- 1 Title: Projected impact of 21st century climate change on diapause in *Calanus finmarchicus*
- 2 Running title: Warming impacts on *C. finmarchicus* diapause
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12 Abstract

13 High latitude zooplankton typically spend over six months diapausing in deep waters each year, where metabolic demands are met by lipid reserves. The potential time an animal can 14 15 diapause for is related to overwintering respiration rates, lipid levels and body size. Climate change is therefore expected to reduce diapause duration, which will affect population 16 dynamics and regional biological carbon pumps. However, geographic variations in the 17 18 impact of climate change and potential behavioural adaptations to changing temperatures are 19 poorly understood. Here, we project changes in potential diapause duration over the 21st century in the key North Atlantic copepod Calanus finmarchicus under IPCC RCP 8.5 using 20 21 a new bioenergetic model. Potential diapause duration was related to surface and deep-water temperature and body size. Climate change has two key effects: increased deep-water 22 23 temperature will increase metabolism and shorten diapause, whereas surface warming will 24 reduce body size and relative lipid reserves and thus shorten diapause. The projections show 25 pronounced geographic variation in the impact of climate change. Northwestern Atlantic 26 populations see reductions in potential diapause duration of over 30%, whereas the overwintering centre in the Norwegian Sea will only see marginal changes. In deep basins 27 28 such as the Labrador Sea animals can potentially diapause in deeper waters, which experience 29 significantly less warming, but the ability to control diapause depth may be limited. Animals exiting diapause significantly earlier in the Northwest Atlantic is likely to have a significant 30 impact on ecosystems and disrupt the biological lipid pump. 31

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36 Introduction

37 The continued and largely unabated emissions of anthropogenic greenhouse gases (Friedlingstein et al., 2014) and the fundamental inertia of global energy systems (Davis and 38 Socolow, 2014) mean that large increases in oceanic temperatures are probably unavoidable. 39 These changes are already affecting marine ecosystems (Doney et al., 2012), and future 40 pressures on fisheries will have significant economic consequences (Barange et al., 2014). 41 42 Changes in zooplankton populations are particularly important because of their impacts on higher trophic levels (Blanchard et al., 2012; Chust et al., 2014) and zooplankton's role as 43 prey for many commercially important fish (Beaugrand and Kirby, 2010). It is therefore a 44 45 priority of zooplankton research to anticipate the responses of individual species and communities to rising temperatures. 46

Calanoid copepod species frequently dominate North Atlantic mesozooplankton communities
(Head *et al.*, 2003). The most well studied, and arguably most important, is *Calanus finmarchicus* (Melle *et al.*, 2014), whose geographic range extends from the Gulf of Maine to
the North Sea (Planque and Fromentin, 1996). Oceanic warming has caused a significant
northward shift in its geographic distribution in recent decades (Barnard *et al.*, 2004; Chust *et al.*, 2013).

Oceanic warming will not be uniform in the North Atlantic (Collins *et al.*, 2013). Some regions will warm significantly less than others (Drijfhout *et al.*, 2012), while deep water warming will lag that at the surface (Li *et al.*, 2013). In addition, weakening of the Atlantic Meridional Overturning Circulation (AMOC) could result in cooling in some regions (Banks and Gregory, 2006). These complexities result in difficult to predict regional changes, but it is clear that they will result in the general northward shift of species (Reygondeau and Beaugrand, 2011).

60 Research on the impact of climate change on C. finmarchicus's biogeography has focused almost exclusively on changes in surface temperatures (e.g. Reygondeau and Beaugrand 61 (2011); Hinder et al. (2013); Chust et al. (2013)). However, in response to low food in 62 63 autumn and winter, C. finmarchicus populations spend over half of the year diapausing in deep waters (Svetlichny et al., 1998; Johnson and Checkley, 2004; Falk-Petersen et al., 64 2009). During diapause, animals reduce their metabolism (Maps et al., 2014) and do not feed 65 66 (Hirche, 1996). C. finmarchicus is probably the only calanoid copepod species where quantitative empirical knowledge is advanced enough to allow modelling of diapause 67 68 duration. Furthermore, diapause of C. finmarchicus acts as an important carbon sink in the North Atlantic (Jønasdøttir et al. 2015). C. finmarchicus is therefore an excellent target 69 species if we want to understand the impacts of climate change on diapause 70 71 Existing models of diapause (Ingvarsdøttir et al., 1999; Saumweber & Durbin, 2006; Maps et

al., 2014) have focused on the key role of lipids (Irigoien, 2004). Before diapausing as copepodite stage C5, animals acquire large lipid reserves to meet the metabolic requirements of overwintering (Ingvarsdøttir *et al.*, 1999) and the energy costs of molting and gonad formation (Rey-Rassat *et al.*, 2002). Potential diapause duration is therefore a result of the combined effects of pre-diapause lipid levels, the costs of metabolism, and the energy requirements of molting and gonad formation (Ingvarsdøttir *et al.*, 1999; Saumweber and Durbin, 2006).

