PECGylation-Dependent Metabolic Rewiring of Macrophages with Silk Fibroin Nanoparticles

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ABSTRACT: Silk fibroin nanoparticles are emerging as promising nanomedicines, but their full therapeutic potential is yet to be realized. These nanoparticles can be readily PEGylated to improve colloidal stability and to tune degradation and drug release profiles; however, the relationship between silk fibroin nanoparticle PEGylation and macrophage activation still requires elucidation. Here, we used in vitro assays and nuclear magnetic resonance based metabolomics to examine the inflammatory phenotype and metabolic profiles of macrophages following their exposure to unmodified or PEGylated silk fibroin nanoparticles. The macrophages internalized both types of nanoparticles, but they showed different phenotypic and metabolic responses to each nanoparticle type. Unmodified silk fibroin nanoparticles induced the upregulation of several processes, including production of proinflammatory mediators (e.g., cytokines), release of nitric oxide, and promotion of antioxidant activity. These responses were accompanied by changes in the macrophage metabolomic profiles that were consistent with a proinflammatory state and that indicated an increase in glycolysis and reprogramming of the tricarboxylic acid cycle and the creatine kinase/phosphocreatine pathway. By contrast, PEGylated silk fibroin nanoparticles induced milder changes to both inflammatory and metabolic profiles, suggesting that immunomodulation of macrophages with silk fibroin nanoparticles is PEGylation-dependent. Overall, PEGylation of silk fibroin nanoparticles reduced the inflammatory and metabolic responses initiated by macrophages, and this observation could be used to guide the therapeutic applications of these nanoparticles.

KEYWORDS: silk, fibroin, silk nanoparticles, NMR metabolomics, macrophages

INTRODUCTION

The clinical approval of Abraxane in 2005 for the treatment of solid breast tumors was followed by a surge in the use of novel materials in the design of intravenous anticancer nanoparticles that target tumors. Drug-loaded nanoparticles can enter tumor tissues either passively, using the enhanced permeability and retention effect, or actively, using targeting moieties such as peptides, monoclonal antibodies, or aptamers. The physicochemical properties of nanoparticles therefore have a direct impact on their biological performance at the systemic, tissue, cellular, and subcellular levels. Factors such as the particle size, shape, and chemical composition dictate the likelihood of undesirable nanoparticle interactions with complement proteins and immunoglobulins in the circulating blood and with extracellular matrix proteins. These interactions then lead to rapid (<1 min) protein corona formation, opsonization of the particle, and clearance of the nanoparticles by professional macrophages of the mononuclear phagocytic system or by tumor-associated macrophages. This undesired clearance can be circumvented by conjugation of “stealth” polymers, such as poly(ethylene glycol) (PEG), to the nanoparticles. These types of conjugation have become standard practice to allow drug-loaded nanoparticles to evade macrophage detection, thereby maximizing therapeutic payload delivery to the tumor site.

Macrophages are versatile innate immune cells, and their processing of foreign materials (including nanosized drug carriers) plays an essential role in the initiation and coordination of inflammatory or anti-inflammatory immune responses. Until recently, the phagocytosis of nanomedicines was viewed as an undesirable event due to the risks of inducing drug hypersensitivity reactions. However, interest is now growing in the immunomodulatory potential of novel nanomaterials as immunological adjuvants that can influence site-specific inflammation, particularly with regard to the tumor microenvironment. This has led to a revival in the demand for rigorous in vitro techniques that can be used to predict the
immunological outcome of novel materials being tested for advanced drug delivery applications.\textsuperscript{11}

We recently proposed the use of NMR-based metabolomics to identify nanomedicine performance. NMR is a robust analytical technique that can be used to identify and quantify molecular structures such as cellular metabolites.\textsuperscript{12} Our previous comparative metabolomic assessment, which evaluated silk as a novel nanomaterial, revealed that unmodified silk fibroin nanoparticles performed in a similar manner to other emerging and clinically established nanomaterials, such as silica and poly(lactic-co-glycolic acid). Silk fibroin nanoparticles were able to drive metabolic reprogramming of RAW 264.7 macrophages toward induction of a proinflammatory M1-like metabolic profile.\textsuperscript{12}

The design of silk nanoparticles for anticancer drug delivery has intensified over the past 5 years. For example, silk from the Bombyx mori cocoon is reverse-engineered into an aqueous silk fibroin solution using a process that removes sericin (i.e., a gumming agent present in the cocoon).\textsuperscript{13,14} The resulting silk fibroin solution is then desolvated to form spherical nanoparticles with an optimal (∼100 nm) size for intravenous administration.\textsuperscript{15,16} Silk fibroin nanoparticles can be readily decorated with PEG,\textsuperscript{17} and both physical and in vitro biological studies have validated the performances of unmodified and PEGylated silk fibroin nanoparticles for anticancer drug delivery applications.\textsuperscript{18–20} PEGylation significantly improves the colloidal stability while modulating the degradation and drug release profiles of silk fibroin nanoparticles\textsuperscript{17,18} but has no negative impacts on the ability of the nanoparticles to conduct lysosomotropic delivery of anticancer payloads.\textsuperscript{19} However, the immune adjuvanticity between unmodified and PEGylated silk fibroin nanoparticles has not yet been compared. Here, we examine the impacts of nanoparticle concentration and PEGylation on the phenotypic and metabolic reprogramming of macrophages induced by silk fibroin nanoparticles.

