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Radical-Mediated C–H Functionalization: A Strategy for Access to Modified Cyclodextrins

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ABSTRACT

A simple and efficient radical C–H functionalization to access modified cyclodextrins (CDs) has been developed. The well-defined conformation of glycosidic and aglyconic bonds in α-, β-, and γ-CDs favors the intramolecular 1,8-hydrogen atom transfer (HAT) promoted by the 61-O-yl radical which abstracts regioselectively the hydrogen at C5II of the contiguous pyranose. The C5II-radical evolves by a polar crossover mechanism to a stable 1,3,5-trioxocane ring between two adjacent
glucoses or alternatively triggers the inversion of one α-D-glucose into a 5-C-acetoxy-β-L-idose unit possessing a $^1\text{C}_4$ conformation. The $6_{\text{IV}}$- and $6_{\text{III}}$-diols of α- and β-CDs behave similarly to the monoalcohols, forming mostly compounds originating from two 1,8-HAT consecutive processes. In the case of $6_{\text{II}}$-diols the proximity of the two 6-O-yl radicals in adjacent sugar units allows the formation of unique lactone rings within the CD framework via a 1,8-HAT–β-scission tandem mechanism. X-Ray diffraction carried out on the crystalline 1,4-bis(trioxocane)-α-CD derivative shows a severe distortion toward a narrower elliptical shape for the primary face.

**INTRODUCTION**

Over the past decades, cyclodextrins (CDs) have been extensively investigated for their valuable applications in several areas.¹ These naturally-occurring cyclic oligosaccharides are composed of glucose units in $^4\text{C}_1$ chair conformations, with a hydrogen bond network formed between the secondary hydroxyl groups, creating a conical shape with an internal hydrophobic cavity, conferring the ability to form host-guest or inclusion complexes with a great variety of molecules and rendering them one of the most important supramolecular host families. Because of these inclusion properties, CDs are displayed as being potentially useful as drug and gene delivery systems,² artificial enzymes³ or catalysts,⁴ just to cite a few.

Considering their unique features and relevant applications, great effort has been invested in developing chemical modifications of native CDs to modulate their chemophysical properties, and thus achieve an enhancement of their solubility, a change in their cavity size or an improvement in their inclusion capability. The strategies are usually based on grafting a wide assortment of substituents taking into account the different accessibility and reactivity of the hydroxyl groups to
obtain diverse patterns of functionalization, which act as platforms to access amphiphilic CDs, and hence, sophisticated molecular architectures.

It should be pointed out that most of these methodologies have addressed the structural changes retaining the $4C_1$ chair conformation of the glucose units, but a few of them describe modifications of the cavity by altering the conformations of these units. Thus, the reported synthesis of 3,6-anhydro-CDs shows the conformational change of the pyranose units to $1C_4$, and consequently, a significant change in their binding properties. Moreover, the formation of a mono-2,3-epoxide-$\beta$-CD followed by a nucleophilic ring opening generates an altrose unit in a $1C_4$ which distorts the cavity from conical to elliptical. The introduction of several altrose moieties reflects a rapid equilibrium between $1C_4$ and $4C_1$ conformers and their ratio depends on the number of residues and their position in the macromolecule. The cycloglycosylation syntheses of cyclodextrin analogues composed exclusively of L-sugars or incorporating both L- and D-sugars residues have been reported. Finally, it has been found that in the crystalline structure of permethylated $\beta$-CD one of the D-glucose residues is inverted from the normal $4C_1$ to the $1C_4$ chair conformation.

Interestingly, in the methodologies employed to create this structural diversity in CDs, regardless of conformational changes, radical reactions are practically unknown and this fact is not surprising if we consider that in simple carbohydrates they mainly involve the anomeric position which is committed in the cyclic CD structure. Only one example has been reported in the literature and is focused on the intermolecular attack of reactive-oxygen-centered free radicals on CDs in aqueous solution using EPR detection experiments.

During the last several years, our group has been immersed in the development of methods for the activation and functionalization of “inert” C–H bonds in carbohydrate chemistry. Of special
interest is a novel 1,8-hydrogen atom transfer reaction (1,8-HAT) which takes place, through a
rather unusual nine-membered transition state (TS), between both pyranose units in a Hexp-(1→4)-Hexp disaccharide system. The hydrogen abstraction at C5H is promoted, in a highly regioselective
manner, by the electrophilic 61-O-yl radical I generated from the corresponding alcohol under
oxidative conditions, producing a C5H radical II (Scheme 1).

Scheme 1. Radical polar crossover mechanism for the HAT reaction promoted by 6-O-yl radicals in Hexp-(1→4)-Hexp systems of carbohydrates. DIB = (diacetoxyiodo)benzene.

This intermediate suffers a one-electron oxidation causing a crossover from the radical to the
oxacarbenium ion III which is subsequently trapped by the alcohol to give the intramolecular
bridged eight-membered 1,3,5-trioxocane IV. The obtained results show that this regioselectivity
is highly dependent on both the four stereogenic centers implicated in the cyclization step (C5H,
C1\textsuperscript{II}, C4\textsuperscript{I}, and C5\textsuperscript{I}) as the conformation of the glycosidic (Φ = H1\textsuperscript{II}–C1\textsuperscript{II}–O4\textsuperscript{I}–C4\textsuperscript{I}) and aglyconic (Ψ = C1\textsuperscript{II}–O4\textsuperscript{I}–C4\textsuperscript{I}–H4\textsuperscript{I}) bonds. In fact, when these stereochemical and conformational requisites are not suitable for the 1,8-HAT to take place, the alkoxyl radical I may alternatively abstract the hydrogen at C1\textsuperscript{II} via a 1,6-HAT to produce a new C1\textsuperscript{II} radical V which, after oxidation to the respective oxacarbenium ion VI and cyclization, gives the spiro ortho ester VII.

For the β-maltose disaccharide, α-D-Glcp-(1→4)-β-D-Glcp, the reaction proceed with complete regioselectivity, yielding compounds formed exclusively from the 1,8-HAT.\textsuperscript{13a} In this case, the corresponding glycosidic and aglyconic bonds in the TS involved (Φ = −32.7, Ψ = −37.3°) are arranged in an exo-syn conformation, where the alkoxyl radical and the H5\textsuperscript{II} are at an optimal distance of 3.1 Å for the initial abstraction step.\textsuperscript{14}

With this result in mind, we decided to investigate whether this radical-polar crossover methodology could be extended to more complex carbohydrates such as CDs, since in these systems the D-glucose units are linked in a similar fashion (cyclo-[α-D-Glcp-(1→4)]\textsubscript{n}). The measures of distances and angles made on the reported X-ray structure of permethylated β-CD\textsuperscript{15} as well as on its minimized structure indicate that there is sufficient inter-residue flexibility in the macromolecule to adopt an exo-syn conformation similar to β-maltose. Therefore, from a theoretical point of view, the 1,8-HAT process could take place between two vicinal units of the macrosystem to provide modified CDs on their primary face, which could exhibit important distortions in their cavities.

Preliminary results of this work have been described recently and we now present here the full details of the process and the extension to other positional isomeric diols and per-6-ol models.\textsuperscript{16}
RESULTS AND DISCUSSIONS

Initially, the most common and readily available β-cyclodextrin was the substrate of choice in order to investigate the viability of this radical process. The corresponding monoalcohol 2\textsuperscript{I-VII},3\textsuperscript{I-VII},6\textsuperscript{I-VII}-icosa-O-methyl-β-CD (1) necessary to assay the reaction was prepared from the commercial β-CD following a well-known three-step sequence consisting in monosilylation of one primary alcohol, O-methylation of the remaining hydroxyl groups and finally removal of silyl-ether.\textsuperscript{17} This protocol has the advantage of using methyl ethers as protecting groups which usually allow an increase in solubility both in water and in organic solvent.\textsuperscript{18} To our delight, visible light irradiation of alcohol 1 with diacetoxyiodobenzene (DIB) (2.2 equiv) and iodine (1 equiv) at 30 °C proceeded nicely in 30 minutes delivering the expected 1,3,5-trioxocane 2 (36%) and the acetate 3 (61%) as the major compound (Table 1, entry 1).\textsuperscript{19} Lower reaction temperatures or fewer equivalents of reagents (DIB or I\textsubscript{2}) only gave rise to slower and incomplete reactions while the increase in the reaction times or the amount of reagents allowed us to obtain exclusively trioxocane 2 in 60% yield (entry 2). The reaction is plausibly explained by a sequential intermolecular nucleophilic attack by acetate anion to give 3 followed by an intramolecular cyclization leading to 2. A competitive addition of both nucleophiles to the oxacarbenium ion intermediate may also be considered.\textsuperscript{20}

Obviously, both products, 2 and 3, derived from the same C5\textsuperscript{VII} radical intermediate, through an intramolecular 1,8-HAT reaction promoted by the 6\textsuperscript{I}-O-yl radical. No compounds resulting from abstraction at C1\textsuperscript{VII} via a 1,6-HAT process were detected.

In both compounds, the oxidation at C5\textsuperscript{VII} was clearly confirmed by 1D and 2D NMR experiments. Thus, for compound 2, the HMBC correlation of the H1\textsuperscript{VII} proton signal at \(\delta\textsuperscript{H} 5.06\) ppm with the C5\textsuperscript{VII} quaternary carbon atom (\(\delta\textsuperscript{C} 101.04\) ppm) corroborates this C5\textsuperscript{VII} functionalization. Moreover,
all D-glucopyranose units are preferentially in $^4C_1$ chair conformations as we can be deduced from the values of the coupling constants of the anomeric hydrogens ($^3J_{1,2} \approx 3.6$ Hz).

Table 1. 1,8-HAT for 6$^1$-ol-$\beta$-CD 1, 6$^1$-ol-$\alpha$-CD 5, and 6$^1$-ol-$\gamma$-CD 8.\textsuperscript{a}

<table>
<thead>
<tr>
<th>entry</th>
<th>substrate</th>
<th>l(III)/I$_2$ (equiv)</th>
<th>products (yield)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>1; $\beta$-CD</td>
<td>DIB/I$_2$ (2.2/1.0)</td>
<td>2 (36%) 3; R = Ac (61%)</td>
</tr>
<tr>
<td>2</td>
<td>1; $\beta$-CD</td>
<td>DIB/I$_2$ (3.0/1.7)</td>
<td>2 (60%)</td>
</tr>
<tr>
<td>3</td>
<td>1; $\beta$-CD</td>
<td>Phi(OCOAd)/I$_2$ (2.2/1.0)$^b$</td>
<td>No reaction</td>
</tr>
<tr>
<td>4</td>
<td>1; $\beta$-CD</td>
<td>Phi(OCO-2,4,6-iPr$_3$Ph)/I$_2$ (2.2/1.0)</td>
<td>No reaction</td>
</tr>
<tr>
<td>5</td>
<td>1; $\beta$-CD</td>
<td>Phi(OCO-2,4,6-Me$_3$Ph)/I$_2$ (2.2/1.0)</td>
<td>2 (46%) n.d.</td>
</tr>
<tr>
<td>6</td>
<td>1; $\beta$-CD</td>
<td>Phi(OCO-2,4,6-Me$_3$Ph)/I$_2$ (2.2/1.0)</td>
<td>2 (21%) 4; R = CO-2,4,6-Me$_3$Ph (33%)</td>
</tr>
<tr>
<td>7</td>
<td>5; $\alpha$-CD</td>
<td>DIB/I$_2$ (2.2/0.5)</td>
<td>6 (31%) 7; R = Ac (30%)</td>
</tr>
<tr>
<td>8</td>
<td>5; $\alpha$-CD</td>
<td>DIB/I$_2$ (2.2/1.0)</td>
<td>6 (59%)</td>
</tr>
<tr>
<td>9</td>
<td>8; $\gamma$-CD</td>
<td>DIB/I$_2$ (2.2/0.5)</td>
<td>9 (36%) 10; R = Ac (41%)</td>
</tr>
<tr>
<td>10</td>
<td>8; $\gamma$-CD</td>
<td>DIB/I$_2$ (2.2/1.0)</td>
<td>9 (44%)</td>
</tr>
</tbody>
</table>

\textsuperscript{a}Reaction conditions: A solution of the alcohol in dry CH$_2$Cl$_2$ containing the organoiodine(III) reagent and I$_2$ was stirred under nitrogen while irradiated with two 80 W tungsten-filament lamps. n.d. = not determined. $^b$Ad = 1-Adamantyl.

With respect to the acetyl derivative 3, the deshielding observed for the ring protons of the unit VII bearing the acetyl group and the magnitude of the $^3J_{H,H}$ coupling constants [$^3J_{1,2} = 1.9$ Hz (calcd 1.3 Hz), $^3J_{2,3} = 2.5$ Hz (calcd 2.6 Hz), and $^3J_{3,4} = 1.9$ Hz (calcd 3.3 Hz)] suggest that the ring exists substantially in a $^1C_4$ inverted chair conformation.\textsuperscript{21} Moreover, the NOE experiment shows an interaction between H1$^{\text{VII}}$ and the acetyl methyl protons, thus tentatively establishing the stereochemistry at C5$^{\text{VII}}$ as R. These data indicate that the acetate group has been incorporated with
inversion of configuration at C5VII and consequently the original α-D-glucopyranose has been transformed into a 5-C-acetoxy-β-L-idopyranose unit,22 flipping from 4C1 to 1C4 chair conformation. As far as we know, this is the first precedent of a β-CD molecule having a β-L-idose unit in its structure. Conversely, in the formation of trioxocane 2, the intramolecular nucleophilic addition to the oxacarbenium ion occurs with retention of configuration at C5VII or by a double inversion if we consider 3 to be an intermediate.

To achieve additional insight into the incorporation of acetate, we decided to explore the reaction with different bis(acyloxy) hypervalent iodine(III) reagents to evaluate the nucleophilic addition of other acyloxyl groups to the C5 position.23 As depicted in Table 1, in the cases of the sterically demanding 1-adamantylcarboxyl (entry 3) or 2,4,6-triisopropylbenzoyloxyl (entry 4), the reaction did not proceed, recovering most of the unchanged starting material in both cases. This may be caused by the steric hindrance of the acyl hypoiolate intermediate which precludes entirely the formation of the 6I-O-yl radical.24 When the ester was the 4-tert-butylbenzoyloxyl (entry 5), TLC analysis showed the completion of the reaction and the presence of two main products. However, after chromatographic separation we were able to isolate only the cyclized compound 2 in 46% yield, presumably due to intramolecular hydrolytic cleavage of the ester. Only the reaction with bis(2,4,6-trimethylbenzoyloxy)iodobenzene allowed us to isolate both products, 2 and 4, albeit in moderate yield, 21% and 33% respectively, together with a significant amount of starting material (39%) (entry 6). The NMR data for 4 show clearly that, also in this case, the ring chair conformation of unit VII has been inverted.

The effectiveness of this methodology was tested on α- and γ-CD models. The corresponding monoalcohols 525 and 826 were prepared following the same three-step protocol employed for the 6I-ol-β-CD derivative 1. Using similar amounts of DIB and iodine, both substrates behaved in an
analogous manner to the β-CD case, generating the corresponding trioxocane derivatives 6 and 9 and acetates 7 and 10, albeit in somewhat lower overall yields (Table 1, entries 7–10). While the trioxocane derivatives are stable for long periods of time at room temperature and can be easily manipulated, the acetates seem to be acid- and base-sensitive compounds. The use of an excess of iodine (entries 8 and 10), aqueous work up or standard silica gel column chromatographic purification preclude their isolation and the reaction gives exclusively the respective trioxocane derivatives 6 and 9 in variable yields. Fortunately, pure acetates 7 and 10 could be obtained using 0.5 equiv of iodine, avoiding any work up and after careful flash chromatography employing silica gel scraped from commercially coated TLC plates (entries 7 and 9).

As described above for the β-CD derivative 3, the structures and conformations of the acetyl derivatives 7 and 10 were corroborated by 1D TOCSY and 2D HSQC and HMBC experiments. Remarkably, the conformation of unit VI of compound 7 exhibits changes in the TOCSY experiments by varying from CDCl₃ to C₆D₆. The observed values of the coupling constants of this unit in CDCl₃ [J₁,₂ = 2.8 Hz (calcd 1.4 Hz), J₂,₃ = 3.2 Hz (calcd 2.5 Hz), and J₃,₄ = 2.8 Hz (calcd 3.3 Hz)] indicate predominantly a ¹C₄ conformation while in C₆D₆ these values [J₁,₂ = 3.5 Hz (calcd 3.2 Hz), J₂,₃ = 8.5 Hz (calcd 9.7 Hz), and J₃,₄ = 7.9 Hz (calcd 9.2 Hz)] preferentially point to a ⁴C₁ conformation.²¹ In contrast, the acetates 3, 4, and 10, derived from β- and γ-CD, show the same ¹C₄ conformation in both solvents. The inversion barrier seems to be significantly lower in the more strained hexameric ring.

Encouraged by these promising results with mono-alcohols, we decided to investigate what would happen in this HAT process if we have two primary hydroxyl groups in the molecule which can react simultaneously, a situation hitherto little studied.²⁷ Taking into account the structure of β- and α-CDs, there are three different positional isomers (6¹III-, 6¹III, and 6¹IV-diol) for each of them.
With the aim of obtaining cyclized compounds, we examined the relative stability of the expected final products, the 1,4-, 1,3-, and 1,2-bis(1,3,5-trioxocane) compounds. A preliminary analysis using molecular mechanic calculations revealed that the energetic differences among the three isomers, in either \( \beta \)- or \( \alpha \)-CD, are very small (approx. 2 kcal/mol), providing evidence for the thermodynamic feasibility of the cyclization step.\(^{28}\)

Therefore, we decided to synthesize the three isomeric diols of each CD using a similar protection-deprotection sequence employed for the monoalcohols, as has been described previously,\(^{17a}\) and assess the scope of this methodology.

The study was begun with the more distant 1,4-diol derivatives, that we believe have greater possibilities of success. Firstly, we carried out the reaction of the 6\(^{1,4}\)-diol-\( \beta \)-CD derivative 11\(^{17a}\) with DIB and I\(_2\) as shown in Scheme 2. Gratifyingly, the process proceeded in good overall yield, affording the 1,4-bis(trioxocane) 12 (20%), the diacetate 13 (49%) as the main product, and a 3:2 chromatographically inseparable mixture of mono-trioxocane–mono-acetyl positional isomers (14%, not shown, see the Experimental Section) which could be transformed into 12 after treatment with iodine. Analogously, compound 12 was obtained when 13 was submitted to these iodine catalyzed conditions. The acetyl methyl protons at C5\(^{III}\) and C5\(^{VII}\) in compound 13 were readily assigned by 2D HSQC and HMBC experiments, whereas the \(^1\)H NMR showed the deshielding of the ring protons in these units III and VII and the values of their coupling constants (\(^3\)J\(_{H,H} \approx 1.9–2.5\) Hz) were consistent with the introduction of both acetates with inversion at C5 and thus, with the presence of two \( \beta \)-L-idose units in \(^1\)C\(_4\) conformations. Although no chair inversion was detected using C\(_6\)D\(_6\) as solvent, the axial H1\(^{III}\) and H1\(^{VII}\) protons started to broaden at 18 °C suggesting that coalescence is just below this temperature, while the signals were sharpened by heating at 70 °C. Additionally, the stereochemistry of the quaternary centers C5\(^{III}\) and C5\(^{VII}\) were established.
tentatively as R on the basis of NOE interactions between H1\textsubscript{III} or H1\textsubscript{VII} and the respective acetyl methyl protons.

**Scheme 2. 1,8-HAT for the 6\textsubscript{I,IV}-diol-\(\beta\)-CD 11 and 6\textsubscript{I,IV}-diol-\(\alpha\)-CD 14**

Next, we explored the behavior of 6\textsubscript{I,IV}-diol-\(\alpha\)-CD derivative 14\textsuperscript{25,29} under these oxidative radical conditions. The reaction afforded the bis- and mono-trioxocanes 15 and 16 as sole compounds, not detecting any product from incorporation of acetate into the molecule, and thus, revealing the greater steric hindrance in this hexamer (Scheme 2). Moreover, compound 15 is a crystalline solid whose structure was unambiguously confirmed by X-ray crystallographic analysis, showing two trioxocane rings in a restricted boat-chair (BC) conformation with a guest n-hexane molecule in the CD cavity (Figure 1).\textsuperscript{30} The simplified \(^1\)H and \(^{13}\)C NMR spectra are consistent with the C\(_2\) symmetry of the proposed structure.

The comparison of the geometrical structural parameters of 15 with those of the related permethylated \(\alpha\)-CD\textsuperscript{31} shows that the presence of the two 1,3,5-trioxocane rings severely distort
the primary face of the molecule. This distortion is clearly observed when comparing the geometrical parameters of the irregular hexagon comprising the side-chain carbon atoms (C6\textsuperscript{a}), which points toward a much narrower elliptical shape for this rim (see Tables S2 and S3 in the Supporting Information). Conversely, the presence of the bis(trioxocane) rings does not significantly alter the secondary face of the molecule. Thus, for instance, the distances and angles between the six interglycosidic oxygen atoms (O4\textsuperscript{a}), as well as the radii of the gravity center of the hexagon formed by these O4\textsuperscript{a} oxygen atoms are quite similar in both X-ray structures. The most significant differences are observed in the tilt angles made by the O4\textsuperscript{a} mean plane and the mean planes through the glucose units principally in residues II and V adjacent to the trioxocane rings. What do not seem to be affected either are the 4\textsuperscript{C1} chair geometries of the six glucose units, as shown by the Cremer–Pople puckering parameters which describe slightly distorted chairs similar to those found in the permethylated \( \alpha \)-CD (see Tables S2 and S3).\textsuperscript{32}

Figure 1. X-ray crystal structure of 15. C blue, O red, 1,3,5-trioxocane ring yellow-green, H atoms have been omitted by clarity. Ellipsoids are set at 50% probability.
We next focused our attention on applying the C–H functionalization to 1,3-diol derivatives. The reaction of 6\textsuperscript{I,III}-dial-\textbeta-CD 17\textsuperscript{17a} under the usual conditions produced one compound derived from the mono-abstraction, the acetate 18 in 21% yield together with four compounds in 47% overall yield arising from double abstraction: the diacetate 19 with incorporation of two external nucleophiles, the two possible mono-trioxocane–mono-acetyl derivatives 20 and 21, and the spiro ortho ester–trioxocane 22 in which only internal nucleophiles are implicated (Scheme 3).
Curiously, the expected bis(trioxocane) was not detected in the crude reaction mixture, even after longer reaction times.

Analogously, the incorporation of the acetate has taken place with inversion at C5 and transformation of the sugar into a β-L-idose unit. This appears to be a general trend for all compounds even in the cases of compounds 13 and 19 where two units in a \( ^{1}C_{4} \) inverted chair conformation exist as part of the heptameric ring.

On the other hand, the positional isomers 20 and 21 can be differentiated by a careful study using 2D HSQC and HMBC and 1D TOCSY and ROESY experiments. The H1\(^{I}\) and H4\(^{II}\) 1D ROESY correlation that has been observed in both compounds can be used to distinguish between both isomers. The H4\(^{II}\) protons, that have been clearly identified by 1D TOCSY, appear at 3.90 ppm (d, \( ^{3}J_{3\,II\,4\,II} = 9.0 \) Hz) for compound 20 and at somewhat lower field 4.11 ppm (d, \( ^{3}J_{3\,II\,4\,II} = 2.9 \) Hz) for the isomer 21, which undoubtedly confirms the proposed structures.

Moreover, in compound 22 where two positional isomers could be a priori possible we were able to assign the correct structure, since the 1D ROESY experiment showed a correlation between H1\(^{III}\) and H4\(^{IV}\) [(3.60 ppm (dd, \( J = 9.2, 9.2 \) Hz)]. In the other hypothetical isomer (spiro ortho ester at unit I and trioxocane at unit III) the analogous ROESY interaction would have been between H1\(^{I}\) and H4\(^{II}\) near the trioxocane and this last proton should be a doublet. Although two stereoisomers at the spiro quaternary carbon at C1\(^{II}\) are possible, only one has been isolated and the configuration tentatively assigned as R. Molecular mechanics calculations show that the R-isomer is more stable by 6 kcal/mol than the S-isomer. The stereochemistry is supposed to be stereoelectronically controlled, in the R-isomer the alcohol approaches the oxacarbenium ion by the α-axial direction to maximize the anomeric effect.\(^{33}\) Additional support was secured by examination of the \(^{1}H\) NMR spectrum, revealing a downfield displacement for the pro-S H6\(^{III}\) proton [4.24 (dd, \( J = 9.1, 7.9 \) Hz)]
Hz), which in this stereoisomer presents a 1,3-diaxial interaction with the C1\textsuperscript{II}–O\textsuperscript{II} bond. An obvious experiment that could be important for the determination of this stereochemistry such as the study of NOE correlations from H2\textsuperscript{II} could not be performed due to signal overlap. Fortunately, this problem does not exist in the \textsuperscript{1}H NMR spectrum of the analogous spiro ortho ester 30 and the results confirm the proposed stereochemistry (vide infra).

In contrast with the previous results, when the reaction is performed with the hitherto unknown 6\textsuperscript{I,III}-dial-\alpha-CD 23, we only obtained the bis- and the two possible mono-trioxocanes 24, 25 and 26 respectively, in 66% overall yield, not detecting products with inverted \beta-L-idose units in the molecule (Scheme 4). This is in agreement with the results obtained in the 6\textsuperscript{I,IV}-dial series where we did not find inverted sugar units in the reaction of 6\textsuperscript{I,IV}-dial-\alpha-CD 14 (Scheme 2).

**Scheme 4. HAT for the 6\textsuperscript{I,III}-dial-\alpha-CD 23**
Although, all structures were analogously determined by NMR spectroscopy the preparation of the acetyl derivatives 27 and 28 from both mono-trioxocanes was necessary to differentiate these positional isomers. The NOE interaction between H1\textsuperscript{I} and H4\textsuperscript{II} [4.26 ppm (d, J = 9.2 Hz)] in compound 27 and the corresponding correlation between H4\textsuperscript{VI} and H1\textsuperscript{V} [5.04 ppm (d, J = 3.2 Hz)] in the isomer 28 firmly established the proposed structures.

Finally, to complete the study of the diols, the reactions of 1,2-diols of β- and α-CDs were carried out. The reaction of the 6\textsuperscript{I,II}-diol-β-CD 29\textsuperscript{17a} under the usual oxidative conditions led to the formation of four compounds in moderate overall yield: the spiro ortho ester–trioxocane 30 (5%), the ten-membered lactone 31 (10%) and a mixture of two products, 32 and 33, which after acetylation was readily separated as the mono-trioxocane 34 and the δ-lactone 35, in 22% and 11% yield respectively (Scheme 5). The relative position of the spiro ortho ester with respect to the trioxocane rings in 30 and the primary acetate with respect to the trioxocane in 34 was readily established by spectroscopic means. The ring coupling constants for the sugar units involved were determined by 1D TOCSY and ROESY experiments but in any case, the alternative structures for the positional isomers should have a very different \textsuperscript{1}H and \textsuperscript{13}C NMR pattern. The spiro quaternary carbon stereochemistry C1\textsuperscript{I} in compound 30 was assigned as R on the basis discussed above for analogous spiro ortho ester 22. The absence of NOE interactions between H2\textsuperscript{I} and the pro-S H6\textsuperscript{II}, which was expected for the S-isomer, provides additional support for the anomeric configuration as R.

The ten-membered lactone 31 possesses a one-carbon dehomologated skeleton and among its more important structural and functional features are: the presence of carboxyl and acetal functions and the disappearance of one of the anomeric carbons of the starting β-CD. The protons of the sugar residues I and II can be differentiated by 1D TOCSY and all the above appears to confirm the
decanolide structure proposed, while additional support is provided by HMBC 1,3-correlations, being of special significance that observed between the carboxyl carbon at C5\textsuperscript{I} and the two protons at H6\textsuperscript{II}.

**Scheme 5. HAT for the 6\textsuperscript{I,II}-diol-\(\beta\)-CD 29**

The \(\delta\)-lactone 35 has also a one carbon dehomologated skeleton and the more notable features of its structure are the large deshielding observed for the chemical shift of the anomeric hydrogen at H\textsuperscript{I} [5.56 ppm (d, J = 3.2 Hz)] and the 1,3-HMBC correlation of this proton with the carboxylic carbon at C5\textsuperscript{I}.