Effective modelling of potential diapause duration therefore requires understanding the key
determinants of lipid levels and overwintering metabolism. Evidence shows that body size
(Miller *et al.*, 2000) and food quantity and quality (Gatten *et al.*, 1980; Hygum *et al.*, 2000)
are the main influences on lipid levels. Furthermore, field evidence shows that there is a steep
allometric scaling of lipid reserves with prosome length (Miller *et al.*, 2000; Saumweber and
Durbin, 2006; Pepin *et al.*, 2011; Bergvik *et al.*, 2012). The two key influences on metabolic

costs are temperature (Saumweber and Durbin, 2006) and the scaling of metabolism with
body size (Maps *et al.*, 2014).

Quantitative knowledge is only sufficiently advanced to explicitly incorporate the influence 87 88 of body size and temperature into a model of diapause duration. Body size varies geographically and is largely determined by temperature (Wilson et al., 2015). We can 89 therefore model the geographic variation of body size and overwintering metabolism with 90 reasonable credibility using depth resolved spatial temperature data. However, there is 91 92 currently insufficient data to model the impact of geographic variations of food quality and quantity on body size and lipid reserves of animals. We therefore produced a model that 93 94 relates potential diapause duration to body size and overwintering temperature. 95 A recent large-scale analysis of field data showed that diapause duration is 200-250 days in the North Atlantic, with a median duration of approximately 200 and 250 days in the 96 97 Northwest and Northeast Atlantic respectively (Melle et al., 2014). Temperature variation 98 was put forward as an explanation for the difference between the eastern and western North 99 Atlantic. However, no existing study has modelled the potential diapause duration of C. finmarchicus populations and compared model predictions with field estimates. The present 100 101 study marks the first attempt to do so and to consider the geographic variation in changes that will result from climate change. 102

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105 Materials and methods

106 **Potential diapause duration model**

107 We define potential diapause duration as the maximum time an animal can diapause for. This was calculated under the following biological and ecological assumptions. Temperature 108 experienced during development determines prosome length. Body size determines maximum 109 110 pre-diapause lipid reserves. Animals must have sufficient lipid reserves remaining after diapause for molting and gonad formation. Structural body mass and temperature determine 111 112 metabolism, and hence the rate at which animals deplete their reserves. Potential diapause duration is therefore the time taken for an animal to deplete its lipid reserves to the level 113 required for post-diapause gonad formation and molting. The model's mathematical 114 equations are detailed in Table 1 and the literature used to derive relevant biological 115 parameters is given in Table S1. Here we will provide an overview of the biological and 116 ecological rationale for the model, while referencing the relevant equations. 117

We assume a linear relationship between prosome length and temperature (equation 1). Food quantity also influences body size (Campbell *et al.*, 2001), however large scale patterns of *C*. *finmarchicus* body size can be successfully reproduced with a linear relationship between prosome length and temperature (Wilson *et al.*, 2015). Furthermore, existing knowledge of the influence of food on body size is insufficient to credibly model body length in terms of food and temperature. Structural, i.e. nitrogen, weight is then calculated using the relevant allometric scaling (equation 2).

Maximum pre-diapause oil sac volume scales allometrically with prosome length (equation
3). Oil sac volume is then converted to wax ester carbon (equation 4). There is limited
knowledge of the lipid requirements for post-diapause molting and gonad formation. We
therefore assume that there is a simple linear relationship between post-diapause lipid

129	requirements and nitrogen weight (equation 5). This is estimated using the laboratory data of
130	Rey-Rassat <i>et al.</i> (2002). Lipid reserves available for metabolism during diapause, WE_{α} , is
131	thus defined as the difference between maximum pre-diapause lipid reserves and the lipid
132	requirements for post-diapause molting and gonad formation (equation 6). This is then
133	simplified to an allometric relationship with length (equation 6).
134	Respiration rates are related to structural weight and temperature (equations 7-10). The
135	oxygen respiration rate (equation 7) is converted to a carbon respiration rate (equation 8).
136	This is then converted from being related to nitrogen weight to being related to length
137	(equation 8).
138	Potential diapause duration is then the time taken to deplete WE_{α} (equation 11). Finally, we
139	simplify the relationship and relate potential diapause duration to prosome length and

140 overwintering temperature (equation 14).