### EXPERIMENTAL SECTION

#### Production and Characterization of Unmodified and PEGylated Silk Fibroin Nanoparticles

Silk fibroin was extracted as detailed previously.\textsuperscript{12} Briefly, Bombyx mori cocoons were cut into 5 × 5 mm pieces and degummed in Na$_2$CO$_3$ (0.02 M) for 60 min to quantitatively remove sericin. Fibers were rinsed with ddH$_2$O, air-dried, and dissolved in LiBr solution (9.3 M) at 60 °C, yielding a 25 wt % solution. This solution was dialyzed (molecular weight cutoff 3500 g/mol) against ddH$_2$O for 72 h to remove the LiBr salt. The resulting aqueous silk fibroin solution was cleared by centrifugation (9500 × g for 20 min at 5 °C and repeated once), diluted to 5% (w/v), and added dropwise to acetone to produce spherical silk fibroin nanoparticles. Silk fibroin nanoparticles were surface-decorated with polyethylene glycol (PEG) using a method that has been reported previously,\textsuperscript{15} and a visual format of silk fibroin nanoparticle production is available.\textsuperscript{15} All samples were washed thoroughly and stored at 4 °C for up to 4 weeks.

A Zetasizer Nano-ZS (Malvern Panalytical, Worcestershire, U.K.) was used to characterize the respective particle size distributions, ζ potentials, and polydispersity indices (PDI) of unmodified and PEGylated silk fibroin nanoparticles using dynamic light scattering (DLS) in ddH$_2$O. A refractive index of 1.33 was used for PDI. For phenotypic and metabolic analyses, silks fibroin nanoparticles were either left in their unmodified form or surface-decorated with unlabeled PEG at a 1:1 w/v ratio (as detailed above). For fluorescence analyses, unmodified silk fibroin nanoparticles were either labeled directly with Alexa Fluor 488 or PEGylated with a 1:4 ratio of FITC-PEG and unlabeled PEG, as described below. A schematic of each formulation is shown in Figure 1A.

#### Fluorescence Labeling of Unmodified and PEGylated Silk Fibroin Nanoparticles

Unmodified silk fibroin nanoparticles (1 mL, 10 mg/mL) were resuspended in NaHCO$_3$ (1 mL, 0.2 M, pH 8.6), and Alexa Fluor 488 succinimidyl ester (100 μL, 1 mg/mL) (Thermo Fisher Scientific, Waltham, MA, USA) was added as detailed previously.\textsuperscript{12} Fluorescent PEGylated silk fibroin nanoparticles were produced by resuspending unlabeled silk fibroin nanoparticles (1 mL, 10 mg/mL) in NaHCO$_3$ (1 mL, 0.2 M, pH 8.6) and adding fluorescein PEG succinimidyl ester (250 μL, 30 mg/mL in DMSO) (FITC-PEG-NHS, MW 5000 g/mol, Nanocs, Boston, MA, USA). The sample was incubated for 4 h at room temperature in the dark with stirring, and then TST-activated mPEG (750 μL, 30 mg/mL)
(5000 g/mol, Sigma-Aldrich, St. Louis, MO, USA) in NaHCO₃ (0.2 M, pH 8.6) was added and reacted overnight at 4 °C with stirring. The fluorescently labeled unmodified and PEGylated silk fibroin nanoparticles were then centrifuged (10000 × g for 45 min), and the free (unconjugated) dye was removed by washing the pellet in acidified water (pH 4.6). This was repeated once more with acidified water and then three more times with ultrapure water. All fluorescent nanoparticles were refrigerated (4 °C) and protected from light until use.

**Cell Culture.** RAW 264.7 cells are a mouse macrophage cell line originally extracted from ascites of an Abelson murine leukemia virus-induced tumor and thought to be derived from peritoneal monocytes. The culture of RAW 264.7 cells in complete medium has been described previously. Subconfluent flasks were passaged by scraping cells, centrifuging at 380 × g for 4 min, and resuspending them at ratios of 1:5–1:10 onto plasma-treated tissue culture polystyrene. The cell passage number was controlled during the study, and all experiments were conducted using cells under passage 20. Seeding for polarization experiments was conducted at a density of 1.5 × 10⁵ cells/cm², and cells were allowed to recover for 24 h prior to conducting the in vitro experiments.

**Scanning Electron Microscopy (SEM).** Unmodified or PEGylated silk fibroin nanoparticles (1 mg/mL) in distilled water were lyophilized and sputter-coated with gold and imaged by SEM with 2000-fold magnification at the critical point (EM CPD300, Leica Microsystems, Wetzlar, Germany). Samples were sputter-coated with gold and imaged by SEM with 40000-fold magnification at the critical point (EM CPD300, Leica Microsystems, Wetzlar, Germany). The amide I region was detailed previously. The amide I region was detailed previously. The samples were subjected to 128 scans at a 4 cm⁻¹ resolution over the wavenumber range of 400 to 4000 cm⁻¹. The amide I region was identified: 1605–1615 cm⁻¹ as side chain/aggregated strands, 1616–1637 and 1697–1703 cm⁻¹ as a β-sheet structure, 1638–1655 cm⁻¹ as a random coil structure, 1656–1662 cm⁻¹ as α-helical bands, and 1663–1696 cm⁻¹ as turns.