According to these results, the formation of all products described in Scheme 5 could be explained in terms of which of the two primary alkoxyl radicals (6\textsuperscript{I}-O-yl or 6\textsuperscript{II}-O-yl) is generated first. In this sense, compound 30 should be formed by the 6\textsuperscript{I}-O-yl radical 1,8-abstraction in the first place followed by a second 1,6-abstraction of the 6\textsuperscript{II}-O-yl radical to generate the spiro ortho ester. Clearly, the primary alcohol 32 should be an intermediate in the formation of 30. On the other
hand, the mechanism for the formation of lactones 31 and 33 from diol 29 certainly implies a prior abstraction of hydrogen at H5\textsuperscript{I} by the 6\textsuperscript{II}-O-yl radical to give the trioxocane VIII (Scheme 6). The excess of reagent generates a new 6\textsuperscript{I}-O-yl radical from the free primary alcohol which by β-scission with loss of formaldehyde produces a C5\textsuperscript{I} tertiary radical IX.\textsuperscript{34} This intermediate evolves following two competitive paths: (i) it may experience a second β-scission of the C1\textsuperscript{I}–O\textsuperscript{I} bond leading to the C1\textsuperscript{I}-radical X, which by one-electron oxidation and nucleophilic attack by the acetate anion finally gives the ten-membered lactone 31 and (ii) it can be directly stabilized by oxidation–nucleophilic attack to give the δ-lactone 33 previous hydrolysis of ortho ester intermediate XI. Neither intermediate VIII nor XI could be detected in the crude reaction mixture.

**Scheme 6. Proposed mechanistic pathway for the formation of lactones 31 and 33**

Analogously to the β-CD derivative, the C–H functionalization of the 6\textsuperscript{II}-dial-α-CD 36\textsuperscript{35} proceeded in a similar way, giving the corresponding ten-membered lactone 37, the mono-trioxocane 38 and the δ-lactone 39 in 51\% overall yield (Scheme 7). All these compounds show NMR features similar to those of the respective compounds 31, 34 and 35 and their formation can be explained by a mechanism identical with that described for the β-CD model (Scheme 6).
Although, as commented previously, the formation of the 1,2-bis(trioxocane)s in α- and β-CDs may be thermodynamically possible, we have been unable to detect any of them. Therefore, the possibility of a second 1,8-HAT on the logical precursors, the isolated mono-trioxocanes 32 and 38, was studied by molecular mechanics. In the minimized structures of these compounds the distances between the 6\textsuperscript{II}-O-yl radical and the extractable hydrogens at H5\textsuperscript{I} are excessively long (4.7 and 5.5 Å, respectively) and 1,8-HAT reactions should be clearly unfavorable (see Figure S2 and Table S4 in the SI for more details). Notwithstanding, the alternative 1,6-HAT of the H1\textsuperscript{I} appears possible in the case of compound 32 (dO6\textsuperscript{II}–H1\textsuperscript{I} = 2.7 Å) that would explain the formation of spiro ortho ester 30 in the β-CD series. This 1,6-HAT reaction is very unlikely in the α-CD series, since in 38 a calculated distance O6\textsuperscript{II}–H1\textsuperscript{I} of 4.6 Å would prevent the formation of the corresponding spiro ortho ester.

Scheme 7. HAT for the 6\textsuperscript{II,III}-diol-α-CD 36
To gain insight into the reaction mechanism and clarify the different behavior of the 6\(^{\text{II}}\)-diols in the C–H functionalization with respect to the other positional isomers, we carried out the process with the corresponding diol in a more simple and conformationally flexible molecule such as the hexamethyl \(\beta\)-maltose derivative 41 obtained in three steps from D-(+)-maltose through the silyl-ether derivatives 40\(\alpha\) and 40\(\beta\) (Scheme 8). In this case, the reaction generated a complex mixture from which we were able to isolate the ten-membered lactone 42 together with compound 43 in only 20% overall yield. Both compounds were found to be unstable toward chromatographic purification. This instability does not prevent complete characterization but is presumably responsible for the small yield obtained.

**Scheme 8. HAT for the 6\(^{\text{II}}\)-dil-\(\beta\)-maltose 41**

The structure and stereochemistry of the ten-membered lactone 42, where all ring protons are directly observable in the \(^1\)H NMR spectrum, can be now more conveniently and unambiguously studied. The NMR data obtained are in quite good agreement with those previously described for the ten-membered lactones 31 and 37, which had been obtained mostly using 1D TOCSY. The
presence of NOE interactions between H1\textsuperscript{I} and all the protons positioned on the β-face of the decanolide ring (H2\textsubscript{I}, H4\textsubscript{I}, H4\textsubscript{II}, and pro-S H6\textsubscript{II}) seems to indicate a stereoselective α-attack of the nucleophile and an R configuration for the C1\textsuperscript{I}. The relative position of the involved ring protons and the $^3\text{J}_{\text{H},\text{H}}$ coupling constants has been checked on a minimized structure where the ten-membered lactone adopts a boat-chair-chair (BCC) conformation with an (R)-configuration for the C1\textsuperscript{I}.\textsuperscript{36}

While the decanolide formation could be explained by the mechanism shown above, the new disaccharide \textsuperscript{43} could arise from initial 1,6-hydrogen abstraction by the 6\textsuperscript{I}-O-yl radical from the proximal methoxyl group at C4\textsuperscript{I} followed by oxidation and cyclization to form a 1,3-dioxane ring.\textsuperscript{37} Then, a second 1,8-abstraction now by the 6\textsuperscript{II}-O-yl radical generates a C5\textsuperscript{I}-radical which finally collapses to the oxacarbenium ion and the incorporation of acetate with inversion of configuration. Also in this case, the coupling constants for the vicinal ring protons account for a preferred $^1$C\textsubscript{4} chair conformation for the L-idose unit. The occurrence of this compound sheds some light on the mechanism, the less hindered environment permitting this second 1,8-abstraction which in neither case was observed when the more restricted 6\textsuperscript{I,II}-diols of β- and α-CDs were used, namely compounds \textsuperscript{29} (Scheme 5) and \textsuperscript{36} (Scheme 7).

Until now, only mono- and di-alcohols have been investigated. We thought that this study would not be complete without trying the reaction with per-6-ol models derived from β- and α-CDs, \textsuperscript{2}\textsuperscript{I-VII,3}\textsuperscript{I-VII}-tetradeca-O-methyl-β-cyclomaltoheptaose\textsuperscript{38,5f} and \textsuperscript{2}\textsuperscript{I-VI,3}\textsuperscript{I-VI}-dodeca-O-methyl-α-cyclomaltohexaose,\textsuperscript{39,5f} to find out if we can access to polyfunctionalized compounds through multiple radical abstractions. In both substrates, several different conditions were screened in which the reagents stoichiometry, temperature, and time were varied, to no avail, only intractable
mixtures of products being observed in all instances. Such a result is not very surprising if we consider the many possibilities of abstraction and the presumed instability of the products formed.

**CONCLUSIONS**

In summary, we have successfully applied for the first time the remote C–H functionalization logic to CD systems. The process is initiated by the 6\textsuperscript{I}-O-yl radical which abstracts with complete regioselectivity the hydrogen atom at C5\textsuperscript{II} located in the adjacent D-glucose by a favored geometrically restricted nine-membered transition state. Using this methodology with monoalcohols derived from α-, β-, and γ-CDs two types of structurally modified CDs can be obtained in moderate to good yields: a) a new stable 1,3,5-trioxocane ring is formed between the two glucopyranose units involved and b) one of the \textsuperscript{4}C\textsubscript{1} glucopyranose residues has been transformed into a \textsuperscript{1}C\textsubscript{4} β-L-idose unit by inversion of configuration at C5\textsuperscript{II}. Interestingly, in the case of α-CD derivative 7, the conformation of the β-L-idose ring is solvent-dependent, and the \textsuperscript{1}C\textsubscript{4} conformer is favored in CDCl\textsubscript{3} while the equilibrium is strongly shifted toward the \textsuperscript{4}C\textsubscript{1} chair in C\textsubscript{6}D\textsubscript{6} solution. This phenomenon that can be readily detected at 26 °C by 1D TOCSY experiments is not observable in β- and γ-CD analogs 3, 4, and 10.\textsuperscript{40}

This methodology was also applied to 6\textsuperscript{I,II}, 6\textsuperscript{I,III}, and 6\textsuperscript{I,IV}-diols of α- and β-CDs with the intention of assessing the viability of two hydrogen abstractions in a simultaneous or tandem fashion, a hitherto unknown process.\textsuperscript{27}

The 6\textsuperscript{I,IV}-dial-β-CD derivative 11 behaved as if the two primary alcohols were independent, the expected products with two trioxocane rings 12 and two inverted β-L-idose units 13 being formed in good overall yield (Scheme 2). In the reaction of 6\textsuperscript{I,IV}-dial-α-CD derivative 14 a bis(trioxocane)
15 was also formed. However, in this more restricted hexameric ring, the corresponding products with one or two β-L-idose units were not detected.

Among the products that can be generated by a double 1,8-abstraction in the reaction of 6\textsuperscript{I\textsc{ili}}-dior-β-CD 17, three (19, 20 and 21) out of four possible have been obtained (Scheme 3). The only one that could not be isolated was the corresponding bis(trioxocane). Instead, the trioxocane–spiro ortho ester 22, formed also by a double abstraction, this time 1,8- followed by 1,6-HAT, was obtained. Unexpectedly, any attempt to cyclize the plausible intermediates 19-21 by acid catalysis failed to produce the bis(trioxocane). A mixture of chromatographically more polar unstable compounds presumably containing CD ring fragmentation products was obtained in all cases.

In sharp contrast to the situation encountered above with 6\textsuperscript{I\textsc{ili}}-dior-β-CD 17, the reaction of 6\textsuperscript{I\textsc{ili}}-dior-α-CD 23 led to the bis(trioxocane) 24 as the major product. (Scheme 4). Apart from the acid instability of the intermediates, we have no satisfactory explanation at the moment to account for the fact that the 1,3-bis(trioxocane) is easily formed in the α-CD and not in the β-CD reaction.

In the case of the 6\textsuperscript{II}-dior- of β- and α-CD, 29 and 36 respectively, the proximity of the two alkoxyl radicals in adjacent sugar units, facilitates the possibility of interaction between them. This situation which has not been presented in the previous diols is responsible for the formation of the ten-membered lactones (31 and 37) and the δ-lactones (35 and 39). These compounds are obtained by the proposed mechanism (vide supra) where the two alkoxyl radicals are involved in the formation of the lactonic rings (Schemes 5, 6, and 7).

To the best of our knowledge, this is the first time that a radical protocol is applied to create structural differentiation on the primary face of CDs. Moreover, the mild reaction conditions and the good efficiency are remarkable features that make this process a powerful tool to access
modified CDs and obtain macrocyclic rings with a different range of functionalization, otherwise difficult to achieve, and with potential synthetic and pharmaceutical applications.

**EXPERIMENTAL SECTION**

**General Experimental Methods.** Melting points were measured on a hot-stage apparatus. Optical rotations were recorded on a polarimeter at a wavelength of 589 nm at room temperature in CHCl$_3$ solutions. IR spectra were recorded on a FT-IR spectrophotometer in film. $^1$H NMR spectra were determined at 500 MHz in CDCl$_3$ or C$_6$D$_6$. Chemical shifts are reported in parts per million (ppm) and are calibrated to residual solvent peaks (CHCl$_3$ 7.26 ppm and C$_6$H$_6$ 7.15 ppm). $^{13}$C NMR spectra were determined at 125.7 MHz in CDCl$_3$ or C$_6$D$_6$. Chemical shifts are reported in parts per million (ppm) and are calibrated to residual solvent peaks (CHCl$_3$ 77.0 ppm and C$_6$H$_6$ 128.0 ppm). NMR peaks assignments and stereochemistries have been established using COSY, TOCSY, DEPT, HMBC, HSQC, and ROESY experiments. Low and high resolution mass spectra were recorded with a TOF analyzer spectrometer by using electrospray (ESI$^+$). Flash column chromatography was performed on Merck silica gel 60 PF (0.063–0.2 mm). Sensitive compounds were purified by medium-pressure column chromatography using TLC silica gel 60 F$_{254}$, scraped from Merck Millipore aluminum sheets (Product No. 1055540001), as adsorbent. Reaction progress was monitored by thin-layer chromatography (TLC) carried out on 0.25 mm coated commercial silica gel plates impregnated with a fluorescent indicator (254 nm). The spray reagents for TLC analysis were conducted with 0.5% vanillin in H$_2$SO$_4$–EtOH (4:1) and further heating until development of color. Commercially available reagents and solvents were analytical grade or were purified by standard procedures prior to use.

Oxidative HAT of 2$^{1-}$VII, 3$^{1-}$VII, 6$^{1-}$VII-icosa-O-methyl-$\beta$-cyclomaltoheptaose (1) with DIB and iodine. Method A: A solution of alcohol 1$^{17b}$ (50 mg, 0.035 mmol) in dry CH$_2$Cl$_2$ (2.4 mL)
containing DIB (33.4 mg, 0.105 mmol) and I₂ (15.5 mg, 0.06 mmol) was stirred under nitrogen at 30 °C for 0.75 h while irradiated with two 80 W tungsten-filament lamps. An excess of solid Na₂S₂O₃ was then added and stirring continued until complete disappearance of the iodine color. The reaction mixture was then filtered and concentrated under reduced pressure. Silica gel [Merck 60 PF (0.063–0.2 mm)] column chromatography of the reaction residue (hexanes–acetone, 65:35) afforded cyclo-5VII,6I-anhydro-(5VIIR)-(2,3,6-tri-O-methyl-α-D-xylo-hexos-5-ulopyranosyl)-(1→4)-2,3-di-O-methyl-α-D-glucopyranosyl-[{(1→4)-2,3,6-tri-O-methyl-α-D-glucopyranosyl}₅ (2) (30 mg, 0.021 mmol, 60%): colorless oil, [α]D +138.5 (c 1.78, CHCl₃); IR (film): 2929, 1143, 1107, 1042 cm⁻¹; ¹H NMR (500 MHz, CDCl₃): δ 3.365 (s, 3H), 3.371 (s, 9H), 3.376 (s, 3H), 3.384 (s, 3H), 3.461 (s, 3H), 3.465 (s, 3H), 3.48 (s, 3H), 3.49 (s, 3H), 3.50 (s, 6H), 3.51 (s, 3H), 3.58 (s, 3H), 3.60 (s, 3H), 3.62 (s, 6H), 3.63 (s, 3H), 3.64 (s, 3H), 3.69 (s, 3H), 5.06 (d, J = 3.5 Hz, 1H), 5.06 (d, J = 3.5 Hz, 1H), 5.07 (d, J = 3.8 Hz, 1H), 5.08 (d, J = 3.2 Hz, 1H), 5.14 (d, J = 3.5 Hz, 1H), 5.20 (d, J = 4.4 Hz, 1H), 5.21 (d, J = 3.8 Hz, 1H); ¹³C NMR (125.7 MHz, CDCl₃): δ 57.78 (CH₃), 57.95 (CH₃), 58.04 (CH₃), 58.36 (CH₃), 58.51 (CH₃), 58.58 (CH₃), 58.86 (3 × CH₃), 59.01 (CH₃), 59.03 (3 × CH₃), 60.91 (CH₃), 61.04 (CH₃), 61.26 (CH₃), 61.37 (CH₃), 61.65 (2 × CH₃), 62.17 (CH₃), 64.81 (CH₂), 66.88 (CH), 70.74 (CH), 70.78 (CH), 70.90 (CH), 70.92 (CH₂), 70.96 (CH), 70.98 (CH), 70.98 (CH₂), 71.23 (CH₂), 71.47 (CH₂), 71.76 (CH₂), 71.94 (CH₂), 77.62 (CH), 79.18 (CH), 79.20 (CH), 79.21 (CH), 80.07 (CH), 80.07 (CH), 80.07 (CH), 80.16 (CH), 80.20 (CH), 80.20 (CH), 80.78 (CH), 81.18 (CH), 81.20 (CH), 81.46 (CH), 81.51 (CH), 81.57 (CH), 81.70 (CH), 81.81 (2 × CH), 82.06 (2 × CH), 82.21 (CH), 82.30 (CH), 82.96 (CH), 96.97 (CH), 97.98 (CH), 98.24 (CH), 99.02 (CH), 99.06 (CH), 99.15 (CH), 99.39 (CH), 101.04 (C); MS (ESI⁺-TOF): m/z (%) 1435 [(M + Na)⁺, 100]; HRMS (ESI⁺-TOF): m/z [M + Na]⁺ calcd for C₆₂H₁₀₈NaO₃₅ 1435.6569; found 1435.6605. Anal. calcd for C₆₂H₁₀₈O₃₅: C, 52.68; H, 7.70. Found: C, 52.31; H, 7.57.
Method B: A solution of alcohol 1 (100 mg, 0.071 mmol) in dry CH$_2$Cl$_2$ (2.9 mL) containing DIB (50.3 mg, 0.156 mmol) and I$_2$ (18 mg, 0.071 mmol) was stirred under nitrogen at 30 °C for 0.5 h while irradiated with two 80 W tungsten-filament lamps. The reaction mixture was then poured into 10% aqueous Na$_2$S$_2$O$_3$, extracted with CH$_2$Cl$_2$, dried over Na$_2$SO$_4$, and concentrated. The residue was purified by silica gel [Merck 60 PF (0.063–0.2 mm)] column chromatography (hexanes–acetone, 65:35) to give 2 (36.3 mg, 0.026 mmol, 36%) and cyclo-(5R)-5$^\text{VII}$-O-acetyl-2,3,6-tri-O-methyl-α-D-xylo-hexos-5-ulpopyranosyl-(1→4)-2,3-di-O-methyl-α-D-glucopyranosyl-[(1→4)-2,3,6-tri-O-methyl-α-D-glucopyranosyl]$_5$ (3) (62.9 mg, 0.043 mmol, 61%). Compound 3: colorless oil, [α]$_D$ = +134.5 (c 1.32, CHCl$_3$); IR (film): 3524, 2929, 1734, 1109, 1041 cm$^{-1}$; $^1$H NMR (500 MHz, CDCl$_3$): δ 2.06 (s, 3H), 3.14 (dd, $J = 10.1$, 3.5 Hz, 1H), 3.15 (dd, $J = 10.1$, 3.8 Hz, 1H), 3.18 (dd, $J = 9.5$, 3.2 Hz, 1H), 3.19 (dd, $J = 10.1$, 3.5 Hz, 1H), 3.20 (dd, $J = 10.1$, 3.5 Hz, 1H), 3.22 (dd, $J = 10.1$, 3.5 Hz, 1H), 3.31 (dd, $J = 9.8$, 9.8 Hz, 1H), 3.367 (s, 6H), 3.373 (s, 3H), 3.38 (s, 3H), 3.39 (s, 3H), 3.40 (s, 3H), 3.42 (s, 3H), 3.45 (s, 3H), 3.475 (s, 3H), 3.478 (s, 3H), 3.49 (s, 3H), 3.51 (s, 3H), 3.53 (s, 3H), 3.58 (s, 3H), 3.61 (s, 3H), 3.62 (s, 6H), 3.637 (s, 6H), 3.644 (s, 3H), 3.96 (d, $J = 10.1$ Hz, 1H, H$_6^\text{VII}$), 3.99 (d, $J = 9.8$ Hz, 1H, H$_6^\text{VI}^\text{II}$), 4.04 (br d, $J = 1.9$ Hz, 1H, H$_4^\text{VII}$), 4.20 (dd, $J = 2.5$, 2.5 Hz, 1H, H$_3^\text{VII}$), 4.31 (br d, $J = 11.7$ Hz, 1H), 5.01 (d, $J = 3.2$ Hz, 1H), 5.04 (d, $J = 3.2$ Hz, 1H), 5.060 (d, $J = 2.8$ Hz, 1H), 5.061 (d, $J = 4.1$ Hz, 1H), 5.12 (d, $J = 3.8$ Hz, 1H), 5.14 (d, $J = 1.9$ Hz, 1H, H$_1^\text{VII}$), 5.18 (d, $J = 3.5$ Hz, 1H); $^1$H NMR (500 MHz, CDCl$_3$), 1D-TOCSY, irradiation at H$_3^\text{VII}$, 4.20 ppm): δ 3.41 (br s, 1H, H$_2^\text{VII}$), 4.04 (br d, $J = 1.9$ Hz, 1H, H$_4^\text{VII}$), 5.14 (d, $J = 1.9$ Hz, 1H, H$_1^\text{VII}$); $^1$H NMR (500 MHz, C$_6$D$_6$): δ 1.72 (s, 3H), 3.10 (dd, $J = 9.1$, 3.2 Hz, 1H), 3.13 (dd, $J = 9.8$, 3.5 Hz, 1H), 3.19 (s, 3H), 3.20 (s, 3H), 3.26 (s, 3H), 3.27 (s, 3H), 3.28 (s, 3H), 3.30 (s, 6H), 3.31 (s, 3H), 3.32 (s, 3H), 3.39 (s, 3H), 3.40 (s, 6H), 3.41 (s, 3H), 3.46 (s, 3H), 3.60 (s, 3H), 3.68 (s, 3H), 3.74 (s, 3H), 3.75 (s, 3H), 3.76 (s, 3H), 3.80 (s, 3H), 5.07 (d, $J =
3.2 Hz, 1H), 5.10 (d, J = 2.8 Hz, 1H), 5.20 (d, J = 3.2 Hz, 1H), 5.27 (d, J = 3.8 Hz, 1H), 5.32 (d, J = 3.5 Hz, 1H), 5.45 (d, J = 3.8 Hz, 1H), 5.52 (d, J = 1.6 Hz, 1H, H1l); $^{13}$C NMR (125.7 MHz, CDCl₃): δ 22.08 (CH₃), 57.27 (CH₃), 57.45 (CH₃), 58.04 (CH₃), 58.37 (CH₃), 58.65 (CH₃), 58.68 (CH₃), 58.72 (CH₃), 58.86 (2 × CH₃), 58.93 (CH₃), 58.96 (3 × CH₃), 59.17 (CH₃), 60.16 (CH₂), 61.18 (CH₃), 61.25 (CH₃), 61.34 (CH₃), 61.45 (CH₃), 61.52 (CH₃), 61.67 (CH₃), 70.33 (CH₂, C6Ⅶ), 70.60 (CH), 70.80 (CH₂), 70.91 (CH), 70.91 (CH₂), 71.01 (CH, C4Ⅶ), 71.13 (2 × CH), 71.13 (2 × CH₂), 71.31 (CH₂), 71.35 (CH), 71.47 (CH), 75.22 (CH, C3Ⅶ), 76.64 (CH, C2Ⅶ), 79.43 (CH), 79.93 (CH), 80.61 (CH), 81.20 (CH), 81.31 (CH), 81.47 (CH), 81.59 (CH), 81.74 (2 × CH), 81.75 (CH), 81.97 (CH), 82.00 (CH), 82.08 (CH), 82.22 (CH), 82.26 (CH), 82.28 (CH), 82.30 (CH), 83.15 (CH), 97.59 (CH, C1Ⅶ), 99.25 (3 × CH), 99.52 (CH), 99.87 (CH), 100.34 (CH), 105.03 (C, C5Ⅶ), 170.19 (C); MS (ESI+ -TOF): m/z (%) 1495 [(M + Na)+, 100]; HRMS (ESI+ -TOF): m/z [M + Na]+ calcd for C₆₄H₁₁₂NaO₃₇ 1495.6780; found 1495.6727. Anal. calcd for C₆₄H₁₁₂O₃₇: C, 52.17; H, 7.66. Found: C, 52.29; H, 7.58.

Oxidative HAT of 2¹Ⅶ,3¹Ⅶ,6²Ⅶ-icosa-O-methyl-β-cyclomaltoheptaose (1) with bis(4-tert-butylbenzoyloxy)iodobenzene and iodine. A solution of alcohol 1 (30 mg, 0.021 mmol) in dry CH₂Cl₂ (0.85 mL) containing bis(4-tert-butylbenzoyloxy)iodobenzene (25.8 mg, 0.046 mmol) and I₂ (5.3 mg, 0.021 mmol) was stirred under nitrogen at 25 °C for 1.5 h while irradiated with two 80 W tungsten-filament lamps. The reaction mixture was then directly loaded onto a silica gel (TLC Silica gel 60 F₂₅₄, scraped from Merck Aluminum sheets) column chromatography (hexanes–acetone, 70:30→65:35) to give 2 (13.7 mg, 0.01 mmol, 46%).