Table 1: Outline of potential diapause duration model

144 $T_{s} = Surface (development) temperature(°C)L = prosome length of diapausers (mm)OSV_max = Maximum oil sac volume (mm3)T_{D} = Diapause temperature (°C)w = Nitrogen weight (µg)WE_max = Maximum pre-diapau(µg C)w = Nitrogen weight (µg)145WE_{\omega} = Post-diapause WErequirements (µg C)r = Respiration rate (µmol O_2g N-1 h-1)RQ = Respiratory quotient(µmol CO_2\mumol O_2^{-1})WE_max = Maximum pre-diapau(µg C)WE_max = WE for diapause metabolR = Respiration rate (µg C µN^{-1})RQ = Respiratory quotient(µmol CO_2\mumol O_2^{-1})Equationw = c \times L^dRemark(1) Length assumed to betemperature (Wilson A(2) Nitrogen weight assumed to betemperature (Wilson A(2009),WE_max = 0.74 \times \frac{900 \times OSV_{max} + 10.8}{1.44}WE_{\omega} = 3 \times wWE_{\omega} = 3 \times wWE_{\omega} = WE_{max} - WE_{\omega} \cong \alpha L^{\beta}(5) Approximation fromal. (2002).r = \mu w^{0.75} Q_{10}^{TD} / 1010^{6}(6) Derived from SaumwDurbin (2006).(7) Respiration rate convcarbon rate.R = \frac{24 \times RQ \times 12.011 \times r}{10^{6}}R = \frac{70}{R_0} \frac{T_D}{R_0}(6) Derived from SaumwDurbin rate radiu$		
144 $L = \text{prosome length of diapausers (mm)} \\ OSV_{max} = Maximum oil sac volume (mm 3) 145 WEw =Post-diapause WE requirements (µg C) r = Respiration rate (µmol O2g N-1 h-1) RQ = Respiratory quotient (µmol O2g mol O2-1) Equation K = Respiratory quotient (µmol O2g mol O2-1)EquationW = c \times L^dOSVmax = e × LfOSVmax = e × LfWE_{wax} = 0.74 \times \frac{900 \times OSV_{max} + 10.8}{1.44}WEw = 3 × wWEw = WEmax - WEw \cong \alpha L^{\beta}R = \frac{24 \times RQ \times 12.011 \times r}{10^6}R = \frac{24 \times RQ \times 12.011 \times r}{10^6}R = \frac{24 \times RQ \times 12.011 \times r}{10^6}R = \frac{7D}{2} (c_{x})R = \frac{7D}{2} (c_{x})R = \frac{7D}{2} (c_{x})R = \frac{24 \times RQ \times 12.011 \times r}{10^6}R = \frac{24 \times 12.011 \times r}{10^6}R = 24 \times 12.011 \times $	T_D = Diapause temperature (°C) w = Nitrogen weight (µ a)	
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$t_{a} = \frac{T_{a}}{\xi c^{0.75} Q_{10}} \frac{T_{D}}{T_{10}}$		
$\frac{a}{2} - \frac{a}{2} $		

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151 Model scenarios

We report two model scenarios. First, we estimated potential diapause duration in the North Atlantic under current environmental conditions (2005-2012). This scenario was used to validate the model compared with field estimates of current diapause duration. We then estimated changes in potential diapause duration throughout the North Atlantic under the IPCC Representative Concentration Pathway 8.5 (RCP 8.5) (Riahi *et al.*, 2011). RCP 8.5 is a high emissions scenario; however, current greenhouse gas emissions are closely tracking this scenario (Sanford *et al.*, 2014).