**Endocytosis of Unmodified and PEGylated Silk Fibroin Nanoparticles.** RAW 264.7 cells were seeded as detailed above onto glass coverslips recovered for 24 h. They were then dosed for 24 h with unmodified or PEGylated silk fibroin nanoparticles (detailed below). Next, cells were fixed (2% v/v glutaraldehyde in phosphate-buffered saline (PBS)), washed with ultrapure water, dehydrated, and dried at the critical point (EM CPD300, Leica Microsystems, Wetzlar, Germany). Samples were sputter-coated with gold and imaged by SEM with 2000-fold magnification at 5 kV.

**Fourier Transform Infrared Spectroscopy (FTIR).** The secondary silk structures were determined for air-dried silk films, 70% ethanol treated silk films, and lyophilized unmodified and PEGylated silk nanoparticles using Fourier transform infrared (FTIR) spectroscopy (TENSOR II FTIR spectrometer, Bruker Optik GmbH, Ettlingen, Germany). As detailed previously, the samples were subjected to 128 scans at a 4 cm⁻¹ resolution over the wavenumber range of 400 to 4000 cm⁻¹. The amide I region was detailed previously. The amide I region was identified: 1605–1615 cm⁻¹ as side chain/aggregated strands, 1616–1637 and 1697–1703 cm⁻¹ as a β-sheet structure, 1638–1655 cm⁻¹ as a random coil structure, 1656–1662 cm⁻¹ as α-helical bands, and 1663–1696 cm⁻¹ as turns.

**Calculation of Colocalization Coefficients.** Colocalization between nanoparticles and lysosomes was determined on representative confocal images using the JACoP ImageJ plug-in to calculate Mander’s overlap coefficients for green (nanoparticle) and magenta (lysosome) channels. Mander’s coefficient ranges from 0 to 1 and indicates the degree of colocalization between two channels. The default JACoP algorithm was used on threshold adjusted images to calculate Mander’s coefficients for each treatment group.

**MTT Assay.** The cytotoxicity of unmodified and PEGylated silk fibroin nanoparticles was verified using 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assays. After seeding and recovery (detailed above), cells were either left untreated or dosed for 24 h in complete DMEM supplemented with (i) 1% v/v Triton X-100, (ii) 0–1 mg/mL unmodified silk fibroin nanoparticles, (iii) 0–1 mg/mL PEGylated silk fibroin nanoparticles, or (iv) PEG (5000 g/mol, Iris Biotech GmbH, Marktredwitz, Germany). MTT (5 mg/mL in PBS) was then added at 10% of the volume of cell culture media, and cells were cultured again at 37 °C for 4 h. The medium was then aspirated, the formazan product was solubilized with DMSO (100 µL), and the absorbance was measured at 570 nm. All data sets were normalized to the Triton X-100 positive control (Figure S1) to determine the percentage of cell viability following each treatment.

**Phenotypic Response to Unmodified and PEGylated Silk Fibroin Nanoparticles.** RAW 264.7 cells were seeded and allowed to recover for 24 h as detailed above. Next, cells were dosed for 24 h in complete medium supplemented with a low (0.1 mg/mL) or high (0.5 mg/mL) concentration of either unmodified or PEGylated silk fibroin nanoparticles (note that macrophages were exposed to silk fibroin nanoparticles without prior priming). A 1% v/v Triton X-100 positive control sample was included to assess plasma membrane integrity. However, for tumor necrosis factor alpha (TNF-α), nitric oxide (NO⁻) production, antioxidant capacity, and reactive oxygen species, a positive control using lipopolysaccharide (1.5 mg/mL) was included instead to provide baseline phenotypic information on RAW 264.7 cells in an activated state. Following the 24 h incubation period, the conditioned growth medium was collected, clarified by centrifugation (380 × g for 10 min), transferred to 2 mL low-protein-binding Eppendorf tubes, and stored at −80 °C. The conditioned growth medium samples were assayed to quantitatively monitor plasma membrane integrity (lactate dehydrogenase assay), TNF-α (TNF-α ELISA), and nitric oxide levels (Griess assay). Cell lysates were generated as detailed below to measure antioxidant capacity and reactive oxygen species.

The lactate dehydrogenase assay was performed with a Pierce LDH Cytotoxicity Assay Kit (Thermo Fisher Scientific, Waltham, MA, USA) following the manufacturer’s instructions to assess membrane integrity in positive and negative controls, nanoparticle, and LPS treated cells.

A mouse TNF-α DuoSet ELISA kit (R&D Systems, Minneapolis, MN, USA) was used to quantify TNF-α levels in a conditioned culture medium. Nitric oxide was measured with the Griess assay as detailed elsewhere.