Oxidative HAT of 2¹Ⅶ,3¹Ⅶ,6²Ⅶ-icosa-O-methyl-β-cyclomaltoheptaose (1) with bis(2,4,6-trimethylbenzoyloxy)iodobenzene and iodine. A solution of alcohol 1 (90 mg, 0.064 mmol) in dry CH₂Cl₂ (2.6 mL) containing bis(2,4,6-trimethylbenzoyloxy)iodobenzene (101 mg, 0.191 mmol)
and I₂ (16.2 mg, 0.064 mmol) was stirred under nitrogen at 28 °C for 2 h while irradiated with two 80 W tungsten-filament lamps. The reaction mixture was then directly loaded onto a silica gel (TLC Silica gel 60 F₂₅₄, scraped from Merck Aluminum sheets) column chromatography (hexanes–acetone, 70:30) to give 2 (19.3 mg, 0.014 mmol, 21%) and cyclo-(5R)-5\(^{\text{VII}}\)-O-(2,4,6-trimethylbenzoyl)-2,3,6-tri-O-methyl-\(\alpha\)-D-xylo-hexos-5-ulopyranosyl-(1→4)-2,3-di-O-methyl-\(\alpha\)-D-glucopyranosyl-[/(1→4)-2,3,6-tri-O-methyl-\(\alpha\)-D-glucopyranosyl]₅ (4) (33.1 mg, 0.021 mmol, 33%). Compound 4: amorphous solid, [\(\alpha\)]\(_D\) +132.5 (c 1.10, CHCl₃); IR (film): 2928, 1727, 1455, 1368, 1192, 1107, 1043 cm\(^{-1}\); \(^1\)H NMR (500 MHz, CDCl₃): \(\delta\) 2.26 (s, 3H), 2.36 (s, 6H), 3.14 (dd, \(J = 6.9, 3.2\) Hz, 1H), 3.17 (m, 5H), 3.21 (dd, \(J = 5.7, 3.8\) Hz, 1H), 3.32 (s, 3H), 3.37 (s, 3H), 3.38 (s, 3H), 3.39 (s, 3H), 3.40 (s, 3H), 3.41 (s, 3H), 3.44 (s, 3H), 3.48 (s, 3H), 3.48 (s, 3H), 3.49 (s, 3H), 3.51 (s, 6H), 3.58 (s, 3H), 3.62 (s, 6H), 3.64 (s, 3H), 3.65 (s, 3H), 3.66 (s, 3H), 4.36 (br d, \(J = 12.0\) Hz), 5.04 (d, \(J = 2.8\) Hz, 1H), 5.07 (m, 3H), 5.13 (d, \(J = 3.8\) Hz, 1H), 5.18 (d, \(J = 3.8\) Hz, 1H), 5.36 (d, \(J = 1.9\) Hz, 1H), 6.81 (s, 2H); \(^1\)H NMR (500 MHz, C₆D₆): \(\delta\) 2.03 (s, 3H), 2.57 (s, 6H), 3.12 (s, 3H), 3.14 (s, 3H), 3.20 (s, 3H), 3.28 (s, 3H), 3.29 (s, 3H), 3.29 (s, 3H), 3.30 (s, 3H), 3.31 (s, 3H), 3.32 (s, 3H), 3.40 (s, 3H), 3.41 (s, 3H), 3.42 (s, 3H), 3.42 (s, 3H), 3.44 (s, 3H), 3.60 (s, 3H), 3.68 (s, 3H), 3.73 (s, 3H), 3.75 (s, 3H), 3.77 (s, 3H), 3.79 (s, 3H), 4.67 (d, \(J = 9.5\) Hz, 1H), 5.09 (d, \(J = 3.2\) Hz, 1H), 5.17 (d, \(J = 2.8\) Hz, 1H), 5.21 (d, \(J = 3.5\) Hz, 1H), 5.26 (d, \(J = 3.8\) Hz, 1H), 5.33 (d, \(J = 3.5\) Hz, 1H), 5.47 (d, \(J = 3.8\) Hz, 1H), 5.74 (d, \(J = 1.9\) Hz, 1H), 6.67 (s, 2H); \(^{13}\)C NMR (125.7 MHz, CDCl₃): \(\delta\) 19.53 (2 × CH₃), 21.08 (CH₃), 57.12 (CH₃), 57.27 (CH₃), 58.07 (CH₃), 58.37 (CH₃), 58.44 (CH₃), 58.68 (CH₃), 58.70 (CH₃), 58.74 (CH₃), 58.88 (2 × CH₃), 58.95 (CH₃), 58.98 (CH₃), 58.99 (CH₃), 59.19 (CH₃), 60.51 (CH₂), 61.00 (CH₃), 61.28 (CH₃), 61.36 (CH₃), 61.50 (CH₃), 61.56 (CH₃), 61.72 (CH₃), 69.93 (CH₂), 70.62 (CH), 70.87 (CH₂), 70.91 (CH₂), 70.91 (CH), 71.04 (CH), 71.11 (CH), 71.16 (CH₂), 71.20 (CH₂), 71.40 (CH), 71.40 (CH₂), 71.51
Oxidative HAT of 2\textsuperscript{I-VI},3\textsuperscript{I-VI},6\textsuperscript{II-VI}-heptadeca-O-methyl-\(\alpha\)-cyclomaltohexaose (5). Method A: A solution of alcohol 5\textsuperscript{25} (30 mg, 0.025 mmol) in dry CH\(_2\)Cl\(_2\) (1 mL) containing DIB (17.6 mg, 0.055 mmol) and I\(_2\) (6.3 mg, 0.025 mmol) was stirred under nitrogen at 21 °C for 1 h while irradiated with two 80 W tungsten-filament lamps. An excess of solid Na\(_2\)S\(_2\)O\(_3\) was then added and stirring continued until complete disappearance of the iodine color. The reaction mixture was then filtered and concentrated under reduced pressure. Silica gel [Merck 60 PF (0.063–0.2 mm)] column chromatography of the reaction residue (hexanes–acetone, 65:35) afforded cyclo-5\textsuperscript{VI},6\textsuperscript{I}-anhydro-(5\textsuperscript{VI}R)-(2,3,6-tri-O-methyl-\(\alpha\)-D-xylo-hexos-5-ulopyranosyl)-(1\(\rightarrow\)4)-2,3-di-O-methyl-\(\alpha\)-D-glucopyranosyl-[1\(\rightarrow\)4]-2,3,6-tri-O-methyl-\(\alpha\)-D-glucopyranosyl\(]_4\) (6) (17.7 mg, 0.015 mmol, 59%) as a colorless oil: [\(\alpha\)]\(_D\) +127.6 (c 1.24, CHCl\(_3\)); IR (film): 3052, 2929, 2832 1454, 1365, 1196, 1107 cm\(^{-1}\); \(^1\)H NMR (500 MHz, CDCl\(_3\)): \(\delta\) 3.12 (dd, J = 10.1, 3.2 Hz, 1H), 3.14 (dd, J = 9.9, 3.3 Hz, 1H), 3.15 (dd, J = 9.3, 3.6 Hz, 1H), 3.18 (dd, J = 9.8, 3.8 Hz, 1H), 3.19 (dd, J = 9.8, 3.5 Hz, 1H), 3.25 (dd, J = 9.8, 3.2 Hz, 1H), 3.357 (s, 3H), 3.360 (s, 3H), 3.36 (s, 3H), 3.37 (s, 3H), 3.378 (s, 3H), 3.379 (s, 3H), 3.469 (s, 3H), 3.470 (s, 3H), 3.483 (s, 3H), 3.488 (s, 3H), 3.493 (s, 3H), 3.51 (s, 3H), 3.58 (s, 3H), 3.60 (s, 3H), 3.627 (s, 3H), 3.632 (s, 3H), 3.70 (s, 3H), 3.73 (s, 3H), 4.98 (d, J = 3.8 Hz, 1H), 5.00 (d, J = 3.5 Hz, 1H), 5.03 (d, J = 3.8 Hz, 1H), 5.04 (d, J = 3.5 Hz, 1H), 5.06 (d, J = 3.2 Hz,
$^1$H NMR (500 MHz, C$_6$D$_6$): \( \delta \) 3.15 (dd, \( J = 9.8, 3.5 \) Hz, 1H), 3.16 (dd, \( J = 9.1, 4.4 \) Hz, 1H), 3.19 (dd, \( J = 9.8, 3.8 \) Hz, 1H), 3.20 (dd, \( J = 9.9, 3.3 \) Hz, 1H), 3.24 (dd, \( J = 9.8, 3.2 \) Hz, 1H), 3.25 (s, 3H), 3.268 (s, 3H), 3.273 (s, 3H), 3.28 (s, 3H), 3.30 (s, 3H), 3.31 (s, 3H), 3.317 (s, 3H), 3.320 (s, 3H), 3.34 (s, 3H), 3.50 (s, 3H), 3.55 (dd, \( J = 8.8, 7.9 \) Hz, 1H), 3.67 (s, 3H), 3.68 (s, 3H), 3.70 (s, 3H), 3.80 (s, 3H), 3.82 (s, 3H), 3.83 (s, 3H), 4.16 (dd, \( J = 9.5, 9.5 \) Hz, 1H), 4.27 (d, \( J = 9.1 \) Hz, 1H), 4.37 (d, \( J = 10.1 \) Hz, 1H), 4.55 (ddd, \( J = 10.1, 5.0, 1.3 \) Hz, 1H), 5.02 (d, \( J = 3.8 \) Hz, 1H), 5.17 (d, \( J = 3.3 \) Hz, 1H), 5.18 (d, \( J = 3.5 \) Hz, 1H), 5.19 (d, \( J = 3.2 \) Hz, 1H), 5.22 (d, \( J = 3.5 \) Hz, 1H), 5.24 (d, \( J = 3.5 \) Hz, 1H); $^{13}$C NMR (125.7 MHz, C$_6$D$_6$): \( \delta \) 57.22 (CH$_3$), 57.70 (CH$_3$), 57.84 (CH$_3$), 57.93 (CH$_3$), 58.15 (CH$_3$), 58.17 (CH$_3$), 58.61 (CH$_3$), 58.72 (CH$_3$), 58.91 (CH$_3$), 59.00 (CH$_3$), 59.03 (CH$_3$), 61.31 (CH$_3$), 61.53 (CH$_3$), 61.58 (CH$_3$), 61.79 (CH$_3$), 62.08 (CH$_3$), 62.15 (CH$_3$), 64.19 (CH$_2$), 66.85 (CH), 71.42 (CH), 71.64 (CH), 71.67 (CH$_2$), 71.83 (CH), 72.52 (CH), 72.87 (CH$_2$), 72.92 (2 × CH$_2$), 73.10 (CH$_2$), 79.14 (CH), 81.49 (CH), 81.60 (2 × CH), 81.69 (CH), 81.76 (CH), 81.89 (CH), 82.03 (CH), 82.09 (CH), 82.35 (CH), 82.67 (CH), 82.92 (CH), 82.94 (2 × CH), 82.97 (CH), 83.34 (CH), 83.37 (CH), 83.48 (CH), 98.07 (CH), 98.58 (CH), 99.11 (CH), 99.17 (CH), 99.95 (CH), 100.42 (CH), 101.31 (C); MS (ESI$^+$-TOF): m/z (%) 1231 [(M + Na)$^+$, 100]; HRMS (ESI$^+$-TOF): m/z [M + Na]$^+$ calcd
for C₅₃H₉₂NaO₃₀ 1231.5571; found 1231.5543. Anal. calcd for C₅₃H₉₂O₃₀: C, 52.64; H, 7.67. Found: C, 52.46; H, 7.76.

Method B: A solution of alcohol 5 (45 mg, 0.037 mmol) in dry CH₂Cl₂ (1.5 mL) containing DIB (26.2 mg, 0.081 mmol) and I₂ (4.7 mg, 0.019 mmol) was stirred under nitrogen at 22 °C for 1.5 h while irradiated with two 80 W tungsten-filament lamps. The reaction mixture was then directly loaded onto a silica gel (TLC Silica gel 60 F₂₅₄, scraped from Merck Aluminum sheets) column chromatography (hexanes–acetone, 65:35) to give 6 (14 mg, 0.012 mmol, 31%) and cyclo-(5R)-5'VI-O-acetyl-2,3,6-tri-O-methyl-α-D-xylo-hexos-5-ulopyranosyl-(1→4)-2,3-di-O-methyl-α-D-glucopyranosyl-[((1→4)-2,3,6-tri-O-methyl-α-D-glucopyranosyl)₄ (7) (13.9 mg, 0.011 mmol, 30%).

Compound 7: colorless oil, [α]D +127.8 (c 0.90, CHCl₃); IR (film): 3509, 2933, 1758, 1107, 1041 cm⁻¹; ¹H NMR (500 MHz, CDCl₃): δ 2.05 (s, 3H), 3.15 (dd, J = 10.1, 3.2 Hz, 1H), 3.15 (dd, J = 10.1, 3.2 Hz, 1H), 3.16 (dd, J = 9.8, 3.5 Hz, 1H), 3.17 (dd, J = 10.1, 3.5 Hz, 1H), 3.21 (dd, J = 9.5, 3.2 Hz, 1H), 3.36 (s, 3H), 3.368 (s, 3H), 3.373 (s, 6H), 3.38 (s, 3H), 3.43 (s, 3H), 3.456 (s, 3H), 3.464 (s, 3H), 3.48 (s, 3H), 3.50 (s, 3H), 3.52 (s, 3H), 3.607 (s, 3H), 3.609 (s, 3H), 3.613 (s, 3H), 3.62 (s, 3H), 3.64 (s, 3H), 3.68 (s, 3H), 3.91 (d, J = 10.4 Hz, 1H), 3.98 (dd, J = 10.1, 3.2 Hz, 1H), 4.02 (d, J = 10.4 Hz, 1H), 4.21 (d, J = 3.2 Hz, 1H), 4.23 (m, 1H), 4.97 (d, J = 3.5 Hz, 1H), 5.02 (d, J = 3.5 Hz, 1H), 5.03 (d, J = 3.8 Hz, 1H), 5.05 (d, J = 3.2 Hz, 1H), 5.06 (d, J = 3.5 Hz, 1H), 5.23 (d, J = 2.8 Hz, 1H); ¹H NMR (500 MHz, CDCl₃, 1D-TOCSY, irradiation at H1VI, 5.23 ppm): δ 3.43 (br s, 1H, H2VI), 3.99 (dd, J = 3.2, 3.2 Hz, 1H, H3VI), 4.21 (d, J = 2.8 Hz, 1H, H4VI); ¹H NMR (500 MHz, C₆D₆): δ 1.89 (s, 3H), 3.12 (dd, J = 9.6, 3.3 Hz, 1H), 3.15 (dd, J = 9.8, 3.2 Hz, 1H), 3.24 (s, 3H), 3.25 (s, 3H), 3.28 (s, 6H), 3.30 (s, 3H), 3.32 (s, 3H), 3.33 (s, 3H), 3.36 (s, 3H), 3.39 (s, 3H), 3.42 (s, 3H), 3.50 (s, 3H), 3.61 (s, 3H), 3.70 (s, 3H), 3.73 (s, 3H), 3.76 (s, 3H), 3.84 (s, 3H), 3.87 (s, 3H), 4.74 (d, J = 7.9 Hz, 1H), 5.04 (d, J = 3.2 Hz, 1H), 5.12 (d, J = 3.2 Hz, 1H), 5.19
(d, J = 3.5 Hz, 1H), 5.20 (d, J = 3.8 Hz, 1H), 5.22 (d, J = 3.5 Hz, 1H), 5.27 (d, J = 3.5 Hz, 1H); \( ^1\text{H} \) NMR (500 MHz, C\(_6\)D\(_6\), 1D-TOCSY, irradiation at H\(_1\)\(^{VI}\), 5.27 ppm): \( \delta \) 3.49 (dd, \( J = 8.5, 2.8 \) Hz, 1H, H\(_2\)\(^{VI}\)), 3.73 (dd, \( J = 7.9, 7.9 \) Hz, 1H, H\(_3\)\(^{VI}\)); \( ^{13}\text{C} \) NMR (125.7 MHz, CDCl\(_3\)): \( \delta \) 21.88 (CH\(_3\)), 57.27 (CH\(_3\)), 57.74 (CH\(_3\)), 57.80 (CH\(_3\)), 58.11 (CH\(_3\)), 58.33 (CH\(_3\)), 58.36 (CH\(_3\)), 58.71 (CH\(_3\)), 58.82 (CH\(_3\)), 58.86 (CH\(_3\)), 58.93 (CH\(_3\)), 59.04 (CH\(_3\)), 59.08 (CH\(_3\)), 61.21 (CH\(_2\)), 61.48 (CH\(_3\)), 61.62 (CH\(_3\)), 61.66 (CH\(_3\)), 61.77 (CH\(_3\)), 61.92 (CH\(_3\)), 70.84 (CH), 70.92 (CH), 71.14 (CH), 71.16 (CH), 71.35 (2 \( \times \) CH\(_2\)), 71.68 (CH), 71.79 (CH), 71.83 (CH\(_2\)), 72.17 (CH), 74.04 (CH), 76.44 (CH), 77.31 (CH), 80.57 (CH), 81.10 (CH), 81.21 (CH), 81.41 (CH), 81.49 (CH), 81.54 (CH), 81.57 (CH), 81.92 (CH), 82.19 (CH), 82.25 (CH), 82.33 (2 \( \times \) CH), 82.40 (CH), 82.57 (CH), 82.65 (CH), 97.26 (CH), 98.28 (CH), 99.78 (CH), 99.89 (CH), 100.41 (CH), 101.39 (CH), 103.99 (C), 169.76 (C); \( ^{13}\text{C} \) NMR (125.7 MHz, C\(_6\)D\(_6\)): \( \delta \) 22.36 (CH\(_3\)), 57.35 (CH\(_3\)), 57.54 (CH\(_3\)), 57.81 (CH\(_3\)), 57.83 (CH\(_3\)), 58.22 (2 \( \times \) CH\(_3\)), 58.80 (CH\(_3\)), 58.82 (CH\(_3\)), 58.93 (CH\(_3\)), 59.01 (CH\(_3\)), 59.87 (CH\(_3\)), 60.41 (CH\(_3\)), 61.60 (CH\(_3\)), 61.71 (CH\(_3\)), 61.75 (CH\(_3\)), 61.79 (CH\(_3\)), 61.80 (CH\(_3\)), 62.77 (CH\(_2\)), 71.94 (CH), 72.01 (CH), 72.01 (CH\(_2\)), 72.17 (2 \( \times \) CH\(_2\)), 72.31 (CH), 72.60 (CH\(_2\)), 72.60 (CH), 73.22 (CH\(_2\)), 73.39 (CH), 79.60 (CH), 80.56 (CH), 80.96 (CH), 81.10 (CH), 81.94 (CH), 81.98 (CH), 82.28 (CH), 82.39 (CH), 82.41 (CH), 82.57 (2 \( \times \) CH), 82.60 (CH), 82.73 (CH), 82.94 (CH), 83.09 (CH), 83.16 (CH), 83.26 (2 \( \times \) CH), 98.68 (CH), 99.22 (CH), 99.58 (CH), 99.99 (CH), 100.28 (CH), 100.62 (CH), 104.57 (C), 169.11 (C); MS (ESI\(^+\)-TOF): \( m/z \) (\%) 1291 [(M + Na\(^+\), 100]; HRMS (ESI\(^+\)-TOF): \( m/z \) [M + Na\(^+\)] calcd for C\(_{55}\)H\(_{96}\)NaO\(_{32}\) 1291.5782; found 1291.5819. Anal. calcd for C\(_{55}\)H\(_{96}\)O\(_{32}\): C, 52.04; H, 7.62. Found: C, 52.25; H, 7.49.

Oxidative HAT of 2\(^{I-VIII}\),3\(^{I-VIII}\),6\(^{II-VIII}\)-tricosa-O-methyl-\( \gamma \)-cyclomaltooctaose (8). Method A: A solution of alcohol 8\(^{36}\) (31 mg, 0.019 mmol) in dry CH\(_2\)Cl\(_2\) (0.78 mL) containing DIB (113.5 mg, 0.042 mmol) and I\(_2\) (4.8 mg, 0.019 mmol) was stirred under nitrogen at 25 \(^\circ\)C for 1 h while
irradiated with two 80 W tungsten-filament lamps. An excess of solid Na$_2$S$_2$O$_3$ was then added and stirring continued until complete disappearance of the iodine color. The reaction mixture was then filtered and concentrated under reduced pressure. Silica gel [Merck 60 PF (0.063–0.2 mm)] column chromatography of the reaction residue (hexanes–acetone, 60:40) afforded cyclo-$\text{VIII,6}$-anhydro-($\text{VIII}R$)-(2,3,6-tri-$O$-methyl-$\alpha$-$D$-xylo-hexos-5-ulopyranosyl)-(1→4)-2,3-di-$O$-methyl-$\alpha$-$D$-glucopyranosyl-[(1→4)-2,3,6-tri-$O$-methyl-$\alpha$-$D$-glucopyranosyl]$\text{6}$ (9) (13.6 mg, 0.008 mmol, 44%): colorless oil, $[\alpha]_D^0 +130.9$ (c 1.380, CHCl$_3$); IR (film): 2929, 1141, 1104, 1039 cm$^{-1}$; $^1$H NMR (500 MHz, CDCl$_3$): $\delta$ 3.14–3.29 (m, 8H), 3.34 (s, 3H), 3.36 (s, 3H), 3.37 (s, 3H), 3.377 (s, 3H), 3.379 (s, 3H), 3.39 (s, 3H), 3.459 (s, 3H), 3.464 (s, 3H), 3.47 (s, 3H), 3.48 (s, 3H), 3.491 (s, 3H), 3.495 (s, 6H), 3.50 (s, 3H), 3.51 (s, 3H), 3.54 (s, 3H), 3.58 (s, 3H), 3.59 (s, 3H), 3.603 (s, 3H), 3.606 (s, 3H), 3.611 (s, 3H), 3.639 (s, 3H), 3.643 (s, 3H), 5.08 (d, $J = 3.8$ Hz, 1H), 5.18 (d, $J = 3.2$ Hz, 1H), 5.20 (d, $J = 3.8$ Hz, 1H), 5.21 (d, $J = 3.8$ Hz, 1H), 5.22 (d, $J = 4.1$ Hz, 1H), 5.24 (d, $J = 2.2$ Hz, 1H), 5.25 (d, $J = 2.8$ Hz, 1H), 5.58 (d, $J = 3.5$ Hz, 1H); $^{13}$C NMR (125.7 MHz, CDCl$_3$): $\delta$ 57.46 (CH$_3$), 57.74 (CH$_3$), 58.34 (CH$_3$), 58.41 (CH$_3$), 58.48 (CH$_3$), 58.51 (CH$_3$), 58.85 (2 × CH$_3$), 58.88 (CH$_3$), 58.93 (2 × CH$_3$), 58.96 (CH$_3$), 58.99 (CH$_3$), 59.20 (2 × CH$_3$), 59.60 (CH$_3$), 59.99 (CH$_3$), 60.37 (CH$_3$), 60.98 (CH$_3$), 61.02 (2 × CH$_3$), 61.20 (CH$_3$), 61.70 (CH$_3$), 64.44 (CH$_2$), 66.25 (CH), 69.00 (CH), 69.90 (CH), 70.18 (CH), 70.22 (CH$_2$), 70.55 (CH), 70.78 (CH), 70.91 (CH), 71.04 (CH$_2$), 71.08 (2 × CH$_2$), 71.21 (CH), 71.64 (CH$_2$), 71.94 (CH$_2$), 72.18 (CH$_2$), 74.47 (CH), 77.13 (CH), 77.62 (CH), 77.92 (CH), 78.05 (CH), 78.58 (CH), 79.62 (CH), 79.72 (CH), 80.67 (CH), 81.17 (CH), 81.58 (CH), 81.63 (CH), 81.69 (3 × CH), 81.81 (CH), 82.28 (CH), 82.31 (CH), 82.46 (CH), 82.61 (CH), 82.76 (CH), 83.02 (CH), 83.18 (CH), 92.88 (CH), 95.20 (CH), 96.85 (CH), 97.59 (CH), 97.84 (CH), 98.35 (CH), 98.82 (CH), 99.55 (CH), 100.64 (C); MS (ESI$^+$-TOF): $m/z$

Method B: A solution of alcohol 8 (45 mg, 0.028 mmol) in dry CH2Cl2 (1.2 mL) containing DIB (20 mg, 0.062 mmol) and I2 (3.6 mg, 0.014 mmol) was stirred under nitrogen at 24 °C for 2.5 h while irradiated with two 80 W tungsten-filament lamps. The reaction mixture was then directly loaded onto a silica gel (TLC Silica gel 60 F254, scraped from Merck Aluminum sheets) column chromatography (hexanes–acetone, 65:35) to give 9 (15.6 mg, 0.01 mmol, 36%) and cyclo-(5R)-5VIII-O-acetyl-2,3,6-tri-O-methyl-α-D-xylo-hexos-5-ulopyranosyl-(1→4)-2,3-di-O-methyl-α-D-glucopyranosyl-[(1→4)-2,3,6-tri-O-methyl-α-D-glucopyranosyl]6 (10) (19.2 mg, 0.011 mmol, 41%). Compound 10: colorless oil, [α]D +131.6 (c 0.91, CHCl3); IR (film): 3539, 2925, 1738, 1152, 1100, 1037 cm⁻¹; 1H NMR (500 MHz, CDCl3): δ 2.06 (s, 3H), 3.16 (dd, J = 9.5, 3.2 Hz, 1H), 3.19 (dd, J = 9.8, 3.5 Hz, 1H), 3.19 (dd, J = 9.8, 3.5 Hz, 1H), 3.21 (dd, J = 9.6, 3.6 Hz, 1H), 3.23 (dd, J = 9.5, 3.8 Hz, 1H), 3.26 (dd, J = 9.8, 3.8 Hz, 1H), 3.30 (s, 3H), 3.334 (s, 3H), 3.34 (s, 3H), 3.36 (s, 3H), 3.374 (s, 3H), 3.40 (s, 3H), 3.42 (s, 3H), 3.44 (br d, J = 3.9 Hz, 1H, H2VIII), 3.473 (s, 6H), 3.476 (s, 3H), 3.477 (s, 3H), 3.51 (s, 3H), 3.556 (s, 3H), 3.558 (s, 3H), 3.58 (s, 3H), 3.60 (s, 6H), 3.62 (s, 3H), 3.64 (s, 3H), 3.649 (s, 3H), 3.654 (s, 3H), 3.71 (s, 3H), 3.99 (d, J = 9.8 Hz, 1H), 4.025 (d, J = 9.8 Hz, 1H), 4.03 (br s, 1H, H4VIII), 4.24 (dd, J = 2.2, 2.2 Hz, 1H, H3VIII), 5.01 (d, J = 3.5 Hz, 1H), 5.08 (d, J = 3.2 Hz, 1H), 5.09 (d, J = 3.5 Hz, 1H), 5.12 (d, J = 3.8 Hz, 1H), 5.18 (d, J = 3.8 Hz, 1H), 5.19 (d, J = 3.5 Hz, 1H), 5.23 (d, J = 1.6 Hz, 1H, H1VIII), 5.60 (d, J = 4.1 Hz, 1H); 1H NMR (500 MHz, CDCl3, 1D-TOCSY, irradiation at H1II, 5.23 ppm): δ 3.45 (d, J = 3.9 Hz, 1H, H2VIII), 4.03 (br s, 1H, H4VIII), 4.25 (dd, J = 2.4, 2.4 Hz, 1H, H3VIII); 1H NMR (500 MHz, C6D6): δ 1.77 (s, 3H), 3.10 (dd, J = 9.5, 2.8 Hz, 1H), 3.13 (dd, J = 9.9, 3.3 Hz, 1H), 3.19 (s, 3H), 3.21 (s, 3H), 3.22 (s, 3H), 3.27 (s, 3H), 3.29 (m, 6H), 3.296
(s, 3H), 3.304 (s, 3H), 3.31 (s, 3H), 3.322 (s, 3H), 3.325 (s, 3H), 3.35 (s, 3H), 3.36 (s, 3H), 3.37 (s, 3H), 3.39 (s, 3H), 3.46 (s, 3H), 3.59 (s, 3H), 3.65 (s, 3H), 3.70 (s, 3H), 3.72 (s, 3H), 3.726 (s, 3H), 3.733 (s, 3H), 3.81 (s, 3H), 4.30 (dd, J = 2.5, 2.5 Hz, 1H), 4.32 (br s), 4.40 (d, J = 9.8 Hz, 1H), 4.46 (br d, J = 12.0 Hz, 1H), 4.51 (d, J = 9.8 Hz, 1H), 5.16 (d, J = 3.2 Hz, 1H), 5.17 (d, J = 3.2 Hz, 1H), 5.29 (d, J = 3.5 Hz, 1H), 5.36 (d, J = 3.8 Hz, 1H), 5.37 (d, J = 3.8 Hz, 1H), 5.57 (d, J = 3.8 Hz, 1H), 5.61 (d, J = 3.8 Hz, 1H), 5.65 (d, J = 1.6 Hz, 1H); $^{13}$C NMR (125.7 MHz, CDCl$_3$): $\delta$ 22.11 (CH$_3$), 57.40 (CH$_3$), 58.07 (CH$_3$), 58.18 (CH$_3$), 58.38 (CH$_3$), 58.40 (CH$_3$), 58.46 (CH$_3$), 58.60 (CH$_3$), 58.84 (CH$_3$), 58.88 (CH$_3$), 58.96 (4 $\times$ CH$_3$), 59.10 (CH$_3$), 59.64 (2 $\times$ CH$_3$), 60.12 (CH$_3$), 60.22 (CH$_2$), 61.02 (CH$_3$), 61.33 (CH$_3$), 61.39 (CH$_3$), 61.46 (CH$_3$), 61.55 (CH$_3$), 61.70 (CH$_3$), 70.18 (CH), 70.42 (CH$_2$), 70.60 (CH), 70.71 (CH$_2$), 70.78 (CH), 70.88 (CH$_2$), 70.92 (CH$_2$), 70.99 (CH$_2$), 71.12 (CH), 71.12 (CH$_2$), 71.21 (2 $\times$ CH), 71.58 (CH), 71.68 (CH$_2$), 72.36 (CH), 72.48 (CH), 75.45 (CH), 76.33 (CH), 78.79 (CH), 78.99 (CH), 79.35 (CH), 80.43 (CH), 80.45 (CH), 81.37 (CH), 81.57 (3 $\times$ CH), 81.74 (2 $\times$ CH), 81.97 (CH), 82.04 (CH), 82.07 (CH), 82.33 (2 $\times$ CH), 82.38 (CH), 82.57 (CH), 82.60 (CH), 82.99 (CH), 96.50 (CH), 97.53 (CH), 97.85 (CH), 98.07 (CH), 98.71 (CH), 99.05 (CH), 100.00 (CH), 101.56 (CH), 105.34 (C), 170.23 (C); MS (ESI$^+$-TOF): m/z (%) 1699 [(M + Na)$^+$, 100]; HRMS (ESI$^+$-TOF): m/z [M + Na]$^+$ calcd for C$_{73}$H$_{128}$NaO$_{42}$ 1699.7778; found 1699.7756. Anal. calcd for C$_{73}$H$_{128}$O$_{42}$: C, 52.26; H, 7.69. Found: C, 52.15; H, 7.53.