The ecological assumptions for the scenarios are as follows. Field estimates show that

diapause begins in late summer (Melle et al., 2014). We therefore assume that animals enter 160 diapause in late summer, with prosome length being determined by the mean temperature at a 161 depth of 20 m in July and August. A biological map of diapause depth was created using a 162 loess smooth through the median diapause depths reported by Heath et al. (2004), who 163 collated an extensive spatial data set of 13th, 50th and 83rd percentile of diapause depths. 164 Oceanic temperatures for the period 2005-2012 were obtained from NOAA's World Ocean 165 Atlas 2013 (WOA13) (Locarnini et al., 2013). WOA13 is resolved at a geographic resolution 166 167 of 1 °W by 1 °N and depths are resolved at a resolution of 5, 25 and 50 m in the intervals 0-100, 100-500 and 500-1500 m. We use the statistical mean temperature at the surface and 168 diapause depths for the relevant months over the period 2005-2012, with overwintering 169

temperature being the mean temperature between September and March.

The physical model used to project changes in oceanic temperature in the 21st century was 171 version 3.2 of the state of the art Nucleus for European Modelling of the Ocean (NEMO) 172 model (Madec, 2012). Complete details of forcings etc. are given in Yool et al. (2013). The 173 horizontal resolution of NEMO is approximately $1 \times 1^{\circ}$, with an increased resolution at the 174 equator to provide a more realistic representation of equatorial upwelling. There are 64 175 vertical levels, with the thickness of levels increasing with depth. Projected temperatures for 176 the decades 2000-2009 and 2090-2099 were calculated by forcing the NEMO model using 177 RCP 8.5. Mean temperature at the surface and diapause depth for each decade were used to 178 179 estimate mean potential diapause duration for each location in the two decades. 180 Diapause duration is potentially sensitive to the overwintering depth of animals because of

temperature variation, and thus respiration rate, with depth. We therefore projected the
change in temperature profile at a series of locations in the North Atlantic which have depth
profiles of diapausing populations (locations shown in Fig. S1), and then compared the

evolution of the temperature profile between 2000-09 and 2090-99 in relation to the current

185 depth profiles of diapausing populations.

187 **Results**

188 Relationship between potential diapause duration and prosome length and 189 overwintering temperature

190 Fig. 1 shows the modelled relationship between potential diapause duration and prosome

191 length and overwintering temperature. Field data indicates that typical temperatures

192 experienced by diapausing animals range from 0 to 5 °C (Heath *et al.*, 2004), whereas

193 prosome length typically varies from to 2.1 to 2.7 mm (Miller *et al.*, 2000; Arashkevich *et al.*,

194 2004; Tarrant *et al.*, 2008; Pepin and Head, 2009).

Diapause duration is relatively sensitive to body length. An animal of prosome length 2.7 mm 195 will have potential diapause duration 91% longer than that of an animal of prosome length 196 2.1 mm. This difference is principally driven by the fact that bigger animals have relatively 197 198 higher energy reserves than do small animals, with the allometric scaling of overwintering metabolism a lesser component. Weight-specific metabolism is 16% lower for a 2.7 mm 199 animal than for a 2.1 mm individual, which will, in itself, result in potential diapause duration 200 being 19% longer in a 2.7 mm animal. Temperature has a large effect of diapause duration 201 because of its influence on metabolism. A 1 °C increase in overwintering temperature will 202 result in a 10% reduction in potential diapause duration, while a 2 °C temperature increase 203 will reduce diapause duration by 19%. 204



Figure 1: Modelled relationship between potential diapause duration (days) and prosome 207 length and overwinter temperature. Field data was used to derive the relationship between 208 prosome length and energy reserves. Potential diapause duration is the time taken for an 209 210 animal to use up its energy reserves while maintaining sufficient reserves for post-diapause molting and gonad formation. Arrows show projected changes in the temperatures and length 211 in locations throughout the North Atlantic under IPCC RCP 8.5 as projected using the NEMO 212 ocean model. Starting and end points are the means for 2000-09 and 2090-99 respectively. 213 Letters refer to the following regions: A - Faroe-Shetland Channel, B - Iceland Basin, C -214 215 Irminger Basin, D - Labrador Sea, E - Mid-Atlantic Ridge, F - Northeast Iceland, G -Norwegian Sea, H - Rockall Basin. 216

218 Current diapause duration

Modelled potential diapause duration in the North Atlantic between 2005 and 2012 are show
in Fig. 2. Potential diapause duration shows significant geographic variation. In particular,
there is a marked difference between the overwintering centres in the Norwegian and
Labrador Seas. The Labrador Sea has potential diapause durations of between 200 and 250

days. The Norwegian Sea has potential diapause durations of between 300 and 350 days.