**Antioxidant Assay.** Cells were treated as detailed above. Next, cells were washed three times with 1× Hank’s balanced salt solution (HBSS), scraped and collected into falcon tubes, and centrifuged at 2000 × g for 10 min at 4 °C. The supernatant was decanted, and the pellet was resuspended in 1 mL HBSS and sonicated on ice. Lysates were then centrifuged at 10000 × g for 15 min at 4 °C, and the supernatant was collected and stored at −80 °C. Total antioxidant capacity was quantified by analyzing supernatant samples with an antioxidant assay kit (Cayman Chemical, MI, USA).

**Reactive Oxygen Species Assay.** Cells were treated as detailed above. Next, cells were washed three times with 1× HBSS, stained for 1 h with CellROX Deep Red Reagent (Thermo Fisher Scientific, Waltham, MA, USA), scraped into 3 mL falcon tubes, and stained with SYTOX Green (ThermoFisherScientific, Waltham, MA, U.S.A.) as an exclusion dye. Cells were analyzed immediately on a BD FACSCanto, with 20000 live events being recorded per sample. Flow cytometry data was subsequently analyzed with FlowJo (FlowJo, LLC, Ashland, OR, USA).

**Cytokine Profile.** A mouse cytokine proteome profiler (Panel A, R&D Systems, Minneapolis, MN, USA) was used, according to the manufacturer’s instructions. Blots from the arrays were exposed to UltraCruz autoradiography film (Santa Cruz Biotech Inc., Dallas, TX, USA), digitized (18 MP Canon EOS100D, Canon Inc., Tokyo, Japan), threshold adjusted, and analyzed using ImageJ v1.51k (National Institutes of Health, Bethesda, MD, USA).

**Metabolomic Sample Preparation.** RAW 264.7 cells were dosed for 24 h with unmodified or PEGylated silk fibroin nanoparticles at either 0.1 or 0.5 mg/mL. Next, the culture medium was collected, clarified by centrifugation (380 × g for 10 min), lyophilized, and stored at −80 °C until analysis. The protocol for
Figure 2. Macrophage response toward unmodified and PEGylated silk fibroin nanoparticles. (A) In vitro cytotoxicity of unmodified (SNP) and PEGylated (PEG-SNP) silk fibroin nanoparticles following a 24 h incubation in complete media supplemented with 0–1000 μg/mL nanoparticles (*, P < 0.05; **, P < 0.01; ***, P < 0.001, one-way ANOVA between control and nanoparticle treated groups; n = 3). (B) Membrane integrity of RAW 264.7 cells following a 24 h incubation in complete media supplemented with 0.1 or 0.5 mg/mL unmodified (SNP) or PEGylated (PEG-SNP) silk fibroin nanoparticles (*, P < 0.05; **, P < 0.01; ***, P < 0.001, one-way ANOVA untreated and nanoparticle treated groups; n = 3). (C) Scanning electron microscope images of RAW 264.7 cells following a 24 h dose with 0.1 mg/mL unmodified (SNP) or PEGylated (PEG-SNP) silk fibroin nanoparticles. Scale bar = 100 μm.

Figure 3. Cellular uptake of unmodified and PEGylated silk fibroin nanoparticles. Live RAW 264.7 macrophages following a 24 h dose of (A) unmodified or (B) PEGylated silk fibroin nanoparticles at concentrations of 0.1 or 0.5 mg/mL. (Scale bars: left = 40 μm; right (zoomed) = 20 μm). Data acquisition using confocal microscopy. Numbers on merged images show Mander’s coefficients calculated for each treatment group. Arrowheads identify colocalization between the nanoparticle-associated and lysosomal signals.

## RESULTS

**Characterization of Unmodified and PEGylated Silk Fibroin Nanoparticles.** DLS was used to quantitatively verify unmodified and PEGylated silk fibroin nanoparticle sizes, ζ potentials, and polydispersity indices of each preparation using different batches (Figure 1). An increase in particle size was noted between the unmodified silk fibroin nanoparticles (96.0 nm) and the PEGylated and FITC-PEGylated silk fibroin nanoparticles (105.3 and 106.4 nm, respectively). The ζ
potential was also significantly influenced by the PEGylation process. In their unmodified state, the silk fibroin nanoparticles had a \( \zeta \) potential of \(-47.9 \) mV. However, this increased to \(-43.4 \) and \(-44.0 \) mV for PEGylated and FITC-PEGylated silk fibroin nanoparticles, respectively. All three silk fibroin nanoparticle types showed a narrow polydispersity (Figure 1B). The FTIR absorbance analyses at 1621 cm\(^{-1}\) confirmed a high abundance of \( \beta \) sheets within the structures of all three nanoparticle formulations when compared to ethanol-treated silk film controls (Figure 1C). Quantitative measurements were supported by SEM images that confirmed the spherical shape of the unmodified and PEGylated silk fibroin nanoparticles (Figure 1D).