2$^{I-VI}$,3$^{I-VI}$,6$^{II,III,V,VI}$-Hexadeca-O-methyl-$\alpha$-cyclomaltohexaose (14), 2$^{I-VI}$,3$^{I-VI}$,6$^{II,IV,VI}$-Hexadeca-O-methyl-$\alpha$-cyclomaltohexaose (23), and 2$^{I-VI}$,3$^{I-VI}$,6$^{III-VI}$-Hexadeca-O-methyl-$\alpha$-cyclomaltohexaose (36). A solution of dry $\alpha$-cyclodextrin (4.69 g, 4.825 mmol) and imidazole (1.148 g, 16.9 mmol) in dry DMF (246 mL) was added TBDMSCl (2.654 g, 9.7 mmol) in one portion at room temperature under nitrogen and the mixture stirred at this temperature for 2 h. After cooling at 0°C, NaH (60%, 10.615 g, 265 mmol) was added in small portions and the mixture
stirred for 30 min and then at room temperature for 1 h. After recooling to 0 °C, MeI (34.2 mL, 549 mmol) was then added dropwise and the stirring continued at room temperature overnight. The excess of NaH was destroyed with MeOH and the mixture poured into ice-water and extracted with CHCl$_3$ (5 × 100 mL). The organic phase was dried over Na$_2$SO$_4$ and concentrated under reduced pressure. The residue in dry MeOH (290 mL) was treated with NH$_4$F (6.28 g, 170 mmol) and heated at reflux temperature under nitrogen for 22.5 h. The solvent was evaporated under reduced pressure and the residue redissolved in CHCl$_3$ (200 mL), filtered over Celite and concentrated to give a crude mixture (12.15 g) that was purified by column chromatography (CHCl$_3$ → CHCl$_3$–MeOH, 96:4) to give the known permethylated α-CD (628 mg, 0.513 mmol, 10%), 2$_{1}$–VI$_{1}$,3$_{1}$–VI$_{1}$,6$_{1}$–VI$_{1}$-heptadeca-O-methyl-α-cyclomaltohexaose (5)$^{25}$ (1602 mg, 1.324 mmol, 27%), and a mixture of the three possible diols that was subjected to a second careful flash chromatography on 60 PF silica gel (0.04–0.063) (CHCl$_3$–MeOH, 98:2→95:5) to give the diols 14$^{25,29}$ (385 mg, 0.322 mmol, 7%), 23 (731 mg, 0.611 mmol, 13%), and 36$^{25}$ (936 mg, 0.783 mmol, 16%). A more polar fraction containing a mixture of triols and other polyols was not studied. Compound 23: colorless oil, [α]$^D$ +141.2 (c 0.97, CHCl$_3$); IR (film): 3472, 2929, 1454, 1365, 1107, 1039 cm$^{-1}$; $^1$H NMR (500 MHz, CDCl$_3$): δ 3.13–3.19 (m, 6H), 3.389 (s, 3H), 3.397 (s, 3H), 3.398 (s, 6H), 3.489 (s, 3H), 3.492 (s, 3H), 3.495 (s, 6H), 3.50 (s, 6H), 3.63 (s, 3H), 3.647 (s, 6H), 3.649 (s, 3H), 3.650 (s, 3H), 3.66 (s, 3H), 5.03 (d, J = 3.8 Hz, 1H), 5.035 (d, J = 3.8 Hz, 1H), 5.05 (d, J = 3.5 Hz, 1H), 5.06 (d, J = 3.5 Hz, 1H), 5.07 (d, J = 3.5 Hz, 1H), 5.08 (d, J = 3.5 Hz, 1H); $^{13}$C NMR (125.7 MHz, CDCl$_3$): δ 57.85 (2 × CH$_3$), 57.90 (2 × CH$_3$), 58.03 (CH$_3$), 58.08 (CH$_3$), 58.93 (CH$_3$), 59.01 (CH$_3$), 59.09 (CH$_3$), 59.15 (CH$_3$), 61.63 (2 × CH$_3$), 61.75 (2 × CH$_3$), 61.77 (CH$_3$), 61.79 (CH$_3$), 62.32 (CH$_2$), 62.40 (CH$_2$), 71.19 (CH), 71.28 (2 × CH), 71.43 (CH$_2$), 71.51 (CH), 71.53 (CH$_2$), 71.55 (CH$_2$), 71.69 (CH$_2$), 72.48 (CH), 72.65 (CH), 81.24 (CH), 81.28 (3 × CH), 81.35 (CH), 81.40 (CH), 81.74 (CH),
81.97 (CH), 82.00 (CH), 82.06 (2 × CH), 82.12 (CH), 82.17 (2 × CH), 82.22 (2 × CH), 82.29 (CH),
82.35 (CH), 99.36 (CH), 99.58 (CH), 99.84 (CH), 99.87 (2 × CH), 99.90 (CH); MS (ESI⁺-TOF):
m/z (%) 1214 [(M + NH₄)⁺, 100]; HRMS (ESI⁺-TOF): m/z [M + NH₄⁺] calc for C₅₂H₉₆NO₃₀
1214.6017; found 1214.6023. Anal. calc for C₅₂H₉₂O₃₀: C, 52.17; H, 7.75. Found: C, 52.05; H,
7.61. Compound 36: colorless oil, [α]D +143.3 (c 1.21, CHCl₃); IR (film): 3468, 2933, 1456, 1365,
1109, 1037 cm⁻¹; ¹H NMR (500 MHz, CDCl₃): δ 3.12–3.19 (m, 6H), 3.386 (s, 6H), 3.390 (s, 6H),
3.477 (s, 3H), 3.485 (s, 6H), 3.492 (s, 3H), 3.497 (s, 3H), 3.501 (s, 3H), 3.632 (s, 3H), 3.637 (s,
3H), 3.640 (s, 3H), 3.642 (s, 3H), 3.644 (s, 3H), 3.65 (s, 3H), 5.01 (d, J = 3.5 Hz, 1H), 5.04 (d, J =
3.2 Hz, 4H), 5.08 (d, J = 3.5 Hz, 1H); ¹³C NMR (125.7 MHz, CDCl₃): δ 57.85 (CH₃), 57.90 (3 ×
CH₃), 57.97 (2 × CH₃), 58.94 (2 × CH₃), 59.00 (CH₃), 59.09 (CH₃), 61.67 (CH₃), 61.72 (3 × CH₃),
61.77 (2 × CH₃), 62.20 (CH₂), 62.54 (CH₂), 71.14 (CH), 71.17 (CH), 71.25 (2 × CH), 71.44 (CH₂),
71.51 (2 × CH₂), 71.60 (CH₂), 72.59 (CH), 72.98 (CH), 81.26 (2 × CH), 81.30 (4 × CH), 81.99 (3
× CH), 82.06 (2 × CH), 82.15 (2 × CH), 82.16 (CH), 82.22 (CH), 82.26 (CH), 82.45 (CH), 82.58
(CH), 99.58 (CH), 99.66 (CH), 99.76 (CH), 99.86 (CH), 99.89 (CH), 99.94 (CH); MS (ESI⁺-TOF):
m/z (%) 1219 [(M + Na)⁺, 100]; HRMS (ESI⁺-TOF): m/z [M + Na⁺] calc for C₅₂H₉₂NaO₃₀
1219.5571; found 1219.5576. Anal. calc for C₅₂H₉₂O₃₀: C, 52.17; H, 7.75. Found: C, 52.30; H,
7.74.

Oxidative HAT of 2⁻,Ⅶ,3⁻,Ⅶ,6⁻,Ⅲ,Ⅵ⁻,Ⅶ-nonadeca-O-methyl-β-cyclomaltoheptaose (11). A
solution of alcohol 11¹⁷a (80 mg, 0.057 mmol) in dry CH₂Cl₂ (2.3 mL) containing DIB (55 mg,
0.171 mmol) and I₂ (24.6 mg, 0.097 mmol) was stirred under nitrogen at 30 °C for 1 h while
irradiated with two 80 W tungsten-filament lamps. The reaction mixture was then poured into 10%
aqueous Na₂S₂O₃, extracted with CH₂Cl₂, dried over Na₂SO₄, and concentrated. The residue was
purified by silica gel [Merck 60 PF (0.063–0.2 mm)] column chromatography (hexanes–acetone,
65:35→50:50) to give cyclo-5\textsuperscript{VII},6\textsuperscript{I}-anhydro-(5\textsuperscript{VII}R)-(2,3,6-tri-O-methyl-\textalpha-D-xylo-hexos-5-ulopyranosyl)-(1→4)-2,3-di-O-methyl-\textalpha-D-glucopyranosyl-(1→4)-2,3,6-tri-O-methyl-\textalpha-D-glucopyranosyl-(1→4)-5\textsuperscript{III},6\textsuperscript{IV}-anhydro-(5\textsuperscript{III}R)-(2,3,6-tri-O-methyl-\textalpha-D-xylo-hexos-5-ulopyranosyl)-(1→4)-2,3-di-O-methyl-\textalpha-D-glucopyranosyl-(1→4)-[(1→4)-2,3,6-tri-O-methyl-\textalpha-D-glucopyranosyl]\textsubscript{2} (12) (16.2 mg, 0.012 mmol, 20%), an inseparable mixture of mono-trioxocane–mono-acetyl positional isomers (11.9 mg, 0.008 mmol, 3:2, 14%) and cyclo-(5\textsuperscript{R})-5\textsuperscript{VII}-O-acetyl-2,3,6-tri-O-methyl-\textalpha-D-xylo-hexos-5-ulopyranosyl-(1→4)-2,3-di-O-methyl-\textalpha-D-glucopyranosyl-(1→4)-[(1→4)-2,3,6-tri-O-methyl-\textalpha-D-glucopyranosyl]\textsubscript{2} (13) (42.3 mg, 0.028 mmol, 49%). Compound 12: colorless oil, [\alpha]_D
+110.9 (c 0.79, CHCl\textsubscript{3}); IR (film): 2933, 1454, 1365, 1139, 1065, 1044 cm\textsuperscript{-1}; \textsuperscript{1}H NMR (500 MHz, CDCl\textsubscript{3}): \delta 3.11 (dd, J = 9.8, 3.2 Hz, 1H), 3.13 (dd, J = 9.5, 3.2 Hz, 1H), 3.179 (dd, J = 9.5, 3.5 Hz, 1H), 3.181 (dd, J = 9.5, 3.5 Hz, 1H), 3.21 (dd, J = 9.8, 3.8 Hz, 1H), 3.28 (dd, J = 9.5, 3.2 Hz, 1H), 3.28 (dd, J = 9.5, 3.2 Hz, 1H), 3.37 (s, 3H), 3.378 (s, 3H), 3.380 (s, 6H), 3.39 (s, 3H), 3.465 (s, 3H), 3.468 (s, 3H), 3.47 (s, 3H), 3.478 (s, 3H), 3.484 (s, 3H), 3.50 (s, 3H), 3.52 (s, 3H), 3.59 (s, 3H), 3.601 (s, 3H), 3.604 (s, 3H), 3.62 (s, 3H), 3.63 (s, 3H), 3.67 (s, 3H), 3.69 (s, 3H), 5.02 (d, J = 3.5 Hz, 1H), 5.04 (d, J = 2.8 Hz, 1H), 5.053 (d, J = 3.5 Hz, 1H), 5.058 (d, J = 3.8 Hz, 1H), 5.066 (d, J = 4.1 Hz, 1H), 5.21 (d, J = 3.8 Hz, 1H), 5.22 (d, J = 3.5 Hz, 1H); \textsuperscript{13}C NMR (125.7 MHz, CDCl\textsubscript{3}): \delta 57.74 (CH\textsubscript{3}), 57.94 (2 × CH\textsubscript{3}), 58.04 (CH\textsubscript{3}), 58.08 (CH\textsubscript{3}), 58.33 (CH\textsubscript{3}), 58.43 (CH\textsubscript{3}), 58.95 (4 × CH\textsubscript{3}), 59.06 (CH\textsubscript{3}), 61.09 (2 × CH\textsubscript{3}), 61.66 (CH\textsubscript{3}), 61.86 (CH\textsubscript{3}), 61.89 (CH\textsubscript{3}), 62.18 (CH\textsubscript{3}), 62.25 (CH\textsubscript{3}), 64.71 (CH\textsubscript{2}, C6\textsuperscript{I} or C6\textsuperscript{IV}), 65.14 (CH\textsubscript{2}, C6\textsuperscript{I} or C6\textsuperscript{IV}), 66.65 (CH, C5\textsuperscript{I} or C5\textsuperscript{IV}), 67.21 (CH, C5\textsuperscript{I} or C5\textsuperscript{IV}), 70.76 (CH\textsubscript{2}), 70.85 (CH), 70.87 (CH), 70.92 (CH), 70.98 (CH\textsubscript{2}), 71.10 (CH\textsubscript{2}), 71.74 (CH\textsubscript{2}), 72.05 (CH\textsubscript{2}), 77.64 (CH, C4\textsuperscript{I} or C4\textsuperscript{IV}), 77.79 (CH, C4\textsuperscript{I} or C4\textsuperscript{IV}), 78.00 (CH),
Some spectroscopic data of the inseparable mixture of mono-trioxocane–mono-acetyl positional isomers could be obtained: colorless oil, $^1$H NMR (500 MHz, CDCl$_3$, complex spectrum, only clearly distinguished signals are reported): $\delta$2.062 (s, 3H), 2.064 (s, 3H), 4.15 (dd, $J = 2.5, 2.5$ Hz, 1H), 4.17 (dd, $J = 2.4, 2.4$ Hz, 1H), 4.96 (d, $J = 3.5$ Hz, 1H), 5.01 (d, $J = 3.5$ Hz, 1H), 5.041 (d, $J = 3.5$ Hz, 1H), 5.05 (d, $J = 4.4$ Hz, 2H), 5.06 (d, $J = 3.2$ Hz, 2H), 5.07 (d, $J = 3.2$ Hz, 1H), 5.10 (d, $J = 3.5$ Hz, 1H), 5.12 (d, $J = 3.5$ Hz, 1H), 5.14 (d, $J = 2.2$ Hz, 1H), 5.17 (d, $J = 2.2$ Hz, 1H), 5.227 (d, $J = 3.4$ Hz, 1H), 5.229 (d, $J = 3.5$ Hz, 1H); $^{13}$C NMR (125.7 MHz, CDCl$_3$, complex spectrum, only clearly distinguished signals are reported): $\delta$22.02 (CH$_3$), 22.06 (CH$_3$), 60.31 (CH$_2$), 60.37 (CH$_2$), 64.84 (CH$_2$), 64.91 (CH$_2$), 66.91 (CH), 67.07 (CH), 96.98 (CH), 97.01 (CH), 97.21 (CH), 97.33 (CH), 97.46 (CH), 97.89 (CH), 99.12 (CH), 99.24 (CH), 99.62 (CH), 99.83 (CH), 100.17 (CH), 100.24 (CH), 100.40 (CH), 101.05 (C), 101.45 (C), 101.93 (CH), 104.51 (C), 104.55 (C), 170.14 (C); MS (ESI$^+$-TOF): m/z (%) 1479 [(M + Na$^+$), 100]; HRMS (ESI$^+$-TOF): m/z [M + Na$^+$] calcd for C$_{63}$H$_{108}$NaO$_{37}$ 1479.6467; found 1479.6475.

Compound 13: colorless oil, $[\alpha]_D +130.8$ (c 0.75, CHCl$_3$); IR (film): 2933, 1732, 1454, 1367, 1193, 1106, 1046 cm$^{-1}$; $^1$H NMR (500 MHz, CDCl$_3$): $\delta$2.04 (s, 6H), 3.12 (dd, $J = 9.3, 2.7$ Hz, 1H), 3.15 (dd, $J = 6.9, 3.2$ Hz, 1H), 3.16 (dd, $J = 7.6, 3.5$ Hz, 1H), 3.22 (dd, $J = 9.8, 3.2$ Hz, 1H), 3.26
(dd, J = 9.8, 3.5 Hz, 1H), 3.367 (s, 6H), 3.370 (s, 3H), 3.38 (s, 3H), 3.39 (s, 3H), 3.40 (s, 6H), 3.458 (s, 3H), 3.461 (s, 3H), 3.463 (s, 3H), 3.49 (s, 3H), 3.50 (s, 3H), 3.52 (s, 3H), 3.588 (s, 3H), 3.591 (s, 3H), 3.595 (s, 3H), 3.598 (s, 3H), 3.617 (s, 3H), 3.619 (s, 3H), 3.90 (d, J = 9.8 Hz, 1H, H6), 3.95 (d, J = 9.8 Hz, 1H, H6), 4.00 (d, J = 9.8 Hz, 1H, H6), 4.03 (d, J = 9.8 Hz, 1H, H6), 4.05 (br d, J = 2.5 Hz, 1H, H4VII), 4.08 (dd, J = 2.5, 2.5 Hz, 1H, H3VIII), 4.16 (dd, J = 2.5, 2.5 Hz, 1H, H3VII), 4.33 (br dd, J = 11.0, 7.3 Hz, 1H), 4.41 (br dd, J = 11.5, 5.8 Hz, 1H), 4.97 (d, J = 2.8 Hz, 1H), 4.98 (d, J = 3.2 Hz, 1H), 5.03 (d, J = 3.5 Hz, 1H), 5.04 (d, J = 3.2 Hz, 1H), 5.09 (d, J = 3.2 Hz, 1H), 5.11 (d, J = 1.9 Hz, 1H, H1III), 5.14 (d, J = 1.9 Hz, 1H, H1VII); 1H NMR (500 MHz, CDCl3, 1D-TOCSY, irradiation at H1VII, 5.14 ppm): δ 3.40 (br s, 1H, H2VII), 4.05 (br s, 1H, H4VII), 4.17 (dd, J = 2.5, 2.5 Hz, 1H, H3VII); 1H NMR (500 MHz, CDCl3, 1D-TOCSY, irradiation at H1III, 5.11 ppm): δ 3.32 (br s, 1H, H2III), 4.02 (br s, 1H, H4III), 4.09 (dd, J = 2.5, 2.5 Hz, 1H, H3III); 1H NMR (500 MHz, C6D6, 18 °C): δ 1.74 (s, 3H), 1.78 (s, 3H), 3.16 (s, 3H), 3.18 (s, 3H), 3.19 (s, 3H), 3.21 (s, 3H), 3.26 (s, 3H), 3.270 (s, 3H), 3.274 (s, 6H), 3.30 (s, 9H), 3.40 (s, 3H), 3.61 (s, 3H), 3.62 (s, 3H), 3.66 (s, 3H), 3.72 (s, 3H), 3.822 (s, 3H), 3.825 (s, 3H), 3.85 (s, 3H), 4.44 (d, J = 9.8 Hz, 1H), 4.51 (d, J = 9.8 Hz, 1H), 5.08 (d, J = 3.2 Hz, 1H), 5.09 (d, J = 3.5 Hz, 1H), 5.15 (d, J = 2.5 Hz, 1H), 5.18 (d, J = 3.8 Hz, 1H), 5.19 (d, J = 2.8 Hz, 1H), 5.59 (d, J = 1.3 Hz, 1H), 5.63 (d, J = 1.3 Hz, 1H); 1H NMR (500 MHz, C6D6, 70 °C): δ 1.79 (s, 3H), 1.83 (s, 3H), 3.21 (s, 3H), 3.23 (s, 3H), 3.25 (s, 3H), 3.27 (s, 3H), 3.29 (s, 3H), 3.308 (s, 3H), 3.312 (s, 3H), 3.313 (s, 3H), 3.319 (s, 3H), 3.322 (s, 3H), 3.34 (s, 3H), 3.39 (s, 3H), 3.60 (s, 3H), 3.61 (s, 3H), 3.699 (s, 3H), 3.702 (s, 3H), 3.79 (s, 3H), 3.80 (s, 3H), 3.83 (s, 3H), 4.10 (dd, J = 11.2, 3.9 Hz, 1H), 4.40 (d, J = 9.8 Hz, 1H), 4.44 (d, J = 9.8 Hz, 1H), 5.05 (d, J = 3.2 Hz, 1H), 5.07 (d, J = 3.5 Hz, 1H), 5.11 (d, J = 2.5 Hz, 1H), 5.13 (d, J = 3.5 Hz, 1H), 5.16 (d, J = 2.8 Hz, 1H), 5.56 (d, J = 1.6 Hz, 1H), 5.65 (d, J = 1.3 Hz, 1H); 13C NMR (125.7 MHz, CDCl3): δ 21.98 (CH3), 22.03 (CH3), 57.16 (CH3), 57.41 (CH3)
57.68 (CH₃), 57.75 (CH₃), 57.86 (CH₃), 58.84 (CH₃), 58.90 (CH₃), 58.99 (4 × CH₃), 59.03 (CH₃), 59.34 (CH₃), 59.68 (CH₃), 59.91 (CH₂), 60.37 (CH₂), 61.02 (CH₃), 61.11 (CH₃), 61.55 (CH₃), 61.72 (CH₃), 61.77 (CH₃), 70.44 (CH), 70.48 (CH₂), 70.60 (CH₂), 70.66 (CH₂), 70.86 (CH₂), 70.87 (CH₂), 71.08 (CH), 71.30 (2 × CH), 71.59 (CH), 71.67 (CH), 72.16 (CH), 76.11 (CH, C₃VII), 76.65 (CH, C₂VII), 76.86 (CH, C³VII), 77.06 (CH, C³VII), 77.99 (CH), 80.39 (CH), 81.55 (CH), 81.60 (2 × CH), 81.67 (CH), 82.02 (CH), 82.06 (CH), 82.15 (CH), 82.17 (CH), 82.20 (CH), 82.34 (CH), 82.68 (CH), 82.84 (CH), 83.13 (CH), 97.70 (CH, C¹VII), 98.39 (CH, C¹VII), 99.67 (CH), 99.70 (CH), 99.82 (CH), 100.24 (CH), 101.71 (CH), 104.92 (C, C⁵VII), 105.15 (C, C⁵VII), 170.08 (C), 170.14 (C); MS (ESI⁺-TOF): m/z (%) 1539 [(M + Na)⁺, 100]; HRMS (ESI⁺-TOF): m/z [M + Na]⁺ caleld for C₆₅H₁₁₂NaO₃₉ 1539.6678; found 1539.6642. Anal. caleld for C₆₅H₁₁₂O₃₉: C, 51.44; H, 7.44. Found: C, 51.49; H, 7.60.

Oxidative HAT of 2¹VI,3¹VI,6¹III,IV-VI-hexadeca-O-methyl-α-cyclomaltohexaose (14). Method A: A solution of alcohol 14²⁵,²⁹ (90 mg, 0.075 mmol) in dry CH₂Cl₂ (3 mL) containing DIB (53 mg, 0.165 mmol) and I₂ (9.5 mg, 0.038 mmol) was stirred under nitrogen at 26 °C for 1.5 h while irradiated with two 80 W tungsten-filament lamps. The reaction mixture was then directly loaded onto a silica gel (TLC Silica gel 60 F₂₅₄, scraped from Merck Aluminum sheets) column chromatography (hexanes–acetone, 65:35→55:45) to give cyclo-5¹VI,6¹-anhydro-(5¹VI⁻R⁻)-(2,3,6-tri-O-methyl-α-D-xylo-hexos-5-ulopyranosyl)-(1→4)-2,3,6-tri-O-methyl-α-D-glucopyranosyl-(1→4)-2,3,6-tri-O-methyl-α-D-glucopyranosyl-(1→4)-5¹III,6¹IV-anhydro-(5¹III⁻R⁻)-(2,3,6-tri-O-methyl-α-D-xylo-hexos-5-ulopyranosyl)-(1→4)-2,3-di-O-methyl-α-D-glucopyranosyl-(1→4)-2,3,6-tri-O-methyl-α-D-glucopyranosyl (15) (25.6 mg, 0.021 mmol, 29%) and cyclo-5¹VI,6¹-anhydro-(5¹VI⁻R⁻)-(2,3,6-tri-O-methyl-α-D-xylo-hexos-5-ulopyranosyl)-(1→4)-2,3-di-O-methyl-α-D-glucopyranosyl-[1(1→4)-2,3,6-tri-O-methyl-α-D-glucopyranosyl]₂-(1→4)-2,3-di-O-methyl-α-D-
glucopyranosyl-(1→4)-2,3,6-tri-O-methyl-α-D-glucopyranosyl (16) (17.5 mg, 0.015 mmol, 19%).

Compound 15: Crystalline solid m.p. 250–252 °C (from n-hexane–EtOAc); [α]D +133.0 (c 1.176, CHCl3); IR (film): 2929, 1454, 1365, 1139, 1109, 1063 cm⁻¹; ¹H NMR (500 MHz, CDCl3): δ 3.14 (dd, J = 9.1, 3.5 Hz, 2H), 3.20 (dd, J = 9.5, 3.5 Hz, 2H), 3.25 (dd, J = 9.8, 3.5 Hz, 2H), 3.368 (s, 6H), 3.369 (s, 6H), 3.47 (s, 6H), 3.48 (s, 6H), 3.50 (s, 6H), 3.53 (dd, J = 9.5, 9.5 Hz, 2H), 3.61 (s, 12H), 3.69 (dd, J = 9.3, 9.3 Hz, 2H), 3.74 (s, 6H), 3.77 (dd, J = 9.5, 9.5 Hz, 2H), 3.93 (d, J = 9.5 Hz, 2H), 3.98 (d, J = 9.8 Hz, 2H), 4.99 (d, J = 3.8 Hz, 2H), 5.02 (d, J = 3.5 Hz, 2H), 5.19 (d, J = 3.2 Hz, 2H); ¹³C NMR (125.7 MHz, CDCl3): δ 57.62 (2 × CH₃), 57.80 (2 × CH₃), 58.13 (2 × CH₃), 58.98 (2 × CH₃), 59.14 (2 × CH₃), 61.28 (2 × CH₃), 62.06 (2 × CH₃), 62.14 (2 × CH₃), 64.01 (2 × CH₂), 66.49 (2 × CH), 70.69 (2 × CH₂), 70.91 (2 × CH), 72.25 (2 × CH₂), 78.32 (2 × CH), 80.38 (2 × CH), 80.67 (2 × CH), 80.83 (2 × CH), 81.24 (2 × CH), 81.42 (2 × CH), 81.73 (2 × CH), 82.07 (2 × CH), 82.21 (2 × CH), 97.28 (2 × CH), 98.63 (2 × CH), 99.13 (2 × CH), 100.37 (2 × C); MS (ESI⁺-TOF): m/z (%) 1215 [(M + Na)⁺, 100]; HRMS (ESI⁺-TOF): m/z [M + Na]⁺ calcld for C₅₂H₈₈NaO₃₁ 1215.5258; found 1215.5253. Anal. calcld for C₅₂H₈₈O₃₁: C, 52.34; H, 7.43. Found: C, 52.41; H, 7.26. Compound 16: colorless oil, [α]D +108.2 (c 1.48, CHCl₃); IR (film): 3475, 2929, 1454, 1365, 1141, 1107, 1046 cm⁻¹; ¹H NMR (500 MHz, CDCl₃): δ 3.12–3.21 (m, 5H), 3.25 (dd, J = 9.8, 3.2 Hz, 1H), 3.36 (s, 3H), 3.37 (s, 3H), 3.379 (s, 3H), 3.380 (s, 3H), 3.47 (s, 3H), 3.47 (s, 6H), 3.48 (s, 3H), 3.50 (s, 3H), 3.53 (s, 3H), 3.60 (s, 3H), 3.61 (s, 3H), 3.618 (s, 3H), 3.620 (s, 3H), 3.71 (s, 3H), 3.73 (s, 3H), 4.96 (d, J = 3.5 Hz, 1H), 4.97 (d, J = 3.5 Hz, 1H), 5.03 (d, J = 3.5 Hz, 1H), 5.04 (d, J = 3.5 Hz, 1H), 5.16 (d, J = 3.8 Hz, 1H), 5.19 (d, J = 3.5 Hz, 1H); ¹³C NMR (125.7 MHz, CDCl₃): δ 57.65 (CH₃), 57.83 (CH₃), 57.85 (CH₃), 57.86 (CH₃), 58.39 (CH₃), 58.72 (CH₃), 59.07 (3 × CH₃), 59.09 (CH₃), 61.25 (2 × CH₃), 61.64 (CH₃), 61.89 (CH₃), 61.95 (CH₂), 61.98 (CH₃), 62.25 (CH₃), 64.05 (CH₂), 66.40 (CH), 70.95 (CH₂), 71.03 (CH), 71.14 (CH), 71.39 (CH₂),
71.81 (CH), 72.03 (CH₂), 72.07 (CH₂), 72.10 (CH), 78.18 (CH), 80.09 (CH), 80.37 (CH), 80.46 (CH), 80.75 (CH), 80.83 (CH), 80.86 (CH), 81.26 (CH), 81.33 (CH), 81.50 (CH), 81.61 (CH), 81.65 (CH), 81.79 (CH), 81.96 (CH), 82.14 (CH), 82.45 (CH), 82.59 (CH), 82.78 (CH), 97.30 (CH), 97.97 (CH), 98.34 (CH), 99.39 (2 × CH), 99.52 (CH), 100.79 (C); MS (ESI⁺-TOF): m/z (%) 1217 [(M + Na)⁺, 100]; HRMS (ESI⁺-TOF): m/z [M + Na]⁺ calcd for C₅₂H₉₀NaO₃₀ 1217.5415; found 1217.5419. Anal. calcd for C₅₂H₉₀O₃₀: C, 52.25; H, 7.59. Found: C, 52.33; H, 7.37.