The main cause of this difference is overwintering temperatures. Fig. 2 shows modelled 224 225 prosome lengths and estimated temperatures experienced during winter by diapausers. Surface temperatures during summer in the Norwegian Sea are higher than in the Labrador 226 Sea, which results in smaller animals. However, the relatively lower potential diapause 227 duration induced by smaller body size is overwhelmed by the effects of overwintering 228 temperature. Overwintering temperatures in much of the Norwegian Sea are 4 °C lower than 229 in the Labrador Sea. In itself a 4 °C reduction in temperature results in a 50% increases in 230 231 diapause duration.

Modelled potential diapause durations are consistent with field knowledge of diapause 232 duration. The model shows potential diapause duration of 330 days in the central Norwegian 233 Sea, in comparison with approximately 250 days as shown by field studies (Hind et al., 2000; 234 Speirs et al., 2006; Melle et al., 2014). A range of field studies indicate that diapause duration 235 236 in the Gulf of Maine is approximately 6 months (Durbin et al., 2000; Johnson et al., 2006). 237 This matches our model, which indicates that potential diapause duration in this region is 238 approximately 220 days. Furthermore, our model implies that higher temperatures are the principal reason for C. finmarchicus having significantly lower diapause durations in the Gulf 239 240 of Maine than in Northeast Atlantic regions. Field data shows that diapause duration is 250-270 days in Disko Bay, west Greenland (Madsen et al., 2001), which compares with 241

- 242 modelled potential diapause duration of approximately 330 days. Field estimates of median
- 243 diapause duration are therefore approximately 20-30% lower than modelled potential
- 244 diapause duration.



Figure 2: Modelled potential diapause duration of *C. finmarchicus*. Potential diapause
duration was estimated assuming that body size was determined by mean surface temperature
in July and August. Overwintering temperatures were estimated by creating a synthetic map
of median diapause depths using field data, and then calculating the temperature at that depth.
The solid line demarcates the approximate southern extent of *C. finmarchicus*'s geographic
distribution. Regions with high sea-ice cover where population modelling (Speirs et al. 2006)
indicates *C. finmarchicus* is not viable were excluded.

254 **Projected changes under climate change**

and potential diapause duration between 2000-2009 and 2090-2099. Most of the North 256 257 Atlantic area studied sees significant increases in summer surface temperatures, which induce a reduction in prosome length. Prosome lengths typically reduce by 0.1-0.2 mm in the 258 Labrador Sea. Parts of the Norwegian Sea have similar reductions in prosome length, 259 however regions north and east of the Faroe Islands see only marginal changes. The most 260 pronounced changes in prosome length are in the region south of the Gulf of St. Lawrence, 261 where there are declines of 0.3-0.4 mm. 262 Changes in overwintering temperature are not uniform throughout the North Atlantic. 263 Temperature experienced by diapausing animals only changes marginally in the Norwegian 264 Sea, whereas projected increases in overwintering temperature in the Labrador Sea are 265 between 2 and 3 °C. 266 Estimated percentage changes in potential diapause duration are shown in Fig. 3. Model 267

Fig. 3 shows projected changes in prosome length of diapausers, overwintering temperature

results indicate that changes in diapause duration will vary significantly. The Norwegian Sea
witnesses marginal reductions in diapause duration, with maximum changes of less than 20%.
In fact, parts of the Norwegian Sea witness increases in diapause duration.

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Figure 3: Projected changes in mean prosome length, overwintering temperature and potential

- diapause duration of *C. finmarchicus* populations (2000-2009 to 2090-2099), under IPCC
- 275 RCP 8.5.

In contrast, potential diapause duration shortens significantly in the Labrador Sea. The central 276 Labrador Sea witnesses a shortening of diapause duration of almost 100 days, with reductions 277 of 30-40%. The region south of the Gulf of St. Lawrence and the Gulf of Maine, with 278 279 reductions of at least 50%, see the most pronounced reduction in potential diapause duration. Projected changes in summer surface and overwintering temperature and thus prosome length 280 and potential diapause duration are shown for a number of locations in Fig. 2. The relative 281 282 influence of summer surface and diapause temperature on the change in potential diapause duration varies significantly. In the Rockall Basin, Irminger Basin, Northeast Iceland and the 283 Iceland Basin the influence of surface and overwintering temperature changes is 284 285 approximately the same. In contrast, in the Labrador Sea, the Mid-Atlantic Ridge and the Faroe-Shetland Channel the influence of overwintering temperature changes is much more 286 pronounced. The Norwegian Sea is the only region where overwintering temperature 287 288 declines, with potential diapause duration staying almost unchanged this century.