Interactions of Unmodified and PEGylated Silk Fibroin Nanoparticles with Macrophages. RAW 264.7 cells were selected as a model mouse macrophage cell line to allow for continuity with our initial study.\(^ {12} \) A combination of biochemical in vitro assays and NMR metabolomics was used to elucidate silk-nanoparticle-induced shifts in phenotypic and metabolic profiles. First, the cytotoxicity of silk fibroin nanoparticles was assessed. Exposure to unmodified silk fibroin nanoparticles caused a significant reduction in cell viability when compared to untreated controls. However, statistical analysis between cells exposed to unmodified or PEGylated silk fibroin nanoparticles at equivalent concentrations revealed no significant differences in cell viability. The unmodified and PEGylated silk fibroin nanoparticles showed cytotoxicities of less than 15\% at exposure levels of up to 100 \( \mu \)g/mL (Figure 2A). At higher concentrations, both particle types reduced cell viability. At the maximum tested concentration, the cell viability was reduced more substantially for the PEGylated silk fibroin nanoparticles than for the unmodified ones; however, the IC\(_{50}\) value remained >1000 \( \mu \)g/mL (Figure 2A). Parallel experiments that tested the cytotoxicity of water-soluble PEG revealed no significant changes in cell viability over the tested concentration range (Figure S2). The macrophage plasma membrane integrity was monitored by lactate dehydrogenase leakage into the culture medium. Exposure of the macrophages to unmodified or PEGylated silk fibroin nanoparticles at either low (0.1 mg/mL) or high (0.5 mg/mL) nanoparticle concentrations caused no leakage of lactate dehydrogenase, indicating no plasma membrane damage (Figure 2B).

We also verified whether endocytic uptake might be involved in the observed effects by studying the intracellular fate of the unmodified and PEGylated silk nanoparticles by live-cell confocal microscopy. We used LysoTracker Red to label the RAW 264.7 cells, and we observed nanoparticle accumulation within lysosomes at 24 h, as evidenced by the substantial colocalization of the silk fibroin nanoparticles with the lysosomal signals. This colocalization occurred regardless of particle PEGylation or concentration, as evidenced by similar Mander’s coefficients (numbers on the merged images in Figure 3). However, the unmodified silk nanoparticles showed some evidence of aggregation at the cell membrane. This aggregation was reduced, and the lysosomes were smaller in the macrophages dosed with PEGylated silk fibroin nanoparticles.

Inflammatory Response to Unmodified and PEGylated Silk Nanoparticles. Next, the inflammatory phenotype of RAW 264.7 macrophages in response to low and high nanoparticle concentrations was determined. Cell exposure to unmodified silk fibroin nanoparticles induced significant TNF-\( \alpha \) release at both nanoparticle concentrations (2.20 and 1.70 ng/mL, respectively) when compared to basal levels (0.14 ng/mL) (Figure 4A). A milder TNF-\( \alpha \) response was observed when macrophages were treated with PEGylated silk fibroin nanoparticles, with only the high concentration causing a significant increase in TNF-\( \alpha \) release (1.14 ng/mL). When assessing nitric oxide levels and reactive oxygen species, neither factor was influenced by a low concentration of either unmodified or PEGylated silk fibroin nanoparticles (Figure 4B,D). However, at the high nanoparticle concentration, unmodified silk fibroin nanoparticles significantly increased nitric oxide levels (111\%) yet decreased reactive oxygen species levels by 64\%. A similar trend was observed with the...
high PEGylated silk fibroin nanoparticle concentration (55% increase of nitric oxide levels and 44% drop in reactive oxygen species). Total antioxidant capacity in RAW 264.7 increased substantially compared to basal levels following treatment with low and high concentrations of unmodified silk fibroin nanoparticles. However, only minimal changes were observed following treatment with PEGylated silk fibroin nanoparticles (Figure 4C).

The cell culture medium was also assayed to further evaluate the cytokine response of RAW 264.7 cells following nanoparticle exposure (Figure 4E). In addition to promoting a marked increase in the expression of TNF-α, the unmodified silk fibroin nanoparticles upregulated the expression of several colony stimulating factors (M-CSF, G-CSF, and GM-CSF), chemokines (CCL1, CXCL 9, CXCL 13), and other essential proinflammatory cytokines (IFN-γ, IL-1α, IL-23, and IL-27). By contrast, PEGylated silk fibroin nanoparticle treatment caused a milder (or negligible) response in proinflammatory cytokine expression when compared to unmodified silk fibroin nanoparticles. The anti-inflammatory cytokine IL-10 remained unchanged regardless of the type of silk fibroin nanoparticle used, whereas IL-1ra expression increased in response to unmodified silk fibroin nanoparticles yet decreased following a dose with PEGylated silk fibroin nanoparticles.

Metabolomic Response to Unmodified and PEGylated Silk Nanoparticles. Next, the metabolic profiles of RAW 264.7 macrophages were analyzed to further characterize the effects of silk fibroin nanoparticle PEGylation and concentration on inflammatory modulation. A score scatter plot (Figure 5A) produced by applying principal component analysis to the 1H NMR spectra of aqueous cell extracts showed a separation between control and nanoparticle treated samples. Samples clustered differently depending on nanoparticle treatment, and in particular the high concentration of unmodified silk nanoparticles separated clearly from the remaining samples along principal component 1 (PC1, explaining 19.5% of total variance). These results were complemented by principal component analysis of lipophilic cell extracts (Figure 5B), which showed some overlap between sample groups, especially in the case of PEGylated particles as they barely separated from controls. By contrast, different clusters were observed for lipophilic samples obtained from cells treated with unmodified silk fibroin nanoparticles, with the 0.5 mg/mL cluster grouping separately from control clusters.