Method B: A solution of alcohol 14 (45 mg, 0.038 mmol) in dry CH₂Cl₂ (1.5 mL) containing DIB (30 mg, 0.095 mmol) and I₂ (14.5 mg, 0.057 mmol) was stirred under nitrogen at 28 °C for 1.5 h while irradiated with two 80 W tungsten-filament lamps. The reaction mixture was then directly loaded onto a silica gel (TLC Silica gel 60 F₂₅₄, scraped from Merck Aluminum sheets) column chromatography (hexanes–acetone, 65:35→55:45) to give 15 (13 mg, 0.011 mmol, 29%) and 16 (7.2 mg, 0.006 mmol, 16%).

Method C: A solution of alcohol 14 (47 mg, 0.039 mmol) in dry CH₂Cl₂ (1.6 mL) containing DIB (37.7 mg, 0.117 mmol) and I₂ (16.8 mg, 0.066 mmol) was stirred under nitrogen at 28 °C for 1.5 h while irradiated with two 80 W tungsten-filament lamps. The reaction mixture was then directly loaded onto a silica gel (TLC Silica gel 60 F₂₅₄, scraped from Merck Aluminum sheets) column chromatography (hexanes–acetone, 65:35) to give 15 (15.9 mg, 0.013 mmol, 34%).

Oxidative HAT of mono-trioxocane alcohol (16). A solution of alcohol 16 (12 mg, 0.01 mmol) in dry CH₂Cl₂ (0.45 mL) containing DIB (7 mg, 0.022 mmol) and I₂ (1.3 mg, 0.005 mmol) was stirred under nitrogen at 28 °C for 2 h while irradiated with two 80 W tungsten-filament lamps. The reaction mixture was then directly loaded onto a silica gel (TLC Silica gel 60 F₂₅₄, scraped from Merck Aluminum sheets) column chromatography (hexanes–acetone, 65:35) to give bis(trioxocane) 15 (6 mg, 0.005 mmol, 50%).
Oxidative HAT of 2\textsuperscript{I}–7, 3\textsuperscript{I}–7, 6\textsuperscript{II}–7–nonadeca-O-methyl-\(\beta\)-cyclomaltoheptaose (17). A solution of alcohol 17\textsuperscript{17a} (119 mg, 0.085 mmol) in dry CH\textsubscript{2}Cl\textsubscript{2} (3.5 mL) containing DIB (60.2 mg, 0.187 mmol) and I\textsubscript{2} (28 mg, 0.110 mmol) was stirred under nitrogen at 30 °C for 2.5 h while irradiated with two 80 W tungsten-filament lamps. The reaction mixture was then directly loaded onto a silica gel (TLC Silica gel 60 F\textsubscript{254}, scraped from Merck Aluminum sheets) column chromatography (CHCl\textsubscript{3}–MeOH, 99:1→98:2) to give the acetyl derivative 18 as an inseparable mixture of positional isomers (26.1 mg, 0.018 mmol, 21%, 70:30), cyclo-(5\(\text{R}\))-5\textsuperscript{VII}-O-acetyl-2,3,6-tri-O-methyl-\(\alpha\)-D-xylo-hexos-5-ulopyranosyl-(1→4)-2,3-di-O-methyl-\(\alpha\)-D-glucopyranosyl-(1→4)-(5\(\text{R}\))-5\textsuperscript{II}-O-acetyl-2,3,6-tri-O-methyl-\(\alpha\)-D-xylo-hexos-5-ulopyranosyl-(1→4)-2,3-di-O-methyl-\(\alpha\)-D-glucopyranosyl-[1(1→4)-2,3,6-tri-O-methyl-\(\alpha\)-D-glucopyranosyl]\textsubscript{3} (19) (19.1 mg, 0.013 mmol, 15%), cyclo-(5\(\text{R}\))-5\textsuperscript{VII}-O-acetyl-2,3,6-tri-O-methyl-\(\alpha\)-D-xylo-hexos-5-ulopyranosyl-(1→4)-2,3-di-O-methyl-\(\alpha\)-D-glucopyranosyl-(1→4)-5\textsuperscript{II}, 6\textsuperscript{III}-anhydro-(5\textsuperscript{II}R)-(2,3,6-tri-O-methyl-\(\alpha\)-D-xylo-hexos-5-ulopyranosyl)-(1→4)-2,3-di-O-methyl-\(\alpha\)-D-glucopyranosyl-[1(1→4)-2,3,6-tri-O-methyl-\(\alpha\)-D-glucopyranosyl]\textsubscript{3} (20) (24.3 mg, 0.018 mmol, 20%), cyclo-5\textsuperscript{VII}, 6\textsuperscript{I}-anhydro-(5\textsuperscript{VII}R)-(2,3,6-tri-O-methyl-\(\alpha\)-D-xylo-hexos-5-ulopyranosyl)-(1→4)-2,3-di-O-methyl-\(\alpha\)-D-glucopyranosyl-(1→4)-(5\(\text{R}\))-5\textsuperscript{II}-O-acetyl-2,3,6-tri-O-methyl-\(\alpha\)-D-xylo-hexos-5-ulopyranosyl-(1→4)-2,3-di-O-methyl-\(\alpha\)-D-glucopyranosyl-[1(1→4)-2,3,6-tri-O-methyl-\(\alpha\)-D-glucopyranosyl]\textsubscript{3} (21) (7 mg, 0.005 mmol, 6%), cyclo-5\textsuperscript{VII}, 6\textsuperscript{I}-anhydro-(5\textsuperscript{VII}R)-(2,3,6-tri-O-methyl-\(\alpha\)-D-xylo-hexos-5-ulopyranosyl)-(1→4)-2,3-di-O-methyl-\(\alpha\)-D-glucopyranosyl-[1(1→4)-2,3,6-tri-O-methyl-\(\alpha\)-D-glucopyranosyl]\textsubscript{3} (22) (6.9 mg, 0.005 mmol, 6%). Compound 18: colorless oil, IR (film): 3516, 2933, 1732, 1455, 1368, 1105, 1036 cm\textsuperscript{-1}; \(^1\)H NMR (500 MHz, CDCl\textsubscript{3}) (only the major isomer is shown): \(\delta\) 2.06 (s, 3H), 3.18 (m, 6H), 3.30 (dd, \(J = 9.8, 9.8\) Hz, 1H), 3.36 (s, 3H), 3.367 (s, 3H), 3.368 (s, 3H),
3.373 (s, 3H), 3.39 (s, 3H), 3.40 (s, 3H), 3.43 (s, 3H), 3.45 (s, 3H), 3.482 (s, 3H), 3.483 (s, 3H), 3.49 (s, 3H), 3.50 (s, 3H), 3.54 (s, 3H), 3.59 (s, 3H), 3.61 (s, 3H), 3.617 (s, 3H), 3.621 (s, 3H), 3.627 (s, 3H), 3.633 (s, 3H), 3.64 (s, 3H), 3.94 (d, \(J = 9.8\) Hz, 1H), 4.02 (d, \(J = 9.8\) Hz, 1H), 4.05 (d, \(J = 1.9\) Hz, 1H), 4.18 (dd, \(J = 2.4, 2.4\) Hz, 1H), 4.30 (dd, \(J = 11.2, 5.8\) Hz, 1H), 5.01 (d, \(J = 3.2\) Hz, 1H), 5.037 (d, \(J = 3.4\) Hz, 1H), 5.044 (d, \(J = 3.8\) Hz, 1H), 5.079 (d, \(J = 4.1\) Hz, 1H), 5.087 (d, \(J = 4.1\) Hz, 1H), 5.12 (d, \(J = 1.9\) Hz, 1H), 5.16 (d, \(J = 3.8\) Hz, 1H); \(^1\)H NMR (500 MHz, CDCl\(_3\)), 1D-ROESY, irradiation at 2.06 ppm: \(\delta\) 5.12 (d, \(J = 1.9\) Hz, 1H, H\(_{VII}\)); \(^{13}\)C NMR (125.7 MHz, CDCl\(_3\)) (only the major isomer is shown): \(\delta\) 22.10 (CH\(_3\)), 57.21 (CH\(_3\)), 57.50 (CH\(_3\)), 58.24 (CH\(_3\)), 58.53 (CH\(_3\)), 58.57 (CH\(_3\)), 58.88 (2 \(\times\) CH\(_3\)), 58.91 (CH\(_3\)), 58.95 (CH\(_3\)), 58.97 (2 \(\times\) CH\(_3\)), 59.01 (CH\(_3\)), 59.20 (CH\(_3\)), 60.20 (CH\(_2\)), 61.22 (CH\(_3\)), 61.23 (CH\(_3\)), 61.37 (CH\(_3\)), 61.41 (CH\(_3\)), 61.50 (CH\(_3\)), 61.67 (CH\(_3\)), 61.82 (CH\(_2\)), 70.33 (CH\(_2\)), 70.33 (CH), 71.02 (2 \(\times\) CH), 71.02 (CH\(_2\)), 71.13 (2 \(\times\) CH\(_2\)), 71.14 (2 \(\times\) CH), 71.27 (CH\(_2\)), 71.42 (CH), 72.21 (CH), 75.42 (CH), 76.81 (CH), 79.73 (CH), 80.17 (CH), 80.31 (CH), 81.13 (2 \(\times\) CH), 81.52 (CH), 81.62 (CH), 81.66 (2 \(\times\) CH), 81.73 (2 \(\times\) CH), 81.90 (CH), 82.02 (CH), 82.30 (CH), 82.33 (CH), 82.44 (2 \(\times\) CH), 83.67 (CH), 97.69 (CH), 99.10 (CH), 99.35 (CH), 99.37 (CH), 99.41 (CH), 99.62 (CH), 100.10 (CH), 104.99 (C), 170.18 (C); MS (ESI\(^+\)-TOF): \(m/z\) (%) \(1481\) [(M + Na\(^+\)), 100]; HRMS (ESI\(^+\)-TOF): \(m/z\) [M + Na\(^+\)] calcld for C\(_{63}\)H\(_{110}\)O\(_37\) \(1481.6624\); found \(1481.6647\). Anal. calcld for C\(_{63}\)H\(_{110}\)O\(_37\): C, 51.84; H, 7.60. Found: C, 51.60; H, 7.45. Compound 19: colorless oil, \([\alpha]_D +118.9\) (c 1.34, CHCl\(_3\)); IR (film): 3521, 2928, 1732, 1455, 1368, 1195, 1105, 1041 cm\(^{-1}\); \(^1\)H NMR (500 MHz, CDCl\(_3\)): \(\delta\) 2.04 (s, 3H), 2.05 (s, 3H), 3.16 (dd, \(J = 9.8, 3.8\) Hz, 2H), 3.17 (dd, \(J = 9.5, 3.2\) Hz, 1H), 3.21 (dd, \(J = 9.8, 3.5\) Hz, 1H), 3.25 (dd, \(J = 9.9, 3.3\) Hz, 1H), 3.30 (dd, \(J = 9.8, 9.8\) Hz, 1H), 3.32 (dd, \(J = 2.2, 2.2\) Hz, 1H, H2), 3.36 (s, 3H), 3.37 (s, 3H), 3.38 (s, 3H), 3.39 (s, 3H), 3.40 (s, 3H), 3.41 (s, 3H), 3.42 (s, 3H), 3.46 (s, 3H), 3.47 (s, 3H), 3.479 (s, 3H), 3.482 (s, 3H), 3.53 (s, 3H), 3.54 (s, 3H), 3.586 (s, 3H),
3.593 (s, 3H), 3.60 (s, 3H), 3.61 (s, 3H), 3.62 (s, 3H), 3.64 (s, 3H), 4.01 (br d, J = 3.2 Hz, 1H, H4), 4.04 (dd, J = 2.5, 2.5 Hz, 1H, H3), 4.06 (d, J = 2.2 Hz, 1H, H4), 4.18 (dd, J = 2.5, 2.5 Hz, 1H, H3), 4.99 (d, J = 3.2 Hz, 1H), 5.00 (d, J = 4.1 Hz, 1H), 5.07 (d, J = 2.2 Hz, 1H, H1), 5.079 (d, J = 3.5 Hz, 1H), 5.082 (d, J = 3.5 Hz, 1H), 5.10 (d, J = 3.5 Hz, 1H), 5.12 (d, J = 1.9 Hz, 1H, H1); 1H NMR (500 MHz, C₆D₆): δ 1.68 (s, 3H), 1.84 (s, 3H), 3.01 (dd, J = 9.1, 2.8 Hz, 1H), 3.10 (dd, J = 9.5, 3.5 Hz, 1H), 3.21 (s, 3H), 3.22 (s, 3H), 3.22 (s, 3H), 3.24 (s, 3H), 3.40 (s, 3H), 3.41 (s, 6H), 3.42 (s, 3H), 3.48 (s, 3H), 3.65 (s, 3H), 3.66 (s, 6H), 3.69 (s, 3H), 5.00 (d, J = 2.8 Hz, 1H), 5.14 (d, J = 3.2 Hz, 1H), 5.18 (d, J = 3.2 Hz, 1H), 5.32 (d, J = 3.5 Hz, 1H), 5.48 (d, J = 2.5 Hz, 1H), 5.53 (d, J = 2.2 Hz, 1H); 1H NMR (500 MHz, CDCl₃, 1D-TOCSY, irradiation at H1ⅡorⅦ, 5.12 ppm): δ 3.42 (br s, 1H, H2), 4.05 (br s, 1H, H4), 4.22 (br s, 1H, H3); 1H NMR (500 MHz, CDCl₃, 1D-TOCSY, irradiation at H1ⅠorⅢ, 5.07 ppm): δ 3.36 (br s, 1H, H2), 4.08 (dd, J = 2.5, 2.5 Hz, 1H, H3), 4.09 (br s, 1H, H4); 1H NMR (500 MHz, CDCl₃, 2D-ROESY): both acetates show NOE interactions with their respective anomeric hydrogens: H1Ⅶand H1Ⅲ; 13C NMR (125.7 MHz, CDCl₃): δ 22.05 (CH₃), 22.08 (CH₃), 57.33 (CH₃), 57.37 (CH₃), 57.58 (CH₃), 58.20 (CH₃), 58.38 (CH₃), 58.71 (CH₃), 58.93 (CH₃), 59.64 (5 × CH₃), 59.36 (CH₃), 59.94 (CH₂), 60.06 (CH₃), 60.36 (CH₂), 61.14 (CH₃), 61.27 (CH₃), 61.45 (2 × CH₃), 61.65 (CH₃), 70.08 (CH), 70.34 (CH₂), 70.57 (CH₂), 70.66 (CH₂), 70.71 (CH₂), 70.88 (CH₂), 70.98 (CH), 71.14 (CH), 71.30 (CH), 71.51 (CH), 71.86 (CH), 72.43 (CH), 75.96 (CH), 77.02 (CH), 77.14 (CH), 77.56 (CH), 78.17 (CH), 80.10 (CH), 81.36 (CH), 81.39 (CH), 81.49 (CH), 81.58 (CH), 81.63 (CH), 81.88 (CH), 81.99 (CH), 82.01 (CH), 82.08 (CH), 82.51 (CH), 82.74 (CH), 83.03 (CH), 83.12 (CH), 97.80 (CH), 97.82 (CH), 99.46 (2 × CH), 99.59 (CH), 100.49 (CH), 101.81 (CH), 104.33 (C), 105.18 (C), 170.01 (C), 170.12 (C); MS (ESI⁻-TOF): m/z (%) 1539 [(M + Na)⁺, 100]; HRMS (ESI⁺-TOF): m/z [M + Na]⁺
calcd for C_{65}H_{112}NaO_{39} 1539.6678; found 1539.6660. Anal. calcd for C_{65}H_{112}O_{39}: C, 51.44; H, 7.44. Compound 20: colorless oil, [α]_D +123.5 (c 0.96, CHCl₃); IR (film): 3540, 2928, 1732, 1455, 1368, 1138, 1105, 1046 cm⁻¹; ¹H NMR (500 MHz, CDCl₃): δ 2.05 (s, 3H), 3.24 (dd, J = 10.1, 3.5 Hz, 1H), 3.25 (dd, J = 10.1, 3.3 Hz, 1H), 3.35 (s, 3H), 3.37 (s, 3H), 3.38 (s, 3H), 3.39 (s, 3H), 3.40 (s, 3H), 3.42 (s, 3H), 3.45 (s, 3H), 3.477 (s, 3H), 3.480 (s, 3H), 3.49 (s, 3H), 3.50 (s, 3H), 3.53 (s, 3H), 3.60 (s, 3H), 3.61 (s, 6H), 3.63 (s, 3H), 3.66 (s, 3H), 3.67 (s, 3H), 3.68 (s, 3H), 3.90 (d, J = 8.8 Hz, 1H), 4.21 (m, 1H), 4.23 (dd, J = 2.5, 2.5 Hz, 1H), 4.27 (br dd, J = 11.7, 6.3 Hz, 1H), 4.98 (d, J = 3.2 Hz, 1H), 5.00 (d, J = 3.2 Hz, 1H), 5.04 (d, J = 3.5 Hz, 1H), 5.09 (d, J = 4.1 Hz, 1H), 5.13 (d, J = 3.5 Hz, 1H), 5.15 (d, J = 1.9 Hz, 1H), 5.17 (d, J = 3.5 Hz, 1H); ¹H NMR (500 MHz, CDCl₃, 1D-TOCSY, irradiation at H₁^{II}, 5.17 ppm): δ 3.24 (dd, J = 9.9, 3.7 Hz, 1H, H₂^{II}), 3.64 (dd, J = 9.2, 9.2 Hz, 1H, H₃^{II}), 3.90 (d, J = 9.0 Hz, 1H, H₄^{II}); ¹H NMR (500 MHz, CDCl₃, 1D-TOCSY, irradiation at H₁^{I}, 5.00 ppm): δ 3.25 (dd, J = 9.3, 3.0 Hz, 1H, H₂^{I}), 3.54 (m, 1H, H₆^{I}), 3.59 (dd, J = 9.4, 9.4 Hz, 1H, H₃^{I}), 3.63 (dd, J = 9.1, 9.1 Hz, 1H, H₄^{I}), 4.21 (br d, J = 10 Hz, 1H, H₅^{I}), 4.27 (br dd, J = 12.6, 5.4 Hz, 1H, H₆^{I}); ¹H NMR (500 MHz, CDCl₃, 1D-ROESY, irradiation at H₁^{I}, 5.00 ppm): δ 3.25 (dd, J = 9.3, 3.3 Hz, 1H, H₂^{I}), 3.90 (d, J = 9.1 Hz, 1H, H₄^{I}); ¹³C NMR (125.7 MHz, CDCl₃): δ 22.05 (CH₃), 57.42 (CH₃), 57.56 (2 × CH₃), 58.30 (CH₃), 58.39 (CH₃), 58.88 (CH₃), 58.90 (CH₃), 58.93 (CH₃), 58.98 (CH₃), 59.01 (CH₃), 59.04 (3 × CH₃), 60.13 (CH₂), 61.13 (2 × CH₃), 61.40 (CH₃), 61.61 (CH₃), 61.69 (CH₃), 62.06 (CH₃), 63.85 (CH₂), 67.06 (CH), 70.48 (CH₂), 70.74 (CH), 70.77 (CH₂), 70.90 (CH), 70.90 (CH₂), 71.03 (CH₂), 71.08 (CH), 71.43 (CH), 72.16 (CH₂), 72.83 (CH), 75.62 (CH), 76.57 (CH), 78.46 (CH), 79.02 (CH), 80.24 (CH), 80.32 (CH), 80.35 (CH), 80.63 (CH), 81.03 (CH), 81.48 (CH), 81.57 (CH), 81.69 (2 × CH), 81.73 (CH), 81.77 (2 × CH), 82.21 (CH), 82.71 (CH), 83.09 (CH), 83.38 (CH), 97.20 (CH), 97.31 (CH), 98.31 (CH), 98.45 (CH), 99.61 (CH), 100.31 (CH), 100.35 (C), 102.03
CH), 105.25 (C), 170.26 (C); MS (ESI$^+$-TOF): \(m/z\) (%) 1479 [(M + Na)$^+$, 100]; HRMS (ESI$^+$-TOF): \(m/z\) [M + Na]$^+$ calec for C$_{63}$H$_{108}$NaO$_{37}$ 1479.6467; found 1479.6483. Anal. calec for C$_{63}$H$_{108}$O$_{37}$: C, 51.92; H, 7.47. Found: C, 52.16; H, 7.75. Compound 21: colorless oil, $\alpha$$^\circ$D +110.6 (c 0.36, CHCl$_3$); IR (film): 3526, 2928, 1732, 1455, 1368, 1143, 1107, 1046 cm$^{-1}$; $^1$H NMR (500 MHz, CDCl$_3$): $\delta$ 2.06 (s, 3H), 3.12 (dd, $J$ = 10.1, 3.5 Hz, 1H), 3.15 (dd, $J$ = 9.1, 3.2 Hz, 1H), 3.20 (dd, $J$ = 9.8, 3.8 Hz, 1H), 3.35 (s, 3H), 3.37 (s, 9H), 3.40 (s, 3H), 3.43 (s, 3H), 3.46 (s, 3H), 3.47 (s, 6H), 3.48 (s, 3H), 3.51 (s, 3H), 3.56 (s, 3H), 3.61 (s, 9H), 3.64 (s, 3H), 3.64 (s, 3H), 3.66 (s, 3H), 3.67 (s, 3H), 3.82 (dd, $J$ = 10.7, 10.7 Hz, 1H, H$_6^{III}$), 3.95 (d, $J$ = 9.8 Hz, 1H, H$_6^{II}$), 4.06 (d, $J$ = 9.8 Hz, 1H, H$_6^{III}$), 4.11 (br s, 2H, H$_4^{II}$ and H$_3^{II}$), 4.37 (ddd, $J$ = 12.0, 6.6, 1.6 Hz, 1H, H$_6^{III}$), 5.01 (d, $J$ = 3.2 Hz, 1H), 5.03 (d, $J$ = 3.5 Hz, 1H), 5.04 (d, $J$ = 4.4 Hz, 1H), 5.08 (d, $J$ = 1.9 Hz, 1H), 5.163 (d, $J$ = 3.2 Hz, 1H), 5.166 (d, $J$ = 2.9 Hz, 1H), 5.21 (d, $J$ = 3.8 Hz, 1H); $^1$H NMR (500 MHz, CDCl$_3$, 1D-TOCSY, irradiation at H$_1^{II}$, 5.08 ppm): $\delta$ 3.24 (br s, 1H, H$_2^{II}$), 4.109 (dd, $J$ = 2.9, 2.9 Hz, 1H, H$_3^{II}$), 4.111 (d, $J$ = 2.9 Hz, 1H, H$_4^{II}$); $^1$H NMR (500 MHz, CDCl$_3$, 1D-TOCSY, irradiation at H$_1^{I}$, 5.01 ppm): $\delta$ 3.16 (br d, $J$ = 10.1 Hz, 1H, H$_2^{I}$), 3.42 (dd, $J$ = 9.3, 9.3 Hz, 1H, H$_3^{I}$), 3.46 (dd, $J$ = 9.0, 9.0 Hz, 1H, H$_4^{I}$), 3.65 (m, 1H, H$_5^{I}$), 3.98 (dd, $J$ = 9.2, 2.8 Hz, 1H, H$_6^{I}$); $^1$H NMR (500 MHz, CDCl$_3$, 1D-ROESY, irradiation at H$_1^{I}$, 5.01 ppm): $\delta$ 4.11 (d, $J$ = 2.9 Hz, 1H, H$_4^{II}$); $^{13}$C NMR (125.7 MHz, CDCl$_3$): $\delta$ 22.13 (CH$_3$), 57.08 (CH$_3$), 57.54 (CH$_3$), 57.84 (CH$_3$), 58.14 (CH$_3$), 58.20 (CH$_3$), 58.84 (CH$_3$), 58.86 (CH$_3$), 58.89 (CH$_3$), 58.98 (CH$_3$), 59.04 (CH$_3$), 59.10 (CH$_3$), 59.14 (CH$_3$), 59.89 (CH$_3$), 60.37 (CH$_2$), 60.90 (CH$_3$), 61.25 (CH$_3$), 61.35 (CH$_3$), 61.67 (CH$_3$), 61.73 (CH$_3$), 62.10 (CH$_3$), 63.91 (CH$_2$), 67.13 (CH), 69.00 (CH), 70.29 (CH$_2$), 70.45 (CH), 70.95 (CH), 70.95 (CH$_2$), 71.33 (CH), 71.47 (CH$_2$), 71.54 (CH), 71.54 (CH$_2$), 71.82 (CH$_2$), 75.65 (CH), 76.98 (CH), 77.20 (CH), 77.55 (CH), 79.22 (CH), 79.90 (CH), 80.36 (CH), 80.86 (CH), 80.96 (CH), 80.99 (CH), 81.07 (CH), 81.36 (CH), 81.79 (CH), 81.87 (CH), 81.95 (CH), 81.97 (CH), 82.34
Compound 22: colorless oil, $[\alpha]_D^{+} +116.9$ (c 0.46, CHCl$_3$); IR (film): 2928, 1455, 1370, 1159, 1102, 1046 cm$^{-1}$; $^1$H NMR (500 MHz, CDCl$_3$): $\delta$ 3.099 (d, $J = 9.8$ Hz, 1H), 3.10 (dd, $J = 9.5$, 3.5 Hz, 1H), 3.24 (dd, $J = 9.8$, 9.8 Hz, 1H), 3.30 (dd, $J = 9.5$, 3.5 Hz, 1H), 3.34 (s, 3H), 3.37 (s, 3H), 3.38 (s, 6H), 3.39 (s, 3H), 3.47 (s, 6H), 3.48 (s, 3H), 3.49 (s, 3H), 3.54 (s, 3H), 3.57 (s, 3H), 3.575 (s, 3H), 3.582 (s, 3H), 3.59 (s, 6H), 3.618 (s, 3H), 3.619 (s, 3H), 3.70 (s, 3H), 3.74 (s, 3H), 4.24 (dd, $J = 9.1$, 7.9 Hz, 1H), 4.30 (dd, $J = 10.7$, 7.9, 7.9 Hz, 1H), 5.02 (d, $J = 3.5$ Hz, 1H), 5.06 (d, $J = 3.5$ Hz, 1H), 5.09 (d, $J = 3.2$ Hz, 1H), 5.094 (d, $J = 3.2$ Hz, 1H), 5.094 (d, $J = 3.2$ Hz, 1H), 5.23 (d, $J = 3.5$ Hz, 1H), 5.30 (d, $J = 3.8$ Hz, 1H); $^1$H NMR (500 MHz, CDCl$_3$, 1D-TOCSY, irradiation at H1$^{\text{III}}$, 5.30 ppm): $\delta$ 3.20 (dd, $J = 9.6$, 3.8 Hz, 1H, H2$^{\text{III}}$), 3.59 (dd, $J = 9.2$, 9.2 Hz, 1H, H3$^{\text{III}}$), 3.64 (dd, $J = 9.4$, 9.4 Hz, 1H, H6$^{\text{III}}$), 3.68 (dd, $J = 9.2$, 9.2 Hz, 1H, H4$^{\text{III}}$), 4.24 (dd, $J = 9.0$, 9.0 Hz, 1H, H6$^{\text{III}}$), 3.99 (ddd, $J = 10.4$, 8.0, 8.0 Hz, 1H, H5$^{\text{III}}$); $^1$H NMR (500 MHz, CDCl$_3$, 1D-TOCSY, irradiation at H1$^{1}$, 5.23 ppm): $\delta$ 3.31 (dd, $J = 9.3$, 3.9 Hz, 1H, H2$^{1}$), 3.78 (d, $J = 9.6$, 9.6 Hz, 1H, H3$^{1}$), 3.85 (d, $J = 9.5$ Hz, 1H, H4$^{1}$); $^1$H NMR (500 MHz, CDCl$_3$, 1D-ROESY, irradiation at H1$^{\text{III}}$, 5.30 ppm): $\delta$ 3.21 (dd, $J = 9.7$, 3.8 Hz, 1H, H2$^{\text{III}}$), 3.60 (dd, $J = 9.2$, 9.2 Hz, 1H, H4$^{\text{IV}}$); $^{13}$C NMR (125.7 MHz, CDCl$_3$): $\delta$ 57.84 (CH$_3$), 57.88 (CH$_3$), 58.02 (CH$_3$), 58.26 (CH$_3$), 58.34 (CH$_3$), 58.81 (CH$_3$), 58.83 (CH$_3$), 58.94 (CH$_3$), 58.98 (CH$_3$), 59.38 (CH$_3$), 59.89 (CH$_3$), 60.75 (2 $\times$ CH$_3$), 61.01 (CH$_3$), 61.10 (CH$_3$), 61.50 (CH$_3$), 61.85 (CH$_3$), 61.96 (CH$_3$), 62.28 (CH$_3$), 62.53 (CH), 62.79 (CH$_2$), 65.12 (CH$_2$), 66.50 (CH), 70.27 (CH), 70.96 (CH$_2$), 71.08 (CH), 71.11 (CH), 71.16 (CH$_2$), 71.42 (CH$_2$), 71.73 (CH$_2$), 71.94 (CH), 72.18 (CH$_2$), 74.72 (CH), 77.17 (CH), 77.61 (CH), 78.77 (CH), 79.92 (CH), 80.57 (CH), 80.83 (CH), 82.87 (CH), 83.09 (CH), 83.45 (CH), 97.08 (CH), 97.66 (CH), 98.06 (CH), 98.21 (CH), 98.77 (CH), 99.10 (CH), 99.28 (CH), 100.76 (C), 105.18 (C), 170.26 (C); MS (ESI$^+$-TOF): m/z (%) 1479 [(M + Na)$^+$, 100]; HRMS (ESI$^+$-TOF): m/z [M + Na]$^+$ calcd for C$_{63}$H$_{108}$NaO$_{37}$ 1479.6467; found 1479.6459. Anal. calcd for C$_{63}$H$_{108}$O$_{37}$: C, 51.92; H, 7.47. Found: C, 52.14; H, 7.39.
81.03 (CH), 81.14 (CH), 81.31 (CH), 81.46 (CH), 81.47 (CH), 81.48 (CH)

1419 [(M + Na)]^+; HRMS (ESI^-TOF): m/z [M + Na]^+ calcd for C_{61}H_{104}NaO_{35} 1419.6256; found 1419.6287. Anal. calcd for C_{61}H_{104}O_{35}: C, 52.43; H, 7.50. Found: C, 52.18; H, 7.74.