Figure 4: Changes in temperature profiles in the first week of January in regions considered.

Solid lines are temperatures in 2000 and 2009 using IPCC RCP 8.5. The light grey bar shows

the depths of the 17th to 50th percentile of diapausing populations and the dark grey bar

shows the depths of the 50th to the 83rd percentile as derived from field data.

Fig. 4 shows the projected changes in the vertical temperature profile in a number of
locations in the North Atlantic and compares them with the current vertical distribution of
diapausing *C. finmarchicus* populations. Changes in the temperature profile show a
noticeable east-west pattern. Temperature changes in deep waters in the Northeast Atlantic
are marginal and have minimal impact on diapause duration. However, changes in deep water
temperatures in the overwintering centre of the Labrador Sea are pronounced.

Notably, the impacts of increased overwintering temperatures could be almost entirely offset in some regions if animals diapause in deeper waters. In the Labrador Sea, animals currently diapause at depths of between 500 and 1000 m. However, overwintering at depths of greater than 2000 m would result in minimal changes in overwintering metabolic costs. This is also true for the Mid-Atlantic Ridge, where temperatures at a depth of 2000 m in 2090-2099 are projected to be similar to those at 1000 m today, where animals currently diapause.

308

309 Discussion

310 The ecological consequences of the projected reductions in potential diapause duration in large parts of the North Atlantic will likely be significant. However, because we have a poor 311 understanding of the causes of diapause exit timing (Johnson et al., 2008) they are difficult to 312 predict. Diapause exit may be triggered by an environmental cue such as photoperiod (Hind 313 et al., 2000) or relate largely to lipid metabolism (Irigoien, 2004). In the first case, animals 314 could exit diapause before they have low lipid levels. Diapause duration will therefore only 315 316 be reduced when potential diapause duration is less than current diapause duration. However, reductions in post-diapause lipid levels will also reduce the ability of females to lay eggs in 317 the absence of food (Richardson et al., 1999) and lower post-diapause survival, which will 318 319 influence population dynamics.

320 On the other hand, if diapause duration is related purely to lipid metabolism, reductions in potential diapause duration will automatically reduce actual diapause duration. Evidence 321 suggests this is the case in the Gulf of Maine. Animals currently exit diapause at the end of 322 323 the year in this region (Durbin et al., 2000), and our model shows that this is likely triggered by low lipid levels. In our model scenario diapause exit time for populations in this region 324 will be in September and October by the end of this century. Overwinter persistence of 325 326 locally produced populations will therefore be extremely challenging, which makes understanding the role of advection of populations from colder waters into this region (Runge 327 328 et al., 2015) important. Reductions in diapause duration are therefore likely to have ecological consequences regardless of what causes animals to exit diapause. 329 One of the most notable aspects of our results is that the large reduction in diapause duration 330 in the Labrador Sea could be significantly mitigated if animals diapause in deeper waters. 331 332 Diapausing at a depth of 2000 m greater than at present will increase diapause duration by approximately 25% in the Labrador Sea at the end of the century. However, it is unclear if 333 animals can diapause in waters of this depth. All studies of overwintering populations show 334 that they never diapause at depths greater than 1500 m (Heath and Jønasdøttir, 1999; Heath 335 336 et al., 2000; Bagøien et al., 2001; Heath et al., 2004; Bonnet et al., 2006; Edvardsen et al., 337 2006; Head and Pepin, 2008; Pepin and Head, 2009). Furthermore, it may be physically implausible for animals to diapause at these depths. Visser and Jønasdøttir (1999) used the 338 biophysical properties of lipids and animal dry matter to derive the buoyancy properties of 339 diapausing animals. Their results imply that animals will be negatively buoyant at the depths 340 required to offset increased temperatures in the Labrador Sea, which may make them 341 incapable of diapausing at these depths. Furthermore, the reduction in prosome length will 342 make animals more negatively buoyant in future than they are today due to the combined 343

effects of lower relative lipid content and non-lipid body mass having higher density than
lipids (Visser and Jønasdøttir, 1999).