The principal component analysis score map (Figure 6A) obtained for samples from macrophages treated with unmodified silk fibroin nanoparticles confirmed the separation between control, low dose, and high dose samples. Partial least squares discriminant analysis (Figure 6B) further revealed robust discrimination between control and nanoparticle-treated cells (Q2 0.79) due to differences in the levels of metabolites highlighted in the corresponding loading plots.
In particular, silk fibroin nanoparticle treated cells (negative scores in first latent variable, LV1) were characterized mainly by higher intracellular levels of lactate, creatine, phosphocreatine, and glycine (signals with negative LV1 loadings), together with lower levels of a few amino acids, choline compounds, betaine, and myo-inositol (positive loadings). In samples from macrophages treated with PEGylated silk fibroin nanoparticles (Figure 6D), the low and high dose samples largely overlapped in the principal component analysis score scatter plot, suggesting little dependence on nanoparticle concentration. When considering the two exposure concentrations within a single class, nanoparticle treated samples strongly differed from the controls by partial least squares discriminant analysis (Figure 6E). The corresponding loading profiles (Figure 6F) suggested decreased levels of branched chain amino acids and glucose, along with increased levels of glutamate, glutathione, and phosphocholine, in treated cells compared with controls.

Spectral integration was then employed to confirm and extend the analysis of quantitative variations in individual metabolites (summarized in Figure 7). Overall, silk fibroin nanoparticles caused changes in 28 intracellular polar metabolites associated with different metabolic pathways. When measuring glycolytic metabolites and metabolites of the tricarboxylic acid cycle (Figure 7A), cells exposed to unmodified silk fibroin nanoparticles displayed a pronounced decrease in pyruvate levels, while lactate, itaconate, and succinate increased significantly. By contrast, succinate and intracellular glucose levels decreased in cells treated with PEGylated silk fibroin nanoparticles. When comparing nanoparticle-induced changes to amino acid metabolism (Figure 7A), unmodified silk nanoparticles decreased in the intracellular levels of aspartate, glutamine, glutamate, and alanine, whereas levels of glycine, lysine, branched chain, and aromatic amino acids increased when compared to untreated controls. Interestingly, PEGylated silk fibroin nanoparticles either produced no change in some of these amino acids or caused variations in the opposite direction to that observed for unmodified particles. Energy-related metabolites were also affected by silk fibroin nanoparticle treatment (Figure 7A). When treated with unmodified particles, significant increases to creatine and phosphocreatine levels were observed at both concentrations, while only the high concentration impacted ATP levels (significant decrease). Again, milder effects were observed for macrophages treated with PEGylated silk fibroin nanoparticles. Marked changes were observed in metabolites that can act as antioxidants and/or osmoregulators or that relate to cell membrane composition (Figure 7A). In general, these metabolites showed dose-dependent decreases in cells treated with unmodified silk fibroin nanoparticles, whereas they showed increased levels in cells treated with PEGylated silk fibroin nanoparticles. Metabolites associated with lipid metabolism (Figure 7A) were also differentially affected by the two nanoparticle types, with increases in 3-hydroxybutyrate, acetate, and formate being seen only in cells treated with unmodified silk fibroin nanoparticles.

Changes in metabolic pathways were also apparent through analysis of the exometabolome (Figure 7B), with cells exposed to PEGylated silk fibroin nanoparticles consuming more glucose and excreting more lactate and itaconate when compared to untreated cells. These effects were more pronounced in cells treated with unmodified silk fibroin nanoparticles, and these cells also excreted more alanine and less citrate when compared to untreated controls. Finally, unmodified silk fibroin nanoparticles caused increases in total/esterified cholesterol and phosphatidylcholine, together with decreases in unsaturated fatty acids (Figure 7C). By contrast, the effects of PEGylated silk fibroin nanoparticles were restricted to mild increases in total cholesterol and phosphatidylcholine.

### DISCUSSION

The biopolymer silk has a long clinical track record in humans, and silk nanoparticles have recently emerged as an interesting platform for drug delivery, including anticancer drugs. However, the interactions of silk nanoparticles with macrophages have yet to be fully characterized, and this lack of knowledge limits our current understanding of what effects these nanoparticles will have on off-target tumor-associated macrophages. Tumor-associated macrophages show a tumorigenic profile, and a high distribution of these macrophages within a tumor is associated with poor patient prognosis. However, these macrophages can undergo reprogramming to...
assume a proinflammatory state that confers tumoricidal properties. Therefore, since anticancer nanoparticles encounter macrophages upon accessing a tumor site, they provide a unique opportunity to modulate the inflammatory profiles of tumor-associated macrophages and thereby maximize the intratumoral performance. The aim of the present study was to examine the baseline performances of both unmodified and PEGylated silk fibroin nanoparticles by monitoring the effects of particle PEGylation and concentration on the inflammatory phenotype and metabolic profiles of macrophages. We have opted to study the response of silk fibroin nanoparticles without an anticancer drug payload to simplify data interpretation and to eliminate drug-mediated confounding effects (e.g., drug-induced cytotoxicity).