**Oxidative HAT of 2^{I-VI}, 3^{I-VI}, 6^{II-IV-VI}-hexadeca-O-methyl-\alpha-cyclomaltohexaose (23).** A solution of alcohol 23 (120 mg, 0.10 mmol) in dry CHCl_3 (4 mL) containing DIB (71 mg, 0.22 mmol) and I_2 (25.4 mg, 0.10 mmol) was stirred under nitrogen at 28 °C for 1.5 h while irradiated with two 80 W tungsten-filament lamps. The reaction mixture was then directly loaded onto a silica gel (TLC Silica gel 60 F_{254}, scraped from Merck Aluminum sheets) column chromatography (hexanes–acetone, 65:35→60:40) to give cyclo-5^{II},6^{III}-anhydro-(5^{II}R)-(2,3,6-tri-O-methyl-\alpha-D-xylo-hexos-5-ulopyranosyl)-(1→4)-2,3-di-O-methyl-\alpha-D-glucopyranosyl-[(1→4)-2,3,6-tri-O-methyl-\alpha-D-glucopyranosyl]_2-(1→4)-5^{VI},6^{I}-anhydro-(5^{VI}R)-(2,3,6-tri-O-methyl-\alpha-D-xylo-hexos-5-ulopyranosyl)-(1→4)-2,3-di-O-methyl-\alpha-D-glucopyranosyl (24) (33.4 mg, 0.028, 28%), cyclo-5^{II},6^{III}-anhydro-(5^{R})-(2,3,6-tri-O-methyl-\alpha-D-xylo-hexos-5-ulopyranosyl)-(1→4)-2,3-di-O-methyl-\alpha-D-glucopyranosyl-[(1→4)-2,3,6-tri-O-methyl-\alpha-D-glucopyranosyl]_2-(1→4)-2,3,6-tri-O-methyl-\alpha-D-glucopyranosyl (25) (22 mg, 0.018 mmol, 18%), and cyclo-5^{VI},6^{I}-anhydro-(5^{VI}R)-(2,3,6-tri-O-methyl-\alpha-D-xylo-hexos-5-ulopyranosyl)-(1→4)-2,3-di-O-methyl-\alpha-D-glucopyranosyl (26) (24 mg, 0.020 mmol, 20%). Compound 24: colorless oil, [\alpha]_D +124.1 (c 1.70, CHCl_3); IR (film): 2928, 1453, 1368, 1140, 1107, 1060 cm^{-1}; ^1H NMR (500 MHz, CDCl_3): \delta 3.14 (m, 4H), 3.22 (dd, J = 9.1, 3.8 Hz, 1H), 3.24
(dd, J = 9.5, 3.5 Hz, 1H), 3.35 (s, 3H), 3.36 (s, 6H), 3.37 (s, 3H), 3.466 (s, 3H), 3.469 (s, 3H), 3.485 (s, 3H), 3.491 (s, 6H), 3.494 (s, 3H), 3.60 (s, 3H), 3.62 (s, 3H), 3.63 (s, 3H), 3.68 (s, 3H), 3.70 (s, 3H), 3.71 (s, 3H), 4.98 (d, J = 3.8 Hz, 1H), 5.00 (d, J = 3.2 Hz, 2H), 5.03 (d, J = 3.8 Hz, 1H), 5.15 (d, J = 3.5 Hz, 1H), 5.16 (d, J = 3.5 Hz, 1H); \(^{13}\)C NMR (125.7 MHz, CDCl\(_3\)): \(\delta\) 57.59 (CH\(_3\)), 57.70 (CH\(_3\)), 57.92 (CH\(_3\)), 58.11 (CH\(_3\)), 58.38 (CH\(_3\)), 58.62 (CH\(_3\)), 58.76 (CH\(_3\)), 59.09 (CH\(_3\)), 59.14 (CH\(_3\)), 61.27 (CH\(_3\)), 61.32 (CH\(_3\)), 61.52 (CH\(_3\)), 61.86 (CH\(_3\)), 61.94 (CH\(_3\)), 62.14 (CH\(_3\)), 63.44 (CH\(_2\)), 63.91 (CH\(_2\)), 65.60 (CH), 66.37 (CH), 70.48 (CH), 71.03 (CH), 71.57 (CH\(_2\)), 71.67 (CH\(_2\)), 71.81 (CH\(_2\)), 72.90 (CH\(_2\)), 78.05 (CH), 78.23 (CH), 79.77 (CH), 79.91 (CH), 80.07 (CH), 80.74 (CH), 80.82 (CH), 80.97 (CH), 81.14 (CH), 81.16 (CH), 81.19 (CH), 81.23 (CH), 81.34 (CH), 81.79 (CH), 82.37 (CH), 82.44 (CH), 82.60 (CH), 83.29 (CH), 96.36 (CH), 96.48 (CH), 97.37 (CH), 97.53 (CH), 100.15 (CH), 100.21 (CH), 100.47 (C), 100.78 (C); MS (ESI\(^{+}\)-TOF): \(m/z\) (%) 1215 [(M + Na\(^{+}\), 100]; HRMS (ESI\(^{+}\)-TOF): \(m/z\) [M + Na\(^{+}\)] calcd for C\(_{52}\)H\(_{88}\)NaO\(_3\): 1215.5258; found 1215.5237. Anal. calcd for C\(_{52}\)H\(_{88}\)O\(_3\): C, 52.34; H, 7.43. Found: C, 52.42; H, 7.43. Compound 25: colorless oil, \([\alpha]_D^{+}+123.7\) (c 1.81, CHCl\(_3\)); IR (film): 3494, 2933, 1454, 1365, 1109, 1039 cm\(^{-1}\); \(^1\)H NMR (500 MHz, CDCl\(_3\)): \(\delta\) 3.11 (dd, J = 10.1, 3.5 Hz, 1H), 3.13 (dd, J = 10.4, 3.5 Hz, 1H), 3.15 (dd, J = 9.8, 3.2 Hz, 1H), 3.18 (dd, J = 10.1, 3.5 Hz, 1H), 3.20 (dd, J = 9.8, 3.5 Hz, 1H), 3.25 (dd, J = 9.6, 3.3 Hz, 1H), 3.35 (s, 3H), 3.369 (s, 3H), 3.371 (s, 3H), 3.38 (s, 3H), 3.476 (s, 3H), 3.479 (s, 3H), 3.487 (s, 3H), 3.490 (s, 6H), 3.495 (s, 3H), 3.58 (s, 3H), 3.60 (s, 3H), 3.638 (s, 3H), 3.644 (s, 3H), 3.70 (s, 3H), 3.71 (s, 3H), 4.98 (d, J = 3.8 Hz, 1H), 4.99 (d, J = 3.8 Hz, 1H), 5.00 (d, J = 3.5 Hz, 1H), 5.01 (d, J = 3.5 Hz, 1H), 5.08 (d, J = 3.5 Hz, 1H), 5.17 (d, J = 3.2 Hz, 1H); \(^{13}\)C NMR (125.7 MHz, CDCl\(_3\)): \(\delta\) 57.71 (CH\(_3\)), 57.84 (2 × CH\(_3\)), 58.17 (CH\(_3\)), 58.42 (CH\(_3\)), 58.49 (CH\(_3\)), 58.95 (2 × CH\(_3\)), 59.00 (CH\(_3\)), 59.14 (CH\(_3\)), 61.28 (CH\(_3\)), 61.33 (CH\(_3\)), 61.61 (CH\(_3\)), 61.90 (CH\(_3\)), 61.94 (CH\(_3\)), 62.06 (CH\(_2\)), 62.20 (CH\(_3\)), 63.76 (CH\(_2\)), 66.25 (CH), 70.96 (CH),
71.05 (CH₂), 71.40 (CH), 71.52 (CH₂), 71.62 (CH), 72.12 (CH₂), 72.22 (CH₂), 78.45 (CH), 80.44 (2 × CH), 80.81 (CH), 80.94 (CH), 81.01 (CH), 81.06 (CH), 81.19 (CH), 81.35 (CH), 81.44 (CH), 81.76 (CH), 81.93 (CH), 82.09 (CH), 82.12 (CH), 82.33 (2 × CH), 82.55 (CH), 82.59 (CH), 97.44 (CH), 97.80 (CH), 98.64 (CH), 99.10 (CH), 99.78 (CH), 100.04 (CH), 100.69 (C); MS (ESI⁻-TOF): m/z (%) 1217 [(M + Na)^⁺, 100]; HRMS (ESI⁺-TOF): m/z [M + Na]^⁺ calcd for C₅₂H₉₀NaO₃₀ 1217.5415; found 1217.5405. Anal. calcd for C₅₂H₉₀O₃₀: C, 52.25; H, 7.59. Found: C, 51.98; H, 7.48. Compound 26: colorless oil, [α]D +112.0 (c 1.65, CHCl₃); IR (film): 3475, 2933, 1454, 1367, 1111, 1044 cm⁻¹; ¹H NMR (500 MHz, CDCl₃): δ 3.10 (dd, J = 10.1, 3.5 Hz, 1H), 3.13 (dd, J = 10.1, 3.5 Hz, 1H), 3.15 (dd, J = 9.2, 3.8 Hz, 1H), 3.18 (dd, J = 9.8, 3.8 Hz, 1H), 3.19 (dd, J = 9.8, 3.2 Hz, 1H), 3.25 (dd, J = 9.6, 3.3 Hz, 1H), 3.35 (s, 3H), 3.37 (s, 3H), 3.37 (s, 3H), 3.38 (s, 3H), 3.47 (s, 3H), 3.48 (s, 3H), 3.48 (s, 3H), 3.49 (s, 3H), 3.53 (s, 3H), 3.59 (s, 3H), 3.60 (s, 3H), 3.63 (s, 3H), 3.63 (s, 3H), 3.70 (s, 3H), 3.74 (s, 3H), 4.96 (d, J = 3.8 Hz, 1H), 5.01 (d, J = 3.8 Hz, 1H), 5.03 (d, J = 3.5 Hz, 1H), 5.04 (d, J = 3.2 Hz, 1H), 5.07 (d, J = 3.5 Hz, 1H), 5.19 (d, J = 3.5 Hz, 1H); ¹³C NMR (125.7 MHz, CDCl₃): δ 57.67 (CH₃), 57.82 (CH₃), 58.47 (CH₃), 58.80 (CH₃), 59.00 (2 × CH₃), 59.09 (CH₃), 61.28 (CH₃), 61.50 (CH₃), 61.54 (CH₃), 61.93 (CH₃), 61.97 (CH₃), 62.25 (CH₃), 62.48 (CH₂), 63.86 (CH₂), 66.28 (CH), 70.87 (CH), 71.03 (CH), 71.10 (CH), 71.12 (CH₂), 71.24 (CH₂), 71.83 (CH₂), 71.99 (CH₂), 72.77 (CH), 78.30 (CH), 80.46 (CH), 80.57 (CH), 80.73 (CH), 80.78 (CH), 81.02 (2 × CH), 81.31 (CH), 81.42 (CH), 81.50 (CH), 81.75 (CH), 81.89 (CH), 81.97 (CH), 82.11 (CH), 82.30 (CH), 82.50 (CH), 82.60 (CH), 82.70 (CH), 97.36 (CH), 97.75 (CH), 98.77 (CH), 99.57 (CH), 99.86 (CH), 100.04 (CH), 100.80 (C); MS (ESI⁺-TOF): m/z (%) 1217 [(M + Na)^⁺, 100]; HRMS (ESI⁺-TOF): m/z [M + Na]^⁺ calcd for C₅₂H₉₀NaO₃₀ 1217.5415; found 1217.5427. Anal. calcd for C₅₂H₉₀O₃₀: C, 52.25; H, 7.59. Found: C, 52.30; H, 7.51.
Cyclo-5Ⅱ,6Ⅲ-anhydro-(5ⅡR)-(2,3,6-tri-O-methyl-α-D-xylo-hexos-5-ulopyranosyl)-(1→4)-2,3-di-O-methyl-α-D-glucopyranosyl-[((1→4)-2,3,6-tri-O-methyl-α-D-glucopyranosyl)₃-(1→4)-6′-O-acetyl-2,3-di-O-methyl-α-D-glucopyranosyl (27). A solution of alcohol 25 (16.7 mg, 0.014 mmol) in dry pyridine (0.9 mL) containing Ac₂O (0.3 mL) and DMAP (0.2 mg, 0.001 mmol) was stirred at room temperature for 16 h. The mixture was then poured into 10% aqueous HCl and extracted with CHCl₃. The organic layer was washed with aqueous saturated NaHCO₃, dried over Na₂SO₄ and concentrated under reduced pressure. The residue was then purified by silica gel (TLC Silica gel 60 F₂₅₄, scraped from Merck Aluminum sheets) column chromatography (CHCl₃–MeOH, 99.5:0.5) to give acetyl-trioxocane 27 (14.5 mg, 0.012 mmol, 84%) as a colorless oil: [α]D +113.2 (c 1.45, CHCl₃); IR (film): 2929, 1742, 1456, 1365, 1109, 1044 cm⁻¹; ¹H NMR (500 MHz, CDCl₃): δ 2.07 (s, 3H), 3.19 (dd, J = 9.8, 3.2 Hz, 2H), 3.25 (dd, J = 9.8, 3.2 Hz, 1H), 3.35 (s, 3H), 3.37 (s, 6H), 3.383 (s, 3H), 3.477 (s, 3H), 3.482 (s, 3H), 3.489 (s, 3H), 3.490 (s, 3H), 3.495 (s, 3H), 3.50 (s, 3H), 3.59 (s, 3H), 3.60 (s, 3H), 3.63 (s, 6H), 3.63 (s, 3H), 3.67 (s, 3H), 3.70 (s, 3H), 3.72 (s, 3H), 4.08 (dd, J = 11.2, 3.0 Hz, 1H), 4.18 (dd, J = 12.0, 4.1 Hz, 1H), 4.38 (dd, J = 12.0, 1.6 Hz, 1H), 5.02 (m, 3H), 5.03 (d, J = 3.2 Hz, 1H), 5.06 (d, J = 3.2 Hz, 1H), 5.19 (d, J = 3.5 Hz, 1H); ¹H NMR (500 MHz, C₆D₆): δ 1.87 (s, 3H), 3.26 (s, 6H), 3.28 (s, 3H), 3.28 (s, 3H), 3.29 (s, 3H), 3.29 (s, 3H), 3.32 (s, 6H), 3.37 (s, 3H), 3.39 (s, 3H), 3.48 (s, 3H), 3.65 (s, 3H), 3.68 (s, 3H), 3.70 (s, 3H), 3.80 (s, 3H), 3.84 (s, 3H), 3.85 (s, 3H), 4.06 (dd, J = 10.7, 10.7 Hz, 1H, H₆(Ⅲ)), 4.61 (m, 2H, H₅(Ⅰ) and H₆(Ⅰ)), 5.02 (br d, J = 10.7 Hz, 1H, H₆(Ⅰ)), 5.08 (d, J = 3.8 Hz, 1H), 5.13 (d, J = 3.8 Hz, 1H), 5.17 (d, J = 3.5 Hz, 1H, H₁(Ⅱ)), 5.18 (d, J = 4.1 Hz, 1H), 5.20 (d, J = 3.5 Hz, 1H), 5.22 (d, J = 3.5 Hz, 1H, H₁(Ⅰ)); ¹H NMR (500 MHz, C₆D₆, 1D-TOCSY, irradiation at H₆(Ⅰ), 5.02 ppm): δ 3.21 (dd, J = 9.6, 3.4 Hz, 1H, H₂(Ⅰ)), 3.58 (dd, J = 9.0, 9.0 Hz, 1H, H₄(Ⅰ)), 3.83 (dd, J = 9.1, 9.1 Hz, 1H, H₃(Ⅰ)), 4.61 (m, 2H, H₅(Ⅰ) and H₆(Ⅰ)), 5.22 (d, J = 2.9 Hz, 1H, H₁(Ⅰ)); ¹H NMR (500 MHz, C₆D₆, 1D-TOCSY, irradiation at H₁(Ⅱ), 5.17 ppm): δ 3.25 (dd, J
= 9.6, 2.7 Hz, 1H, H2\textsuperscript{ii}), 4.15 (dd, \(J = 9.3, 9.3\ Hz, 1H, H3\textsuperscript{ii}), 4.26 (d, \(J = 9.4\ Hz, 1H, H4\textsuperscript{ii}); \textsuperscript{1}H NMR (500 MHz, CDCl\textsubscript{3}, 1D-ROESY, irradiation at H1\textsuperscript{i}, 5.22 ppm): \(\delta\) 4.26 (d, \(J = 9.2\ Hz, 1H, H4\textsuperscript{ii}); 13C NMR (125.7 MHz, CDCl\textsubscript{3}): \(\delta\) 20.98 (CH\textsubscript{3}), 57.77 (CH\textsubscript{3}), 57.83 (2 \times CH\textsubscript{3}), 58.12 (CH\textsubscript{3}), 58.20 (CH\textsubscript{3}), 58.39 (CH\textsubscript{3}), 58.92 (CH\textsubscript{3}), 58.99 (CH\textsubscript{3}), 59.09 (CH\textsubscript{3}), 59.17 (CH\textsubscript{3}), 61.30 (CH\textsubscript{3}), 61.56 (CH\textsubscript{3}), 61.89 (CH\textsubscript{3}), 62.02 (CH\textsubscript{3}), 62.30 (CH\textsubscript{3}), 63.91 (CH\textsubscript{2}), 64.07 (CH\textsubscript{2}), 66.10 (CH), 69.20 (CH), 70.75 (CH\textsubscript{2}), 70.86 (CH), 71.21 (CH\textsubscript{2}), 71.33 (CH\textsubscript{2}), 71.38 (CH), 71.48 (CH), 72.02 (CH\textsubscript{2}), 78.26 (CH), 80.03 (CH), 80.60 (CH), 80.61 (CH), 80.83 (CH), 81.04 (CH), 81.06 (CH), 81.15 (CH), 81.35 (CH), 81.42 (CH), 81.79 (2 \times CH), 82.09 (CH), 82.11 (CH), 82.16 (CH), 82.35 (CH), 82.50 (CH), 82.60 (CH), 97.39 (CH), 97.46 (CH), 99.09 (CH), 99.35 (CH), 100.05 (CH), 100.71 (CH), 100.75 (C), 170.86 (C); 13C NMR (125.7 MHz, C\textsubscript{6}D\textsubscript{6}): \(\delta\) 20.78 (CH\textsubscript{3}), 57.24 (CH\textsubscript{3}), 57.67 (CH\textsubscript{3}), 57.78 (2 \times CH\textsubscript{3}), 58.15 (CH\textsubscript{3}), 58.25 (CH\textsubscript{3}), 58.84 (CH\textsubscript{3}), 58.97 (CH\textsubscript{3}), 59.01 (CH\textsubscript{3}), 59.10 (CH\textsubscript{3}), 61.29 (CH\textsubscript{3}), 61.56 (CH\textsubscript{3}), 61.65 (CH\textsubscript{3}), 61.79 (CH\textsubscript{3}), 62.11 (CH\textsubscript{3}), 62.14 (CH\textsubscript{3}), 64.31 (CH\textsubscript{2}), 64.68 (CH\textsubscript{2}), 66.70 (CH), 69.93 (CH), 71.74 (CH\textsubscript{2}), 71.79 (CH), 72.23 (CH), 72.25 (CH\textsubscript{2}), 72.50 (CH), 72.72 (CH\textsubscript{2}), 73.00 (CH\textsubscript{2}), 79.05 (CH), 81.53 (CH), 81.57 (CH), 81.60 (CH), 81.63 (CH), 81.82 (CH), 81.84 (CH), 82.04 (CH), 82.13 (2 \times CH), 82.68 (CH), 82.72 (CH), 82.96 (CH), 83.00 (CH), 83.06 (CH), 83.17 (CH), 83.31 (CH), 83.36 (CH), 98.06 (CH), 98.52 (CH), 99.08 (CH), 99.18 (CH), 99.93 (CH), 100.88 (CH), 101.20 (C), 170.11 (C); MS (ESI\textsuperscript{+}-TOF): \textit{m/z} (\%) 1259 [\(\text{[M + Na]}\textsuperscript{+}\), 100]; HRMS (ESI\textsuperscript{+}-TOF): \textit{m/z} [\text{M + Na}]\textsuperscript{+} calcd for C\textsubscript{54}H\textsubscript{92}NaO\textsubscript{31} 1259.5520; found 1259.5526. Anal. calcd for C\textsubscript{54}H\textsubscript{92}O\textsubscript{31}: C, 52.42; H, 7.49. Found: C, 52.35; H, 7.56.

\textit{Cyclo-5\textsuperscript{VI},6\textsuperscript{I}-anhydro-(5\textsuperscript{VI}R)-(2,3,6-tri-O-methyl-\textgreek{a}-D-xylo-hexos-5-ulopyranosyl)-(1\rlap{\textsuperscript{I}}\textsuperscript{IV})-2,3-di-O-methyl-\textgreek{a}-D-glucopyranosyl-(1\rlap{\textsuperscript{I}}\textsuperscript{IV})-2,3,6-tri-O-methyl-\textgreek{a}-D-glucopyranosyl-(1\rlap{\textsuperscript{I}}\textsuperscript{IV})-6\textsuperscript{III}-O-acetyl-2,3-di-O-methyl-\textgreek{a}-D-glucopyranosyl-[(1\rlap{\textsuperscript{I}}\textsuperscript{IV})-2,3,6-tri-O-methyl-\textgreek{a}-D-glucopyranosyl]\textsubscript{2} (28). A solution of alcohol 26 (24 mg, 0.02 mmol) in dry pyridine (0.9 mL) containing Ac\textsubscript{2}O (0.3
mL) and DMAP (0.3 mg, 0.002 mmol) was stirred at room temperature for 14 h. The mixture was then poured into 10% aqueous HCl and extracted with CHCl₃. The organic layer was washed with aqueous saturated NaHCO₃, dried over Na₂SO₄ and concentrated under reduced pressure. The residue was then purified by silica gel (TLC Silica gel 60 F₂₅₄, scraped from Merck Aluminum sheets) column chromatography (CHCl₃–MeOH, 99.5:0.5) to give acetyl-trioxocane 28 (10 mg, 0.008 mmol, 40%), as a colorless oil: [α]D +130.8 (c 0.98, CHCl₃); IR (film): 2929, 1744, 1454, 1367, 1109, 1046 cm⁻¹; ¹H NMR (500 MHz, CDCl₃): δ 2.07 (s, 3H), 3.179 (dd, J = 10.1, 3.5 Hz, 1H), 3.198 (dd, J = 9.8, 3.5 Hz, 1H), 3.198 (dd, J = 9.8, 3.5 Hz, 1H), 3.257 (dd, J = 9.5, 3.2 Hz, 1H), 3.34 (s, 3H), 3.37 (s, 3H), 3.381 (s, 3H), 3.384 (s, 3H), 3.47 (s, 3H), 3.476 (s, 3H), 3.489 (s, 3H), 3.494 (s, 3H), 3.50 (s, 3H), 3.52 (s, 3H), 3.59 (s, 3H), 3.60 (s, 3H), 3.638 (s, 3H), 3.642 (s, 3H), 3.70 (s, 3H), 3.73 (s, 3H), 3.93 (d, J = 9.5 Hz, 1H, H⁴VII), 4.39 (dd, J = 12.1, 4.3 Hz, 1H, H⁶III), 4.56 (dd, J = 12.3, 1.9 Hz, 1H, H⁶III), 4.98 (d, J = 3.8 Hz, 2H), 5.04 (d, J = 3.5 Hz, 1H), 5.05 (d, J = 3.5 Hz, 2H), 5.19 (d, J = 3.2 Hz, 1H, H¹I); ¹H NMR (500 MHz, CDCl₃, 1D-TOCSY, irradiation at H⁶III, 4.56 ppm): δ 3.14 (dd, J = 10.0, 3.3 Hz, 1H, H²III), 3.57 (dd, J = 9.1, 9.1 Hz, 1H, H⁴III), 3.69 (dd, J = 9.4, 9.4 Hz, 1H, H³III), 3.77 (m, 1H, H⁵III), 4.40 (dd, J = 11.7, 3.9 Hz, 1H, H⁶III), 5.05 (d, J = 2.5 Hz, 1H, H¹III); ¹H NMR (500 MHz, CDCl₃, 1D-TOCSY, irradiation at H¹VI, 5.19 ppm): δ 3.26 (dd, J = 9.7, 3.2 Hz, 1H, H²VI), 3.72 (dd, J = 9.3, 9.3 Hz, 1H, H³VI), 3.94 (d, J = 9.4 Hz, 1H, H⁴VI); ¹H NMR (500 MHz, CDCl₃, 1D-ROESY, irradiation at H⁴VI, 3.93 ppm): δ 5.04 (d, J = 3.2 Hz, 1H, H¹V); ¹³C NMR (125.7 MHz, CDCl₃): δ 20.90 (CH₃), 57.69 (CH₃), 57.82 (CH₃), 58.11 (2 × CH₃), 58.28 (CH₃), 58.46 (CH₃), 58.83 (CH₃), 58.93 (CH₃), 59.05 (CH₃), 59.10 (CH₃), 61.29 (CH₃), 61.48 (CH₃), 61.59 (CH₃), 61.85 (CH₃), 62.08 (CH₃), 62.27 (CH₃), 63.69 (CH₂), 63.85 (CH₂), 66.21 (CH), 69.89 (CH), 70.77 (CH₂), 70.86 (CH), 70.98 (CH), 71.26 (CH), 71.35 (CH₂), 71.56 (CH₂), 71.96 (CH₂), 78.29 (CH), 79.87 (CH), 80.61 (CH), 80.76 (2 × CH), 81.04 (CH), 81.09 (CH), 81.17 (CH),
81.39 (CH), 81.51 (CH), 81.75 (CH), 82.04 (CH), 82.17 (CH), 82.38 (CH), 82.49 (CH), 82.71 (2 × CH), 97.32 (CH), 97.37 (CH), 99.50 (CH), 99.61 (CH), 99.78 (CH), 100.17 (CH), 100.83 (C), 170.55 (C); MS (ESI⁺-TOF): m/z (%) 1259 [(M + Na)⁺, 100]; HRMS (ESI⁺-TOF): m/z [M + Na]⁺ calcd for C₅₄H₉₂NaO₃₁ 1259.5520; found 1259.5504. Anal. calcd for C₅₄H₉₂O₃₁: C, 52.42; H, 7.49. Found: C, 52.20; H, 7.65.

