*. 2005. **131**(5): p. 734-744.
- 1716 78. Abdulgader, M., et al., *Performance and kinetics analysis of an aerobic sequencing batch flexible*
1717 *fibre biofilm reactor for milk processing wastewater treatment*. Journal of Environmental
1718 Management, 2020. **255**.
- 1719 79. Silva, A.J., et al., *KINETIC MODELING AND MICROBIAL ASSESSMENT BY FLUORESCENT IN SITU*
1720 *HYBRIDIZATION IN ANAEROBIC SEQUENCING BATCH BIOFILM REACTORS TREATING SULFATE-*
1721 *RICH WASTEWATER*. Brazilian Journal of Chemical Engineering, 2011. **28**(2): p. 209-219.
- 1722 80. Goh, C.P., et al., *Performance of sequencing batch biofilm and sequencing batch reactors in*
1723 *simultaneous p-nitrophenol and nitrogen removal*. Environmental Technology, 2009. **30**(7): p.
1724 725-736.
- 1725 81. Valdivia, A., et al., *Biological nitrogen removal with three different SBBR*. 2007. **55**(7): p. 245-254.
- 1726 82. Garcia, M.L., et al., *Effects of bed materials on the performance of an anaerobic sequencing*
1727 *batch biofilm reactor treating domestic sewage*. Journal of Environmental Management, 2008.
1728 **88**(4): p. 1471-1477.
- 1729 83. Abdulgader, M., et al., *Biological treatment of milk processing wastewater in a sequencing batch*
1730 *flexible fibre biofilm reactor*. Asia-Pacific Journal of Chemical Engineering, 2009. **4**(5): p. 698-703.

- 1731 84. Terada, A., et al., *Sequencing batch membrane biofilm reactor for simultaneous nitrogen and*
1732 *phosphorus removal: Novel application of membrane-aerated biofilm*. Biotechnology and
1733 Bioengineering, 2006. **94**(4): p. 730-739.
- 1734 85. Wang, Y.L., et al., *Investigate of in situ sludge reduction in sequencing batch biofilm reactor:*
1735 *Performances, mechanisms and comparison of different carriers*. Frontiers of Environmental
1736 Science & Engineering, 2018. **12**(5).
- 1737 86. Tran, H.T., et al., *A study on start-up operation of fixed-bed Biofilm Sequencing batch reactor*
1738 *(FbSBR) for piggery wastewater treatment*. Journal of Industrial and Engineering Chemistry,
1739 2007. **13**(6): p. 985-991.
- 1740 87. Costa, R.H.R., D.B. Wolff, and V.S. Souto, *Performance and Kinetics Aspects of Nitrogen*
1741 *Removal in a Biofilm Sequencing Batch Reactor*. International Journal of Environmental
1742 Research, 2013. **7**(2): p. 513-522.
- 1743 88. Cai, F.R., L.R. Lei, and Y.M. Li, *Rapid start-up of single-stage nitrogen removal using anammox*
1744 *and partial nitrification (SNAP) process in a sequencing batch biofilm reactor (SBBR) inoculated*
1745 *with conventional activated sludge*. International Biodeterioration & Biodegradation, 2020. **147**.
- 1746 89. Eker, S., F.J.E. Kargi, and m. technology, *Kinetic modeling and parameter estimation in biological*
1747 *treatment of 2, 4-dichlorophenol containing wastewater using rotating perforated tubes biofilm*
1748 *reactor*. 2006. **38**(6): p. 860-866.
- 1749 90. Zheng, D., et al., *Performance comparison of biofilm and suspended sludge from a sequencing*
1750 *batch biofilm reactor treating mariculture wastewater under oxytetracycline stress*.
1751 Environmental Technology, 2016. **37**(18): p. 2391-2404.
- 1752 91. Zheng, D., et al., *Performance evaluation and microbial community of a sequencing batch biofilm*
1753 *reactor (SBBR) treating mariculture wastewater at different chlortetracycline concentrations*.
1754 Journal of Environmental Management, 2016. **182**: p. 496-504.
- 1755 92. Li, Z.W., et al., *Impact of sulfadiazine on performance and microbial community of a sequencing*
1756 *batch biofilm reactor treating synthetic mariculture wastewater*. Bioresource Technology, 2017.
1757 **235**: p. 122-130.
- 1758 93. Gao, F., et al., *Effect of florfenicol on performance and microbial community of a sequencing*
1759 *batch biofilm reactor treating mariculture wastewater*. Environmental Technology, 2018. **39**(3):
1760 p. 363-372.
- 1761 94. Zheng, D., et al., *Performance and microbial community of a sequencing batch biofilm reactor*
1762 *treating synthetic mariculture wastewater under long-term exposure to norfloxacin*. 2016. **222**:
1763 p. 139-147.
- 1764 95. Hou, J., et al., *Effects of CeO₂ nanoparticles on biological nitrogen removal in a sequencing batch*
1765 *biofilm reactor and mechanism of toxicity*. 2015. **191**: p. 73-78.
- 1766 96. Xu, Y., et al., *Influence of CeO₂ NPs on biological phosphorus removal and bacterial community*
1767 *shifts in a sequencing batch biofilm reactor with the differential effects of molecular oxygen*.
1768 Environmental Research, 2016. **151**: p. 21-29.
- 1769 97. Xu, Y., et al., *Effects of cerium oxide nanoparticles on the species and distribution of phosphorus*
1770 *in enhanced phosphorus removal sequencing batch biofilm reactor*. Bioresource Technology,
1771 2017. **227**: p. 393-397.
- 1772 98. Xu, Y., et al., *Long term effects of cerium dioxide nanoparticles on the nitrogen removal, micro-*
1773 *environment and community dynamics of a sequencing batch biofilm reactor*. Bioresource
1774 Technology, 2017. **245**: p. 573-580.

1775