Similar to our previous reports, we found that PEGylation of silk fibroin nanoparticles increased the hydrodynamic radius and moderated the surface charge when compared to unmodified nanoparticles (Figure 1B); an observation that aligns well with the literature (reviewed by Rabanel et al.). We now provide additional evidence that the interaction of either unmodified or PEGylated silk fibroin nanoparticles with the macrophage plasma membrane caused no damage to the cell membrane (Figure 2B). As we have observed previously, the unmodified silk nanoparticles had a higher propensity to aggregate in physiological media when compared to the PEGylated silk fibroin nanoparticles (Figure 2C). However, this aggregation did not affect the macrophages’ ability to phagocytose and traffic unmodified silk fibroin particles into lysosomes (Figure 3A,B). The somewhat larger appearance of the lysosomes containing unmodified silk fibroin nanoparticles, when compared with those containing PEGylated silk fibroin nanoparticles, might be a consequence of the larger size of the internalized nanoparticle aggregates.

Both silk fibroin nanoparticle formulations were internalized and trafficked into lysosomes, but the downstream effects on the inflammatory profile of the macrophages were dependent on the surface modification and particle concentration. A high concentration of unmodified silk fibroin nanoparticles primed the RAW 264.7 macrophages toward a proinflammatory phenotype, as evidenced by significant increases in levels of classical proinflammatory mediators (e.g., TNF-α, nitric oxide, Figure 3A,B), enhanced total antioxidant activity, and reduction in levels of reactive oxygen species (Figure 3C,D). Reactive oxygen species levels are implicated in the differentiation of macrophages into wound-healing and tumor-associated phenotypes. However, even at the low nanoparticle concentration, unmodified silk fibroin nanoparticles were able to induce a proinflammatory shift in the macrophage phenotype, although to a lesser extent than the shift observed at high concentrations (i.e., less production of nitric oxide and reactive oxygen species, but similar promotion of TNF-α and antioxidant levels when compared to the high nanoparticle dose).

The biological significance of macrophage priming at the high silk fibroin nanoparticle dose was also evident by the observed upregulation of the macrophage cytokine profile (e.g., colony stimulating factors, chemokines, and other essential proinflammatory substances) (Figure 3C). However, PEGylation apparently modulated the immunomodulatory effects of silk fibroin nanoparticles. At the high nanoparticle concentration, PEGylation moderated the production of proinflammatory mediators, including TNF-α, nitric oxide, total antioxidant activity, and reactive oxygen species when compared to the response to unmodified silk fibroin nanoparticles (Figure 4). At the high concentration, the PEGylated silk fibroin nanoparticles only evoked subtle differences in cytokine production between the treated and control cells (Figure 4D). Overall, these observations are important because they suggest the possibility of differential priming of macrophages, even at high nanoparticle concentrations.

While nanoparticles for drug delivery are typically designed to evade the immune system, a proimmunogenic phenotype could be exploited to maximize intratumoral performance. We and others have shown a link between metabolic reprogramming and cell behavior, such as macrophage activation status. Therefore, we examined the baseline performances of both unmodified and PEGylated silk nanoparticles to monitor nanoparticle-induced metabolic changes. Exposure to silk fibroin nanoparticles affected macrophage metabolism in a nanoparticle-type- and concentration-dependent fashion (Figure 6). In agreement with our previous study, similar to the metabolic phenotype described for lipopolysaccharide-activated proinflammatory macrophages, cells treated with unmodified silk nanoparticles increased cellular glycolytic activity, as indicated by significantly increased glucose consumption, upregulated pyruvate to lactate conversion, and increased lactate and alanine excretion. Increased glycolysis was also apparent in cells treated with PEGylated nanoparticles, although the lactate levels in this case were only elevated in the medium supernatants, not intracellularly.