Oxidative HAT of 2ᴵ-VᴵII,3ᴵ-VᴵII,6ᴵᴵ-VĪII-nonadeca-O-methyl-β-cyclomaltoheptaose (29). A solution of alcohol 29ᴵᴵᵃ (160 mg, 0.114 mmol) in dry CH₂Cl₂ (4.6 mL) containing DIB (110 mg, 0.34 mmol) and I₂ (49 mg, 0.19 mmol) was stirred under nitrogen at 28 °C for 2.5 h while irradiated with two 80 W tungsten-filament lamps. The reaction mixture was then directly loaded onto a silica gel (TLC Silica gel 60 F₂₅₄, scraped from Merck Aluminum sheets) column chromatography (hexanes–acetone, 65:35→55:45) to give cyclo-5ᴵᴵ,6ᴵ-anhydro-(5ᴵᴵIR)-(2,3,6-tri-O-methyl-α-D-xylo-hexos-5-ulopyranosyl)-(1→4)-(1¹R)-4¹I,6¹II-O-(2,3-di-O-methyl-D-glucopyranosylidene)-2,3-di-O-methyl-α-D-glucopyranosyl-[1-(1→4)-2,3,6-tri-O-methyl-α-D-glucopyranosyl]₄ (30) (8 mg, 0.0057 mmol, 5%), ten-membered lactone 3¹ (17.1 mg, 0.012 mmol, 11%), and a mixture (54.1 mg) of two alcohols 3₂ and 3₃ that was separated after acetylation as mentioned below. To a solution of this mixture in anhydrous pyridine (3 mL) were added Ac₂O (1 mL) and DMAP (1 mg). After 14 h at room temperature, the solution was poured into ice-water and extracted with CH₂Cl₂. The organic phase was successively washed with aqueous solutions of HCl (10%) and saturated NaHCO₃, dried over Na₂SO₄, and concentrated under reduced pressure. The residue (86.5 mg) was purified by silica gel (TLC Silica gel 60 F₂₅₄, scraped from Merck Aluminum sheets) column chromatography (CHCl₃–MeOH, 99:1) to give cyclo-5ᴵᴵ,6ᴵ-anhydro-(5ᴵᴵIR)-(2,3,6-tri-O-methyl-α-D-xylo-hexos-5-ulopyranosyl)-(1→4)-2,3-di-O-methyl-α-D-glucopyranosyl-(1→4)-6¹II-O-acetyl-2,3-di-O-methyl-α-D-glucopyranosyl-[1-(1→4)-2,3,6-tri-O-methyl-α-D-glucopyranosyl]₄ (3₄) (36
mg, 0.025 mmol, 22%) and the lactone 35 (16 mg, 0.013 mmol, 11%). Compound 30: colorless oil, [α]D +108.0 (c 0.45, CHCl₃); IR (film): 2925, 1454, 1369, 1143, 1091, 1042 cm⁻¹; ¹H NMR (500 MHz, CDCl₃): δ 3.06 (d, J = 9.5 Hz, 1H), 3.17 (dd, J = 9.5, 3.7 Hz, 1H), 3.19 (dd, J = 9.5, 3.2 Hz, 1H), 3.209 (dd, J = 9.2, 4.1 Hz, 1H), 3.211 (dd, J = 9.5, 3.5 Hz, 1H), 3.22 (dd, J = 9.5, 3.8 Hz, 1H), 3.27 (dd, J = 9.8, 3.8 Hz, 1H), 3.33 (s, 3H), 3.35 (s, 3H), 3.37 (s, 3H), 3.39 (s, 3H), 3.39 (s, 3H), 3.47 (s, 3H), 3.47 (s, 3H), 3.50 (dd, J = 9.5, 3.2 Hz, 1H), 3.52 (s, 3H), 3.55 (s, 3H), 3.56 (s, 3H), 3.59 (s, 9H), 3.64 (s, 3H), 3.64 (s, 3H), 3.66 (s, 3H), 3.70 (s, 3H), 4.22 (dd, J = 10.4, 8.2 Hz, 1H), 5.04 (d, J = 3.5 Hz, 1H), 5.07 (d, J = 3.2 Hz, 2H), 5.20 (d, J = 3.8 Hz, 1H), 5.23 (d, J = 4.4 Hz, 1H), 5.34 (d, J = 4.1 Hz, 1H); ¹H NMR (500 MHz, CDCl₃, 1D-TOCSY, irradiation at H2Ⅰ, 3.06 ppm): δ 3.40 (dd, J = 9.7, 9.7 Hz, 1H, H3Ⅰ), 3.54 (dd, J = 9.3, 9.3 Hz, 1H, H4Ⅰ), 3.75 (m, 2H, H6Ⅰ and H5Ⅰ), 3.90 (dd, J = 9.9, 9.9 Hz, 1H, H6Ⅰ); ¹H NMR (500 MHz, CDCl₃, 1D-TOCSY, irradiation at H6Ⅱ, 4.22 ppm): δ 3.21 (dd, J = 9.6, 4.3 Hz, 1H, H2Ⅱ), 3.57 (dd, J = 9.2, 9.2 Hz, 1H, H3Ⅱ), 3.63 (dd, J = 10.3, 6.8 Hz, 1H, H6Ⅱ), 3.72 (dd, J = 10.0, 10.0 Hz, 1H, H4Ⅱ), 4.00 (m, 1H, H5Ⅱ), 5.23 (d, J = 4.0 Hz, 1H, H1Ⅱ); ¹³C NMR (125.7 MHz, CDCl₃): δ 57.80 (CH₃), 57.88 (CH₃), 58.07 (CH₃), 58.22 (CH₃), 58.91 (CH₃), 59.02 (2 × CH₃), 59.04 (CH₃), 59.14 (CH₃), 59.44 (CH₃), 60.04 (CH₃), 60.57 (CH₃), 60.70 (CH₃), 60.96 (CH₃), 61.07 (CH₃), 61.18 (CH₃), 61.67 (CH₃), 61.70 (CH₃), 61.84 (CH₂), 62.04 (CH₃), 63.66 (CH), 64.36 (CH₂), 67.62 (CH), 70.05 (CH), 70.07 (CH), 71.06 (CH), 71.09 (CH₂), 71.41 (CH), 71.49 (CH₂), 71.83 (CH₂), 71.84 (CH₂), 71.93 (CH₂), 74.10 (CH), 77.50 (CH), 77.87 (CH), 78.37 (CH), 79.20 (CH), 80.18 (CH), 80.35 (CH), 81.02 (CH), 81.05 (CH), 81.11 (CH), 81.37 (CH), 81.70 (CH), 81.80 (CH), 81.88 (CH), 81.92 (CH), 82.28 (CH), 82.34 (CH), 82.45 (CH), 82.48 (CH), 82.76 (CH), 84.92 (CH), 97.04 (CH), 97.99 (CH), 98.61 (CH), 98.83 (CH), 99.81 (CH), 100.31 (CH), 101.04 (C), 111.58 (C); MS (ESI⁺-TOF): m/z (%) 1419 [(M + Na)⁺, 100]; HRMS (ESI⁺-TOF): m/z [M + Na]⁺ calcd for C₆₁H₁₀₄NaO₃₅ 1419.6256; found
1419.6281. Anal. calcd for C_{61}H_{104}O_{35}: C, 52.43; H, 7.50. Found: C, 52.35; H, 7.73. Compound 31: colorless oil, [α]_D +121.8 (c 0.68, CHCl₃); IR (film): 2929, 1755, 1745, 1454, 1367, 1161, 1106, 1039 cm⁻¹; ^1H NMR (500 MHz, CDCl₃): δ 2.14 (s, 3H), 3.04 (dd, J = 9.8, 3.5 Hz, 1H), 3.16 (dd, J = 9.8, 3.5 Hz, 1H), 3.16 (dd, J = 9.8, 3.5 Hz, 1H), 3.18 (dd, J = 9.8, 3.2 Hz, 1H), 3.19 (dd, J = 9.8, 3.5 Hz, 1H), 3.24 (dd, J = 9.5, 3.8 Hz, 1H), 3.35 (s, 3H), 3.361 (s, 3H), 3.365 (s, 3H), 3.37 (s, 3H), 3.38 (s, 3H), 3.46 (s, 6H), 3.485 (s, 3H), 3.49 (s, 3H), 3.51 (s, 3H), 3.53 (s, 3H), 3.58 (s, 3H), 3.59 (s, 3H), 3.61 (s, 3H), 3.62 (s, 3H), 3.63 (s, 3H), 3.66 (s, 3H), 3.69 (s, 3H), 3.73 (s, 3H), 4.36 (ddd, J = 9.9, 9.9, 5.8 Hz, 1H), 4.58 (dd, J = 10.7, 5.7 Hz, 1H), 4.96 (d, J = 3.8 Hz, 1H), 5.00 (d, J = 3.5 Hz, 1H), 5.09 (d, J = 3.8 Hz, 2H), 5.17 (d, J = 3.5 Hz, 1H), 5.26 (d, J = 3.8 Hz, 1H), 6.30 (d, J = 1.9 Hz, 1H); ^1H NMR (500 MHz, CDCl₃, 1D-TOCSY, irradiation at H₅ᴵ, 4.36 ppm): δ 3.05 (dd, J = 9.9, 3.4 Hz, 1H, H₂ᴵ), 3.45 (dd, J = 8.8, 8.8 Hz, 1H, H₃ᴵ), 3.55 (dd, J = 9.3, 9.3 Hz, 1H, H₄ᴵ), 3.81 (dd, J = 10.4, 10.4 Hz, 1H, H₆ᴵ), 4.59 (dd, J = 10.8, 5.7 Hz, 1H, H₆ᴵ), 5.00 (d, J = 2.9 Hz, 1H, H₁ᴵ); ^1H NMR (500 MHz, CDCl₃, 1D-TOCSY, irradiation at H₁ᴵ, 6.30 ppm): δ 3.34 (d, J = 9.0 Hz, 1H, H₂ᴵ), 3.60 (m, 1H, H₃ᴵ), 3.90 (d, J = 9.4 Hz, 1H, H₄ᴵ); ^1H NMR (500 MHz, CDCl₃, 1D-ROESY, irradiation at H₁ᴵ, 6.30 ppm): δ 3.55 (dd, J = 9.3, 9.3 Hz, 1H, H₄ᴵ), 3.90 (d, J = 9.4 Hz, 1H, H₄ᴵ); ^13C NMR (125.7 MHz, CDCl₃): δ 21.43 (CH₃), 57.86 (CH₃), 58.29 (2 × CH₃), 58.39 (CH₃), 58.40 (CH₃), 58.83 (CH₃), 58.94 (CH₃), 58.99 (CH₃), 59.01 (CH₃), 59.04 (2 × CH₃), 60.97 (CH₃), 61.25 (CH₃), 61.50 (CH₃), 61.69 (2 × CH₃), 61.73 (CH₃), 62.20 (2 × CH₃), 63.34 (CH), 65.84 (CH₂), 70.17 (CH), 70.69 (CH), 70.82 (CH₂), 70.93 (2 × CH), 71.09 (CH), 71.12 (CH₂), 71.14 (CH₂), 71.35 (CH₂), 71.66 (CH₂), 78.64 (CH), 79.61 (CH), 80.14 (CH), 80.52 (CH), 80.64 (CH), 80.89 (CH), 81.32 (2 × CH), 81.42 (CH), 81.58 (CH), 81.64 (CH), 81.68 (CH), 81.81 (2 × CH), 81.84 (CH), 82.01 (CH), 82.05 (CH), 82.13 (CH), 82.27 (CH), 83.42 (CH), 84.59 (CH), 94.21 (CH), 97.86 (CH), 98.70 (CH), 98.83 (CH), 99.11 (CH), 99.38
(CH), 99.98 (CH), 170.81 (C), 170.98 (C); MS (ESI<sup>+</sup>-TOF): m/z (%) 1449 [(M + Na)<sup>+</sup>, 100]; HRMS (ESI<sup>+</sup>-TOF): m/z [M + Na]<sup>+</sup> calc for C<sub>62</sub>H<sub>106</sub>NaO<sub>36</sub> 1449.6362; found 1449.6360. Anal. calc for C<sub>62</sub>H<sub>106</sub>O<sub>36</sub>: C, 52.17; H, 7.48. Found: C, 51.91; H, 7.48. Compound 34: colorless oil, [α]<sub>D</sub> +105.2 (c 0.94, CHCl<sub>3</sub>); IR (film): 2933, 1745, 1454, 1369, 1161, 1109, 1041 cm<sup>−1</sup>; <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ 2.08 (s, 3H), 3.13 (dd, <i>J</i> = 9.5, 3.5 Hz, 1H), 3.28 (dd, <i>J</i> = 9.6, 3.6 Hz, 1H), 3.36 (s, 3H), 3.37 (s, 3H), 3.38 (s, 6H), 3.40 (s, 3H), 3.46 (s, 6H), 3.50 (s, 9H), 3.51 (s, 6H), 3.58 (s, 3H), 3.61 (s, 3H), 3.62 (s, 3H), 3.64 (s, 3H), 3.65 (s, 3H), 3.69 (s, 3H), 4.29 (dd, <i>J</i> = 12.3, 3.8 Hz, 1H), 4.49 (dd, <i>J</i> = 12.0, 1.9 Hz, 1H), 4.92 (d, <i>J</i> = 3.5 Hz, 1H), 5.07 (d, <i>J</i> = 3.5 Hz, 1H), 5.074 (d, <i>J</i> = 3.8 Hz, 1H), 5.09 (d, <i>J</i> = 3.5 Hz, 1H), 5.13 (d, <i>J</i> = 3.5 Hz, 1H), 5.20 (d, <i>J</i> = 3.8 Hz, 1H), 5.23 (d, <i>J</i> = 3.8 Hz, 1H); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>, 1D-TOCSY, irradiation at H1<sup>i</sup>, 4.92 ppm): δ 3.14 (dd, <i>J</i> = 9.0, 2.8 Hz, 1H, H2<sup>i</sup>), 3.53 (dd, <i>J</i> = 8.7, 8.7 Hz, 1H, H4<sup>i</sup>), 3.59 (dd, <i>J</i> = 9.5, 9.5 Hz, 1H, H3<sup>i</sup>); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>, 1D-TOCSY, irradiation at H6<sup>ii</sup>, 4.29 ppm): δ 3.19 (dd, <i>J</i> = 9.7, 3.9 Hz, 1H, H2<sup>ii</sup>), 3.40 (dd, <i>J</i> = 9.2, 9.2 Hz, 1H, H3<sup>ii</sup>), 3.57 (dd, <i>J</i> = 9.7, 9.7 Hz, 1H, H4<sup>ii</sup>), 4.49 (br d, <i>J</i> = 11.6 Hz, 1H, H6<sup>ii</sup>), 5.08 (d, <i>J</i> = 3.3 Hz, 1H, H1<sup>ii</sup>); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>, 1D-ROESY, irradiation at H1<sup>i</sup>, 4.92 ppm): δ 3.14 (dd, <i>J</i> = 9.4, 3.2 Hz, 1H, H2<sup>i</sup>), 3.56 (dd, <i>J</i> = 9.4, 9.4 Hz, 1H, H4<sup>ii</sup>); <sup>13</sup>C NMR (125.7 MHz, CDCl<sub>3</sub>): δ 20.84 (CH<sub>3</sub>), 57.86 (CH<sub>3</sub>), 57.89 (CH<sub>3</sub>), 58.09 (CH<sub>3</sub>), 58.42 (CH<sub>3</sub>), 58.45 (CH<sub>3</sub>), 58.72 (CH<sub>3</sub>), 58.88 (CH<sub>3</sub>), 58.97 (CH<sub>3</sub>), 59.00 (2 × CH<sub>3</sub>), 59.07 (CH<sub>3</sub>), 59.11 (CH<sub>3</sub>), 60.81 (CH<sub>3</sub>), 61.02 (CH<sub>3</sub>), 61.35 (CH<sub>3</sub>), 61.55 (CH<sub>3</sub>), 61.66 (CH<sub>3</sub>), 61.85 (CH<sub>3</sub>), 62.16 (CH<sub>3</sub>), 63.36 (CH<sub>2</sub>), 64.58 (CH<sub>2</sub>), 67.04 (CH), 69.46 (CH), 70.67 (CH), 70.89 (CH), 70.96 (CH), 70.96 (CH<sub>2</sub>), 70.99 (CH), 71.03 (CH<sub>2</sub>), 71.36 (CH<sub>2</sub>), 71.74 (CH<sub>2</sub>), 71.83 (CH<sub>2</sub>), 77.62 (CH), 78.85 (CH), 79.05 (CH), 80.19 (CH), 80.29 (CH), 80.51 (CH), 80.82 (CH), 81.13 (CH), 81.21 (CH), 81.31 (CH), 81.42 (CH), 81.46 (CH), 81.51 (CH), 81.79 (2 × CH), 81.80 (CH), 81.83 (CH), 81.98 (CH), 82.27 (CH), 82.35 (CH), 82.67 (CH), 97.02 (CH), 97.81 (CH), 98.95 (CH),
Compound 35: colorless oil, [α]D +120.3 (c 0.64, CHCl3); IR (film): 2929, 1745, 1454, 1369, 1159, 1107, 1041 cm⁻¹; ¹H NMR (500 MHz, CDCl3): δ 2.10 (s, 3H), 3.18 (dd, J = 9.8, 3.8 Hz, 4H), 3.25 (dd, J = 9.8, 3.8 Hz, 1H), 3.35 (s, 3H), 3.37 (s, 3H), 3.38 (s, 3H), 3.385 (s, 3H), 3.389 (s, 3H), 3.45 (dd, J = 9.8, 2.8 Hz, 1H), 3.49 (s, 3H), 3.49 (s, 9H), 3.51 (s, 3H), 3.54 (s, 3H), 3.60 (s, 3H), 3.62 (s, 9H), 3.63 (s, 3H), 3.64 (s, 3H), 3.67 (s, 6H), 3.95 (dd, J = 9.5, 9.5 Hz, 1H), 3.99 (ddd, J = 9.8, 3.9, 1.7 Hz, 1H), 4.17 (d, J = 8.5 Hz, 1H), 4.21 (ddd, J = 10.1, 1.9, 1.9 Hz, 1H), 4.29 (dd, J = 12.5, 3.9 Hz, 1H), 4.45 (dd, J = 12.3, 1.9 Hz, 1H), 5.11 (d, J = 3.5 Hz, 1H), 5.12 (d, J = 3.8 Hz, 1H), 5.13 (d, J = 3.8 Hz, 1H), 5.17 (d, J = 3.8 Hz, 1H), 5.23 (d, J = 3.8 Hz, 1H), 5.25 (d, J = 3.2 Hz, 1H), 5.56 (d, J = 3.2 Hz, 1H); ¹H NMR (500 MHz, CDCl3, 1D-TOCSY, irradiation at H1¹, 5.56 ppm): δ 3.45 (dd, J = 9.6, 2.9 Hz, 1H, H2¹), 3.96 (dd, J = 9.0, 9.0 Hz, 1H, H3¹), 4.18 (d, J = 8.8 Hz, 1H, H4¹); ¹H NMR (500 MHz, CDCl3, 1D-TOCSY, irradiation at H6¹, 4.45 ppm): δ 3.19 (dd, J = 9.7, 3.4 Hz, 1H, H2¹), 3.57 (dd, J = 9.3, 9.3 Hz, 1H, H3¹), 3.71 (dd, J = 9.5, 9.5 Hz, 1H, H4¹), 3.99 (m, 1H, H5¹), 4.30 (dd, J = 12.4, 3.8 Hz, 1H, H6¹), 5.12 (d, J = 3.0 Hz, 1H, H1¹); ¹³C NMR (125.7 MHz, CDCl3): δ 20.85 (CH₃), 58.16 (CH₃), 58.22 (CH₃), 58.28 (CH₃), 58.42 (CH₃), 58.62 (CH₃), 58.79 (CH₃), 58.97 (2 × CH₃), 59.02 (CH₃), 59.06 (CH₃), 59.13 (CH₃), 59.18 (CH₃), 60.53 (CH₃), 61.22 (CH₃), 61.23 (CH₃), 61.27 (CH₃), 61.43 (CH₃), 61.47 (2 × CH₃), 62.87 (CH₂), 68.67 (CH), 70.77 (CH₂), 70.77 (CH), 70.88 (CH), 70.90 (CH), 70.92 (CH), 70.99 (CH₂), 71.12 (CH), 71.14 (CH₂), 71.21 (CH₂), 71.41 (CH₂), 78.45 (CH), 78.63 (CH), 78.80 (CH), 79.29 (CH), 79.98 (CH), 80.03 (CH), 80.19 (CH), 80.39 (CH), 80.77 (CH), 81.31 (CH), 81.46 (CH), 81.71 (3 × CH), 81.80 (CH), 81.97 (CH), 82.14 (CH), 82.19 (CH), 82.27 (2 × CH), 82.38 (CH), 97.91 (CH), 98.11 (CH),
98.64 (CH), 99.02 (CH), 99.11 (CH), 99.22 (CH), 99.60 (CH), 168.56 (C), 170.41 (C); MS (ESI<sup>+</sup>-TOF): m/z (%) 1449 [(M + Na)<sup>+</sup>, 100]; HRMS (ESI<sup>+</sup>-TOF): m/z [M + Na]<sup>+</sup> calcd for C<sub>62</sub>H<sub>106</sub>NaO<sub>36</sub> 1449.6362; found 1449.6410. Anal. calcd for C<sub>62</sub>H<sub>106</sub>O<sub>36</sub>: C, 52.17; H, 7.48. Found: C, 52.46; H, 7.40.

Oxidative HAT of 2<sup>I-VI</sup>,3<sup>I-VI</sup>,6<sup>III-VI</sup>-hexadeca-O-methyl-α-cyclomaltohexaose (36). A solution of alcohol 36<sup>35</sup> (150 mg, 0.125 mmol) in dry CH<sub>2</sub>Cl<sub>2</sub> (5 mL) containing DIB (88 mg, 0.27 mmol) and I<sub>2</sub> (32 mg, 0.125 mmol) was stirred under nitrogen at 28 °C for 1.5 h while irradiated with two 80 W tungsten-filament lamps. The reaction mixture was then directly loaded onto a silica gel (TLC Silica gel 60 F<sub>254</sub>, scraped from Merck Aluminum sheets) column chromatography (hexanes–acetone, 65:35→55:45) to give the ten-membered lactone 37 (23 mg, 0.019 mmol, 16%), cyclo-5<sup>VI</sup>,6<sup>Ι</sup>-anhydro-(5<sup>VI</sup>R)-(2,3,6-tri-O-methyl-α-D-xylo-hexos-5-ulopyranosyl)-[(1→4)-2,3-di-O-methyl-α-D-glucopyranosyl]<sub>2</sub>[(1→4)-2,3,6-tri-O-methyl-α-D-glucopyranosyl]<sub>3</sub> (38) (37 mg, 0.031 mmol, 25%), and the lactone-alcohol 39 (14.6 mg, 0.012 mmol, 10%). Compound 37: colorless oil, [α]<sub>D</sub> +131.7 (c 1.31, CHCl<sub>3</sub>); IR (film): 2929, 1760, 1745, 1454, 1367, 1107, 1046 cm<sup>-1</sup>; <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ 2.12 (s, 3H), 3.09 (dd, J = 9.9, 3.3 Hz, 1H), 3.12 (dd, J = 10.1, 3.2 Hz, 1H), 3.17 (dd, J = 9.8, 3.5 Hz, 2H), 3.20 (dd, J = 10.1, 3.5 Hz, 1H), 3.36 (s, 3H), 3.36 (s, 3H), 3.36 (s, 3H), 3.38 (s, 3H), 3.44 (s, 3H), 3.48 (s, 3H), 3.49 (s, 3H), 3.50 (s, 3H), 3.52 (s, 6H), 3.55 (s, 3H), 3.59 (s, 3H), 3.60 (s, 3H), 3.60 (s, 3H), 3.70 (s, 3H), 3.74 (s, 3H), 3.87 (dd, J = 11.0, 3.8 Hz, 1H), 4.09 (d, J = 5.7 Hz, 1H), 4.33 (ddd, J = 10.1, 10.1, 5.4 Hz, 1H), 4.48 (dd, J = 10.7, 5.4 Hz, 1H), 4.61 (ddd, J = 10.1, 10.1, 5.4 Hz, 1H), 4.95 (d, J = 3.5 Hz, 2H), 5.02 (d, J = 3.5 Hz, 1H), 5.07 (d, J = 3.2 Hz, 2H), 6.37 (d, J = 1.3 Hz, 1H); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>, 1D-TOCSY, irradiation at H<sub>I</sub>δ, 6.37 ppm): δ 3.59 (d, J = 7.7 Hz, 1H, H<sub>2</sub>δ), 3.73 (m, 1H, H<sub>3</sub>δ), 4.09 (d, J = 5.8 Hz, 1H, H<sub>4</sub>δ); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>, 1D-
TOCSY, irradiation at H5\textsuperscript{II}, 4.61 ppm): $\delta$ 3.08 (dd, J = 9.9, 3.3 Hz, 1H, H2\textsuperscript{II}), 3.44 (dd, J = 9.1, 9.1 Hz, 1H, H3\textsuperscript{II}), 3.59 (dd, J = 9.6, 9.6 Hz, 1H, H4\textsuperscript{II}), 3.91 (dd, J = 10.6, 10.6 Hz, 1H, H6\textsuperscript{II}), 4.48 (dd, J = 10.8, 5.3 Hz, 1H, H6\textsuperscript{II}), 4.94 (d, J = 2.5 Hz, 1H, H1\textsuperscript{II}); $^1$H NMR (500 MHz, CDCl\textsubscript{3}, 1D-TOCSY, irradiation at H5\textsuperscript{VI}, 4.33 ppm): $\delta$ 3.19 (dd, J = 9.9, 2.9 Hz, 1H, H2\textsuperscript{VI}), 3.49 (dd, J = 9.6, 9.6 Hz, 1H, H4\textsuperscript{VI}), 3.68 (dd, J = 9.6, 9.6 Hz, 1H, H3\textsuperscript{VI}), 4.94 (d, J = 2.7 Hz, 1H, H1\textsuperscript{VI}); $^1$H NMR (500 MHz, CDCl\textsubscript{3}, 1D-ROESY, irradiation at H1\textsuperscript{I}, 6.37 ppm): $\delta$ 3.59 (dd, J = 9.3, 9.3 Hz, 1H, H4\textsuperscript{I}), 4.09 (d, J = 5.5 Hz, 1H, H4\textsuperscript{I}); $^1$H NMR (500 MHz, CDCl\textsubscript{3}, 1D-ROESY, irradiation at H4\textsuperscript{I}, 4.09 ppm): $\delta$ 4.95 (d, J = 3.5 Hz, 1H, H1\textsuperscript{VI}); $^{13}$C NMR (125.7 MHz, CDCl\textsubscript{3}): $\delta$21.48 (CH\textsubscript{3}), 57.71 (CH\textsubscript{3}), 57.76 (CH\textsubscript{3}), 58.02 (CH\textsubscript{3}), 58.25 (CH\textsubscript{3}), 58.33 (CH\textsubscript{3}), 58.56 (CH\textsubscript{3}), 58.89 (CH\textsubscript{3}), 58.91 (CH\textsubscript{3}), 58.99 (CH\textsubscript{3}), 60.13 (CH\textsubscript{3}), 60.68 (CH\textsubscript{3}), 61.30 (CH\textsubscript{3}), 61.40 (CH\textsubscript{3}), 61.66 (CH\textsubscript{3}), 61.99 (CH\textsubscript{3}), 62.10 (CH\textsubscript{3}), 62.60 (CH), 66.24 (CH\textsubscript{2}), 70.79 (CH), 70.82 (CH), 71.07 (CH\textsubscript{2}), 71.17 (CH), 71.35 (2 × CH\textsubscript{2}), 71.39 (CH), 71.51 (CH\textsubscript{2}), 79.60 (CH), 81.04 (CH), 81.11 (2 × CH), 81.14 (CH), 81.26 (CH), 81.40 (CH), 81.43 (CH), 81.52 (2 × CH), 81.65 (CH), 81.83 (2 × CH), 81.90 (CH), 81.96 (2 × CH), 82.33 (CH), 83.00 (CH), 94.66 (CH), 98.56 (CH), 98.90 (CH), 99.53 (CH), 99.64 (CH), 99.80 (CH), 170.02 (C), 170.23 (C); MS (ESI\textsuperscript{+}-TOF): m/z (%) 1245 [(M + Na\textsuperscript{+}), 100]; HRMS (ESI\textsuperscript{+}-TOF): m/z [M + Na\textsuperscript{+}] calcd for C\textsubscript{53}H\textsubscript{90}NaO\textsubscript{31} 1245.5364; found 1245.5369. Anal. calcd for C\textsubscript{53}H\textsubscript{90}O\textsubscript{31}: C, 52.04; H, 7.42. Found: C, 52.15; H, 7.33. Compound 38: colorless oil, [α]\textsubscript{D} +134.3 (c 1.32, CHCl\textsubscript{3}); IR (film): 3475, 2929, 1454, 1367, 1139, 1107, 1046 cm\textsuperscript{-1}; $^1$H NMR (500 MHz, CDCl\textsubscript{3}): $\delta$3.10–3.17 (m, 4H), 3.19 (dd, J = 9.9, 3.3 Hz, 1H), 3.25 (dd, J = 9.8, 3.2 Hz, 1H), 3.35 (s, 3H), 3.36 (s, 3H), 3.369 (s, 3H), 3.372 (s, 3H), 3.46 (s, 6H), 3.48 (s, 3H), 3.49 (s, 3H), 3.50 (s, 3H), 3.51 (s, 3H), 3.58 (s, 3H), 3.60 (s, 3H), 3.62 (s, 3H), 3.64 (s, 3H), 3.70 (s, 3H), 3.73 (s, 3H), 5.00 (d, J = 3.8 Hz, 1H), 5.01 (d, J = 3.8 Hz, 1H), 5.025 (d, J = 3.8 Hz, 1H), 5.033 (d, J = 4.1 Hz, 1H), 5.06 (d, J = 3.2 Hz, 1H), 5.18 (d, J = 3.5 Hz, 1H); $^{13}$C NMR (125.7 MHz, CDCl\textsubscript{3}): $\delta$57.66
(CH$_3$), 57.82 (CH$_3$), 57.94 (2 × CH$_3$), 58.34 (CH$_3$), 58.51 (CH$_3$), 58.74 (CH$_3$), 58.79 (CH$_3$), 59.07 (CH$_3$), 59.14 (CH$_3$), 61.26 (CH$_3$), 61.44 (CH$_3$), 61.52 (CH$_3$), 61.86 (CH$_2$), 61.93 (CH$_3$), 61.98 (CH$_3$), 62.24 (CH$_3$), 63.80 (CH$_2$), 66.28 (CH), 70.90 (CH), 70.93 (CH), 71.24 (CH$_2$), 71.57 (CH), 71.80 (CH$_2$), 71.94 (CH$_2$), 72.04 (CH), 78.29 (CH), 80.35 (CH), 80.56 (CH), 80.74 (CH), 80.96 (CH), 81.03 (CH), 81.07 (CH), 81.26 (CH), 81.40 (2 × CH), 81.72 (CH), 81.96 (CH), 82.01 (2 × CH), 82.23 (CH), 82.31 (CH), 82.59 (CH), 82.72 (CH), 97.35 (CH), 97.79 (CH), 98.77 (CH), 99.62 (CH), 99.90 (CH), 100.07 (CH), 100.81 (C); MS (ESI$^+$-TOF): m/z (%) 1217 [(M + Na)$^+$, 100]; HRMS (ESI$^+$-TOF): m/z [M + Na]$^+$ calcd for C$_{52}$H$_{90}$NaO$_{30}$ 1217.5415; found 1217.5419. Anal. calcd for C$_{52}$H$_{90}$O$_{30}$: C, 52.25; H, 7.59. Found: C, 52.43; H, 7.61. **Compound 39**: colorless oil, $[\alpha]_D +111.4$ (c 0.96, CHCl$_3$); IR (film): 3468, 2929, 1757, 1454, 1367, 1137, 1109, 1044 cm$^{-1}$; $^1$H NMR (500 MHz, CDCl$_3$): $\delta$ 3.13–3.20 (m, 3H), 3.23 (dd, J = 9.8, 3.5 Hz, 1H), 3.38 (s, 3H), 3.385 (s, 6H), 3.39 (s, 3H), 3.48 (s, 6H), 3.49 (s, 3H), 3.50 (s, 3H), 3.51 (s, 3H), 3.58 (s, 3H), 3.63 (s, 6H), 3.64 (s, 3H), 3.65 (s, 3H), 3.66 (s, 3H), 3.67 (s, 3H), 3.87 (dd, J = 8.5, 8.5 Hz, 1H), 4.03 (d, J = 8.5 Hz, 1H), 4.08–4.15 (m, 1H), 4.24 (ddd, J = 10.1, 2.8, 1.6 Hz, 1H), 5.06 (d, J = 3.5 Hz, 2H), 5.065 (d, J = 3.8 Hz, 1H), 5.12 (d, J = 3.5 Hz, 1H), 5.16 (d, J = 3.5 Hz, 1H), 5.47 (d, J = 2.8 Hz, 1H); $^1$H NMR (500 MHz, CDCl$_3$, 1D-TOCSY, irradiation at H$^1$I, 5.47 ppm): $\delta$ 3.45 (dd, J = 10.0, 2.7 Hz, 1H, H$^2$I), 3.88 (dd, J = 9.3, 9.3 Hz, 1H, H$^3$I), 4.03 (d, J = 8.7 Hz, 1H, H$^4$I); $^1$H NMR (500 MHz, CDCl$_3$, 1D-TOCSY, irradiation at H$^5$VI, 4.24 ppm): $\delta$ 3.24 (dd, J = 9.7, 3.6 Hz, 1H, H$^2$VI), 3.60 (dd, J = 10.0, 9.4 Hz, 1H, H$^4$VI), 3.69 (dd, J = 9.4, 9.4 Hz, 1H, H$^3$VI), 3.89 (dd, J = 10.7, 3.1 Hz, 1H, H$^6$VI), 5.17 (d, J = 2.9 Hz, 1H, H$^1$VI); $^1$H NMR (500 MHz, CDCl$_3$, 1D-ROESY, irradiation at H$^4$I, 4.03 ppm): $\delta$ 5.16 (d, J = 3.4 Hz, 1H, H$^1$I); $^{13}$C NMR (125.7 MHz, CDCl$_3$): $\delta$ 57.91 (4 × CH$_3$), 58.35 (CH$_3$), 58.91 (2 × CH$_3$), 58.98 (2 × CH$_3$), 59.14 (CH$_3$), 61.24 (CH$_3$), 61.56 (CH$_3$), 61.56 (CH$_2$), 61.62 (CH$_3$), 61.65 (CH$_3$), 61.75 (2 × CH$_3$), 71.08 (2 × CH), 63
71.08 (CH$_2$), 71.13 (CH), 71.21 (CH$_2$), 71.25 (CH$_2$), 71.34 (CH$_2$), 71.36 (CH), 71.65 (CH), 80.02 (CH), 80.13 (CH), 80.29 (CH), 81.20 (CH), 81.23 (CH), 81.26 (CH), 81.34 (CH), 81.51 (CH), 81.78 (CH), 81.84 (2 × CH), 81.89 (CH), 81.97 (CH), 82.01 (CH), 82.11 (2 × CH), 82.16 (CH), 82.32 (CH), 99.26 (CH), 99.39 (CH), 99.68 (CH), 99.74 (CH), 100.34 (CH), 100.37 (CH), 169.42 (C); MS (ESI$^+$-TOF): m/z (%) 1203 [(M + Na)$^+$, 100] HRMS (ESI$^+$-TOF): m/z [M + Na]$^+$ calcd for C$_{51}$H$_{88}$NaO$_{30}$ 1203.5258; found 1203.5254. Anal. calcd for C$_{51}$H$_{88}$O$_{30}$: C, 51.86; H, 7.51. Found: C, 51.74; H, 7.40.

*Methyl 6-O-tert-Butyldiphenylsilyl-2,3,4-tri-O-methyl-α-D-glucopyranosyl-(1→4)-6-O-tert-butyldiphenylsilyl-2,3-di-O-methyl-β-D-glucopyranoside (40β) and Methyl 6-O-tert-Butyldiphenylsilyl-2,3,4-tri-O-methyl-α-D-glucopyranosyl-(1→4)-6-O-tert-butyldiphenylsilyl-2,3-di-O-methyl-α-D-glucopyranoside (40α).* To a solution of dry D-(+) maltose (5 g, 14.6 mmol), imidazole (8.9 g, 131.6 mmol), and DMAP (8.9 g, 73.1 mmol) in dry DMF (75 mL) was added TBDPSCl (22.8 mL, 87.7 mmol) at 0°C. The mixture was stirred at room temperature for 20 h, poured into water, and extracted with CH$_2$Cl$_2$. The organic phase was washed with HCl (10%), saturated aqueous NaHCO$_3$, and concentrated under reduced pressure to give a residue that was dried over P$_2$O$_5$ in a high vacuum desiccator for 24 h and used in the subsequent reaction as a mixture without further purification. NaH (60%, 7 g, 175 mmol) was added in portions to a solution of the crude residue (32 g) in dry DMF (1 mL) cooled to 0°C and the mixture stirred at this temperature for 1 h. MeI (13.7 mL, 219.3 mmol) was then added dropwise and the stirring continued for 3 h at room temperature. The excess of NaH was destroyed with MeOH and the mixture poured into water, extracted with CH$_2$Cl$_2$ and concentrated under reduced pressure. The residue was purified by silica gel column chromatography (hexanes–EtOAc, 9:1→8:2) and only the disilylated anomeric isomers 40β (4.3 g, 4.76 mmol, 33%) and 40α (1.6 g, 1.78 mmol, 12%)
were studied. Compound 40β: $R_f = 0.49$ (n-hexane–EtOAc, 7:3), colorless oil, $[\alpha]_D +44.5$ (c 1.19, CHCl$_3$); IR (film): 2933, 1148, 1104 cm$^{-1}$; $^1$H NMR (500 MHz, CDCl$_3$): $\delta$ 0.94 (s, 9H), 1.00 (s, 9H), 3.04 (dd, $J = 9.1$, 7.9 Hz, 1H), 3.12 (dd, $J = 9.6$, 3.9 Hz, 1H), 3.31 (m, 1H), 3.39 (m, 3H), 3.45 (dd, $J = 8.8$, 8.8 Hz, 1H), 3.48 (m, 2H), 3.53 (s, 3H), 3.55 (s, 3H), 3.57 (s, 3H), 3.59 (s, 3H), 3.60 (s, 3H), 3.64 (s, 3H), 3.66 (dd, $J = 9.8$, 8.8 Hz, 1H), 3.74 (dd, $J = 11.0$, 5.7 Hz, 1H), 3.85 (dd, $J = 11.0$, 2.2 Hz, 1H), 4.17 (d, $J = 7.6$ Hz, 1H), 5.55 (d, $J = 4.1$ Hz, 1H), 7.15–7.66 (m, 20H); $^1$H NMR (500 MHz, CDCl$_3$, 1D-TOCSY, irradiation at H1$^1$, 4.17 ppm): $\delta$ 3.04 (dd, $J = 8.8$, 8.1 Hz, 1H, H2$^1$), 3.41 (m, 1H, H5$^1$), 4.45 (dd, $J = 8.9$, 8.9 Hz, 1H, H3$^1$), 3.66 (dd, $J = 9.3$, 9.3 Hz, 1H, H4$^1$), 3.74 (dd, $J = 11.2$, 5.9 Hz, 1H, H6a$^1$), 3.84 (dd, $J = 10.9$, 2.3 Hz, 1H, H6b$^1$); $^1$H NMR (500 MHz, CDCl$_3$, 1D-TOCSY, irradiation at H1$^{II}$, 5.55 ppm): $\delta$ 3.11 (dd, $J = 9.3$, 3.4 Hz, 1H, H2$^{II}$), 3.31 (m, 1H, H5$^{II}$), 3.37 (m, 2H, H3$^{II}$ and H4$^{II}$), 3.46 (br d, $J = 11.1$ Hz, 1H, H6a$^{II}$), 3.52 (br d, $J = 11.3$ Hz, 1H, H6b$^{II}$); $^{13}$C NMR (125.7 MHz, CDCl$_3$): $\delta$ 19.23 (C), 19.31 (C), 26.79 (3 × CH$_3$), 26.81 (3 × CH$_3$), 56.67 (CH$_3$), 59.15 (CH$_3$), 59.58 (CH$_3$), 60.16 (CH$_3$), 60.18 (CH$_3$), 60.71 (CH$_3$), 62.14 (CH$_2$), 63.61 (CH$_2$), 71.89 (CH), 72.06 (CH), 75.11 (CH), 78.84 (CH), 81.90 (CH), 83.38 (CH), 83.77 (CH), 86.41 (CH), 96.12 (CH), 104.04 (CH), 127.43 (2 × CH), 127.47 (2 × CH), 127.49 (2 × CH), 127.54 (2 × CH), 129.36 (CH), 129.43 (CH), 129.47 (2 × CH), 133.27 (C), 133.34 (C), 133.72 (C), 133.86 (C), 135.38 (2 × CH), 135.56 (2 × CH), 135.77 (4 × CH); MS (ESI$^+$-TOF): $m/z$ (%) 925 [(M + Na)$^+$, 100]; HRMS (ESI$^+$-TOF): $m/z$ [M + Na]$^+$ calcd for C$_{50}$H$_{70}$NaO$_{11}$Si$_2$ 925.4354; found 925.4366. Anal. calcd for C$_{50}$H$_{70}$O$_{11}$Si$_2$: C, 66.49; H, 7.81. Found: C, 66.75; H, 7.93. Compound 40α: $R_f = 0.42$ (n-hexane–EtOAc, 7:3), colorless oil, $[\alpha]_D +86.3$ (c 1.067, CHCl$_3$); IR (film): 2933, 1156, 1104, 1029 cm$^{-1}$; $^1$H NMR (500 MHz, CDCl$_3$): $\delta$ 0.94 (s, 9H), 0.99 (s, 9H), 3.11 (dd, $J = 9.8$, 4.1 Hz, 1H), 3.17 (dd, $J = 9.6$, 3.6 Hz, 1H), 3.36 (s, 3H), 3.51 (s, 6H), 3.56 (s, 3H), 3.60 (s, 3H), 3.63 (s, 3H), 4.78 (d, $J = 3.5$ Hz, 1H), 5.55 (d, $J = 4.1$ Hz, 1H), 7.16–7.66 (m, 20H); $^{13}$C NMR
(125.7 MHz, CDCl$_3$): $\delta$ 19.24 (C), 19.30 (C), 26.81 (6 × CH$_3$), 54.59 (CH$_3$), 58.69 (CH$_3$), 59.52 (CH$_3$), 60.00 (CH$_3$), 60.14 (CH$_3$), 60.70 (CH$_3$), 62.22 (CH$_2$), 63.63 (CH$_2$), 70.66 (CH), 71.92 (CH), 72.48 (CH), 78.86 (CH), 81.93 (CH), 82.43 (CH), 83.37 (CH), 83.52 (CH), 96.27 (CH), 96.50 (CH), 127.42 (2 × CH), 127.44 (2 × CH), 127.46 (2 × CH), 127.53 (2 × CH), 129.35 (CH), 129.39 (CH), 129.43 (CH), 129.45 (CH), 133.38 (C), 133.41 (C), 133.77 (C), 133.88 (C), 135.48 (2 × CH), 135.60 (2 × CH), 135.72 (2 × CH), 135.80 (2 × CH); MS (ESI$^+$-TOF): $m/z$ (%) 925 [(M + Na)$^+$, 100]; HRMS (ESI$^+$-TOF): $m/z$ [M + Na]$^+$ calcd for C$_{50}$H$_{70}$NaO$_{11}$Si$_2$ 925.4354; found 925.4358.

Methyl 2,3,4-Tri-O-methyl-α-D-glucopyranosyl-(1→4)-2,3-di-O-methyl-β-D-glucopyranoside (41). To a solution of 40β (2 g, 2.2 mmol) in THF (50 mL) was added TBAF (1 M in THF, 6.6 mL, 6.6 mmol) and the mixture stirred at room temperature for 7 h. The THF was the removed under vacuum and the resulting oil purified by column chromatography (hexanes–EtOAc, 3:7→EtOAc) to give the compound 41 (675 mg, 1.58 mmol, 72%) as a colorless oil; $[\alpha]_D^{+} + 74.1$ (c 1.077, CHCl$_3$); IR (film): 3460, 2936, 1144, 1081, 1029 cm$^{-1}$; $^1$H NMR (500 MHz, CDCl$_3$): $\delta$ 3.00 (dd, $J = 9.8, 8.8$ Hz, 1H), 3.03 (dd, $J = 9.1, 7.9$ Hz, 1H), 3.15 (dd, $J = 9.8, 4.1$ Hz, 1H), 3.35 (ddd, $J = 9.8, 2.8, 2.8$ Hz, 1H), 3.43 (dd, $J = 10.4, 9.1$ Hz, 1H), 3.43 (dd, $J = 8.8, 8.8$ Hz, 1H), 3.52 (s, 3H), 3.53 (s, 3H), 3.55 (s, 3H), 3.57 (s, 3H), 3.58 (s, 3H), 3.60 (s, 3H), 3.63 (s, 3H), 3.65 (dd, $J = 11.4, 6.0$ Hz, 1H), 3.81 (dd, $J = 12.3, 2.8$ Hz, 1H), 3.85 (dd, $J = 12.3, 3.2$ Hz, 1H), 3.85 (dd, $J = 11.7, 2.2$ Hz, 1H), 3.88 (dd, $J = 9.5, 9.5$ Hz, 1H), 4.19 (d, $J = 7.9$ Hz, 1H), 5.61 (d, $J = 4.1$ Hz, 1H); $^{13}$C NMR (125.7 MHz, CDCl$_3$): $\delta$ 57.0 (CH$_3$), 59.9 (CH$_3$), 60.1 (CH$_3$), 60.2 (CH$_3$), 60.6 (CH$_3$), 60.8 (CH$_3$), 60.9 (CH$_2$), 61.9 (CH$_2$), 71.3 (CH), 72.2 (CH), 74.2 (CH), 80.5 (CH), 81.8 (CH), 83.5 (CH), 84.3 (CH), 86.5 (CH), 96.5 (CH), 104.3 (CH); MS (ESI$^+$-TOF): $m/z$ (%) 449 [(M + Na)$^+$, 100]; HRMS

Oxidative HAT of Methyl 2,3,4-Tri-O-methyl-α-D-glucopyranosyl-(1→4)-2,3-di-O-methyl-β-D-glucopyranoside (41). A solution of alcohol 41 (100 mg, 0.235 mmol) in dry CH₂Cl₂ (9.4 mL) containing DIB (166.5 mg, 0.516 mmol) and I₂ (60 mg, 0.235 mmol) was stirred under nitrogen at 32 °C for 3 h while irradiated with two 80 W tungsten-filament lamps. The reaction mixture was then directly loaded onto a silica gel (TLC Silica gel 60 F₂₅₄, scraped from Merck Aluminum sheets) column chromatography (hexanes–acetone, 60:40→25:75) to give the ten-membered lactone 42 (11 mg, 0.024 mmol, 9%) and the acetyl-derivative 43 (11 mg, 0.023 mmol, 10%). Compound 42: colorless oil, [α]D –24.2 (c 0.520, CHCl₃); IR (film): 2938, 1746, 1455, 1372, 1126, 1095 cm⁻¹; ¹H NMR (500 MHz, CDCl₃): δ 2.13 (s, 3H), 2.89 (dd, J = 9.3, 7.7 Hz, 1H, H₂II), 3.15 (dd, J = 8.8, 8.8 Hz, 1H, H₃II), 3.31 (dd, J = 7.7, 2.0 Hz, 1H, H₂I), 3.45 (s, 3H), 3.49 (s, 3H), 3.53 (s, 3H), 3.56 (dd, J = 8.4, 8.4 Hz, 1H, H₃I), 3.59 (s, 3H), 3.60 (s, 3H), 3.61 (s, 3H), 3.62 (dd, J = 9.1, 9.1 Hz, 1H, H₄I), 3.75 (d, J = 8.8 Hz, 1H, H₄II), 3.80 (ddd, J = 9.8, 9.8, 5.4 Hz, 1H, H₅II), 4.12 (d, J = 7.9 Hz, 1H, H₁I), 4.18 (dd, J = 11.4, 9.8 Hz, 1H, H₆I), 4.59 (dd, J = 11.2, 5.2 Hz, 1H, H₆II), 6.23 (d, J = 2.2 Hz, 1H, H₁I); ¹H NMR (500 MHz, CDCl₃, 1D-TOCSY, irradiation at H₁I, 6.23 ppm): δ 3.31 (dd, J = 7.8, 1.9 Hz, 1H, H₂I), 3.56 (dd, J = 8.4, 8.4 Hz, 1H, H₃I), 3.76 (d, J = 8.7 Hz, 1H, H₄I); ¹H NMR (500 MHz, CDCl₃, 1D-TOCSY, irradiation at H₂II, 2.89 ppm): δ 3.15 (dd, J = 8.9, 8.9 Hz, 1H, H₃II), 3.63 (dd, J = 9.2, 9.2 Hz, 1H, H₄II), 3.81 (ddd, J = 9.3, 9.3, 5.4 Hz, 1H, H₅II), 4.13 (d, J = 7.8 Hz, 1H, H₁II), 4.19 (dd, J = 10.6, 10.6 Hz, 1H, H₆II), 4.60 (dd, J = 11.0, 5.2 Hz, 1H, H₆II); ¹³C NMR (125.7 MHz, CDCl₃): δ 21.34 (CH₃), 57.08 (CH₃), 59.00 (CH₃), 60.43 (CH₃), 61.64 (2 × CH₃), 62.00 (CH₃), 65.14 (CH₂, C₆II), 67.32 (CH, C₅II), 81.51 (CH, C₄I), 82.37 (CH, C₄II), 83.04 (CH, C₃I), 83.22 (CH, C₂II), 83.88 (CH, C₂I), 84.71 (CH, C₃II), 94.51 (CH, C₁I), 103.82 (CH,
C1\textsuperscript{Ii}, 170.32 (C), 170.60 (C); MS (ESI\textsuperscript{+}-TOF): m/z (%) 475 [(M + Na\textsuperscript{+}), 100]; HRMS (ESI\textsuperscript{+}-TOF): m/z [M + Na\textsuperscript{+}] calcd for C\textsuperscript{19}H\textsubscript{32}NaO\textsubscript{12} 475.1791; found 475.1793. Anal. calcd for C\textsuperscript{19}H\textsubscript{32}O\textsubscript{12}: C, 50.44; H, 7.13. Found: C, 50.16; H, 7.21. Compound 43: colorless oil, [\alpha]_D \textsuperscript{+} 43.8 (c 0.820, CHCl\textsubscript{3}); IR (film): 3520, 2933, 1745, 1456, 1369, 1107, 1081, 1055 cm\textsuperscript{-1}; \textsuperscript{1}H NMR (500 MHz, CDCl\textsubscript{3}): \delta 2.05 (s, 3H), 3.05 (dd, J = 9.1, 7.6 Hz, 1H, H2\textsuperscript{II}), 3.29 (ddd, J = 9.8, 3.0, 3.0 Hz, 1H, H5\textsuperscript{II}), 3.34 (dd, J = 9.0, 9.0 Hz, 1H, H3\textsuperscript{II}), 3.47 (s, 3H), 3.52 (s, 3H), 3.56 (s, 3H), 3.58 (s, 3H), 3.61 (s, 3H), 3.78 (dd, J = 9.5, 9.5 Hz, 1H, H4\textsuperscript{II}), 3.86 (dd, J = 12.3, 2.8 Hz, 1H, H6\textsuperscript{II}), 3.94 (d, J = 12.0 Hz, 1H, H6\textsuperscript{I}), 4.09 (dd, J = 12.3, 3.2 Hz, 1H, H6\textsuperscript{II}), 4.09 (d, J = 1.6 Hz, 1H, H4\textsuperscript{I}), 4.18 (d, J = 7.6 Hz, 1H, H1\textsuperscript{II}), 4.59 (d, J = 11.7 Hz, 1H, H6\textsuperscript{I}), 4.69 (d, J = 6.3 Hz, 1H), 5.09 (d, J = 6.3 Hz, 1H), 5.58 (d, J = 2.5 Hz, 1H, H1\textsuperscript{I}); \textsuperscript{1}H NMR (500 MHz, CDCl\textsubscript{3}, 1D-TOCSY, irradiation at H2\textsuperscript{II}, 3.05 ppm): \delta 3.29 (ddd, J = 9.9, 2.9, 2.9 Hz, 1H, H5\textsuperscript{II}), 3.34 (dd, J = 9.0, 9.0 Hz, 1H, H3\textsuperscript{II}), 3.78 (dd, J = 9.4, 9.4 Hz, 1H, H4\textsuperscript{II}), 3.86 (dd, J = 12.2, 3.4 Hz, 1H, H6\textsuperscript{II}), 4.09 (dd, J = 12.0, 2.4 Hz, 1H, H6\textsuperscript{II}), 4.18 (d, J = 7.6 Hz, 1H, H1\textsuperscript{II}), 4.59 (d, J = 11.7 Hz, 1H, H6\textsuperscript{I}), 4.69 (d, J = 6.3 Hz, 1H), 5.09 (d, J = 6.3 Hz, 1H), 5.58 (d, J = 2.5 Hz, 1H, H1\textsuperscript{I}); \textsuperscript{13}C NMR (125.7 MHz, CDCl\textsubscript{3}): \delta 21.89 (CH\textsubscript{3}), 56.82 (CH\textsubscript{3}), 57.88 (CH\textsubscript{3}), 59.83 (CH\textsubscript{3}), 60.23 (CH\textsubscript{3}), 60.43 (CH\textsubscript{3}), 61.17 (CH\textsubscript{2}, C6\textsuperscript{II}), 71.11 (CH\textsubscript{2}, C6\textsuperscript{I}), 74.44 (CH, C4\textsuperscript{I}), 74.66 (CH, C5\textsuperscript{II}), 75.13 (CH, C4\textsuperscript{II}), 76.40 (CH, C2\textsuperscript{I} or C3\textsuperscript{I}), 79.17 (CH, C2\textsuperscript{I} or C3\textsuperscript{I}), 84.11 (CH, C2\textsuperscript{II}), 86.22 (CH, C3\textsuperscript{II}), 92.88 (CH\textsubscript{2}), 96.45 (C, C5\textsuperscript{I}), 97.16 (CH, C1\textsuperscript{I}), 104.29 (CH, C1\textsuperscript{II}), 168.79 (C); MS (ESI\textsuperscript{+}-TOF): m/z (%) 505 [(M + Na\textsuperscript{+}), 100]; HRMS (ESI\textsuperscript{+}-TOF): m/z [M + Na\textsuperscript{+}] calcd for C\textsubscript{20}H\textsubscript{34}NaO\textsubscript{13} 505.1897; found 505.1905. Anal. calcd for C\textsubscript{20}H\textsubscript{34}O\textsubscript{13}: C, 49.79; H, 7.10. Found: C, 49.46; H, 7.15.
The Supporting Information is available free of charge on the ACS Publications website at DOI:

The material includes crystal structure of 15; copies of spectra for all new compounds; absolute energy and Cartesian coordinates of minimized structures of compounds 32 and 38 (PDF).

Crystallographic data (CIF).

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Notes

The authors declare no competing financial interest.

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(21) Coupling constants and dihedrals were calculated with Maestro 9.9.013 on minimized structures using MacroModel and the AMBER* force field with the GB/SA solvent model for CHCl₃, Schrödinger, LLC, New York, NY, 2009. The experimental constants were determined on the basis of 1D TOCSY experiments.

(22) Although, according to the IUPAC nomenclature the unit VII is more usually named as (5R)-5₁VII-O-acetyl-2,3,6-tri-O-methyl-α-D-xylo-hexos-5-ulopyranosyl we prefer to see it as a 5-C-acetoxy-β-L-idopyranosyl derivative, which described better the inversion at C₅₁VII and the 1₄C₄ conformation of the pyranose ring and is also alternatively accepted by the IUPAC (see a similar example in Nomenclature of Carbohydrates, Rule 2-Carb-16.1).


(30) CCDC 1039442 contains the supplementary crystallographic data for the compound 15. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif.
(31) Steiner, T.; Saenger, W. Carbohydr. Res. 1996, 282, 53–63. Crystallographic data were obtained by courtesy of The Cambridge Crystallographic Data Centre (Database code CSD = TEVCEC).


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