Cells treated with unmodified silk fibroin nanoparticles further displayed enhanced production of succinate and itaconate, which is consistent with the reprogramming of the tricarboxylic acid cycle associated with a proinflammatory state. Succinate is known to inhibit cytosolic prolyl hydroxylases, thereby stabilizing HIF1α, whose target genes include glycolytic enzymes and inflammatory factors. Additionally, succinate oxidation by succinate dehydrogenase drives the production of reactive oxygen species. By contrast, itaconate, which is produced from citrate via α-ketoglutarate, inhibits succinate dehydrogenase and acts as a counterbalance to limit inflammation. This metabolite inhibits succinate dehydrogenase and activates the anti-inflammatory and antioxidant transcription factor Nrf2, thus restricting downstream inflammatory gene expression and limiting the production of proinflammatory mediators like IL-1β and IL-6. Activation of the Nrf2/antioxidant response element pathway could result in increased expression of antioxidant enzymes (e.g., catalase, superoxide dismutase), as well as increased glutathione synthesis. Therefore the observed itaconate increase and enhanced antioxidant capacity could ultimately equip cells better to neutralize reactive oxygen species. This proposal is consistent with the observed decrease in reactive oxygen species upon nanoparticle treatment. Therefore, our results corroborate a close link between rewiring of the tricarboxylic acid cycle, the cell redox status, and the production of proinflammatory cytokines. Another consequence of this type of metabolic reprogramming could be the observed ATP depletion in cells exposed to a high concentration of unmodified silk fibroin nanoparticles, which, as noted in our previous study, could be due to both inhibition of succinate dehydrogenase (and hence oxidative phosphorylation) and downregulation of substrate-level phosphorylation.
The marked decrease in intracellular levels of some amino acids in cells treated with unmodified silk fibroin nanoparticles further supports tricarboxylic acid cycle reprogramming. In particular, glutamine/glutamate and aspartate could be used to replenish α-ketoglutarate and oxaloacetate, respectively, and so could be involved in the aspartate-arginosuccinate shunt, which is important in lipopolysaccharide-induced macrophage activation.45 By contrast, a different set of amino acids (glycine, lysine, branched chain, and aromatic amino acids) increased in cells treated with unmodified silk nanoparticles, possibly reflecting protein catabolism.12 The PEGylated silk fibroin nanoparticles did not have a similar effect on the tricarboxylic acid cycle. No intracellular increases were noted in succinate or itaconate upon nanoparticle treatment although succinate levels were decreased at the higher nanoparticle dose. Clear reduction in tricarboxylic acid cycle rewiring and itaconate production coincided with a reduced inflammatory phenotype in RAW 264.7 macrophages, and this supports observations made with unmodified silk fibroin nanoparticles.

Another prominent effect of unmodified silk fibroin nanoparticles was the significant intracellular upregulation of creatine and phosphocreatine. Although the functional relevance of these metabolites in macrophage responses to nanoparticles remains to be determined, one possibility is that the creatine kinase/phosphocreatine system plays an important role, possibly in relation to energy buffering and phagocytic function.56 Interestingly, PEGylated silk fibroin nanoparticles produced a milder increase in phosphocreatine and no change in creatine levels, suggesting a less extensive impact on the creatine kinase system. One other explanation for the increased lysosome size observed in response to unmodified silk fibroin nanoparticles might be a differential demand on the creatine kinase system of the phagocytosing macrophages (Figures 2 and 3). The lower aggregation of PEGylated silk fibroin nanoparticles could possibly place less demand on the creatine kinase system. This observation warrants further investigation.

The levels of several metabolites involved in osmoregulation, antioxidant protection, and/or cell membrane composition also showed distinct variation patterns in cells exposed to unmodified or PEGylated silk fibroin nanoparticles. In response to unmodified silk fibroin nanoparticles the levels of antioxidant metabolites glutathione (albeit increased at the lower dose treatment), taurine, and betaine decreased, which may possibly reflect their use to counterbalance reactive oxygen species generation by proinflammatory macrophages.57,58 The observed decrease in reactive oxygen species levels in nanoparticle-treated cells is in line with these observations. By contrast, the levels of glutathione and betaine, to a lesser extent, increased in cells treated with PEGylated silk nanoparticles. Opposite variations were also found for myo-inositol, phosphocholine, and glycophosphocholine, which are metabolites with known involvement in membrane remodeling and osmotic balance.

Last, unmodified silk fibroin nanoparticles, but not PEGylated ones, caused increases in metabolites that could arise from enhanced lipid metabolism: 3-hydroxybutyrate, acetate, and formate. Changes in cellular lipid composition were also apparent from the analysis of lipophilic extracts (Figure 6C). Treatment with unmodified silk fibroin nanoparticles and, to a lesser extent, PEGylated particles caused cells to increase their cholesterol and phospholipid content. Additionally, the levels of unsaturated fatty acids decreased upon exposure to unmodified nanoparticles, again possibly reflecting remodeling of membrane composition and fluidity.

### CONCLUSIONS

In summary, we have demonstrated that unmodified—and, to a lesser extent, PEGylated—silk fibroin nanoparticles modulated macrophage activation and metabolism in a concentration-dependent and particle-type-dependent manner. Specifically, unmodified nanoparticles induced a proinflammatory phenotype that was characterized by increased cytokine release, enhanced nitric oxide production, and elevated antioxidant levels, as well as decreased reactive oxygen species levels. Extensive rewiring of the metabolic profiles, including glycolysis, the tricarboxylic acid cycle, and amino acid and antioxidant metabolite biosynthesis, as well as creative kinase system activity, was also observed. This ability to modulate the macrophage phenotype could be especially useful in the context of anticancer drug delivery, as silk fibroin nanoparticles would act both as carriers for chemotherapeutics and as synergistic attenuators of tumor-associated macrophages toward a more tumoricidal phenotype.

### ASSOCIATED CONTENT

#### Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acsami.8b18716. Positive and negative controls used to calculate mean cell viability in MTT cytotoxicity assays \((n = 3)\); in vitro cytotoxicity, determined by MTT, of PEG following a 24 h incubation in complete media supplemented with 0–1000 μg/mL nanoparticles \((n = 3)\) (PDF)

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**Notes**

The authors declare no competing financial interest.

All data created during this research are openly available from the University of Strathclyde-Pure athttps://doi.org/10.15129/fo3c40f2-1eef-44a0-891a-628abbe5cb16.

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