The importance of lipid reserves has been disputed by some researchers, who have argued 346 that they are mostly used for activities other than metabolism (Jønasdøttir, 1999; Fiksen, 347 2004; Irigoien, 2004). However, our model shows that use of lipid reserves during diapause is 348 of vital importance. Potential diapause duration is typically 50-100 days greater than 349 350 observed diapause durations in the Northeast and Northwest Atlantic (Melle et al., 2014); therefore our model implies that over half of pre-diapause lipid reserves are used to meet 351 metabolic costs. This is consistent with field studies in the Labrador Sea (Pepin and Head, 352 353 2009), Irminger Sea (Heath et al., 2008) and Malengen, Northern Norway (Pasternak et al., 2001), which showed that over half of lipid reserves are used up during diapause. 354

Lipid quality can influence diapause because of energy density variations (Kattner and 355 Hagen, 1995) and the effect of wax ester un-saturation level (Pond, 2012). There is some 356 evidence that animals in temperate regions have fewer long chain fatty acids than more 357 358 northern animals (Kattner, 1989), which may have an impact on diapause duration. However, the close resemblance of lipid composition in diapausing animals across latitudes (Kattner 359 360 and Hagen, 2009) means geographic variations in energy density will be relatively low. Liquid to solid phase transitions in copepods may influence diapause depth (Pond, 2012). The 361 temperature and depth at which these occur are related to wax ester un-saturation level, but 362 they are not likely to be of ecological significance for *C. finmarchicus* (Wilson et al., 2013). 363 However, for higher latitude species, for example Calanoides acutus (Pond and Tarling, 364 365 2011), these phase transitions may play a critical role in influencing the impact of climate change on diapause. Extending our model to other species may therefore require 366 consideration of lipid quality. 367

368 A fundamental uncertainty of our model is the reliability of experimental estimates of overwintering respiration rates, which involve taking animals from depths of up to 1000 m 369 370 and then measuring their respiration rates in a laboratory at the surface. Changes in water pressure, and the inevitable disturbance of animals, possibly cause animals to partially exit 371 diapause. However, there is a consistent pattern across *Calanus* species, with overwintering 372 metabolism typically 25% of that in surface animals (Maps et al., 2014). Our model uses the 373 374 experimental work of Saumweber and Durbin (2006), which did not measure respiration rates at temperatures below 3.6 °C. However, the model provides a close fit to respiration rates 375 376 recorded at 0 °C by Ingvarsdøttir et al. (1999) (Saumweber and Durbin, 2006).

377 The significantly higher scaling of maximum lipid levels than structural weight with prosome length used in our model is shown by all studies that report lipid levels and prosome length 378 (Miller et al., 2000; Saumweber and Durbin, 2006; Pepin et al., 2011; Bergvik et al., 2012). 379 380 Furthermore, a quantitatively similar relationship is found between median lipid levels and prosome length of diapausers (Pepin and Head, 2009). The reasons for this relationship are 381 unclear. Miller et al. (2000) suggested that this was due to body organs taking up relatively 382 less space in larger individuals, however direct evidence for this is currently lacking. Other 383 384 influences could potentially explain a large part of this trend of bigger individuals having 385 relatively more lipid. Individuals can be smaller due to higher temperatures and lower food concentrations (Hygum et al., 2000). Length relationships are also potentially biased by 386 animals who diapause later in the year being smaller due to higher temperatures. Reliable 387 estimates of lipids available for diapause would therefore benefit from advances in our 388 understanding of the biochemistry of animals, and on the development of methods to 389 determine when animals have entered diapause. 390

An important future enhancement to our model would be the ability to relate lipid reservesexplicitly to environmental conditions. We assumed a simplified relationship between

393 prosome length and lipid reserves based on field data. This data comes exclusively from the Northwest Atlantic, and may not be reflective of other regions. Laboratory and field evidence 394 shows that lipid levels in *Calanus* are strongly influenced by food quantity and food quality 395 396 (Gatten et al., 1980; Hygum et al., 2000), and life history modelling shows that the timing of prey availability may be of key importance (Varpe, 2012). Estimates of the large-scale 397 geographic variations of food quality, in particular lipid levels in phytoplankton, do not exist, 398 and a large-scale research programme would be required to attain them. However, a study on 399 the relationship between prosome length and lipid reserves and temperature and food 400 401 concentration would provide important clarifications on the relationship between environmental conditions and potential diapause duration. 402

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References

414	Arashkevich EG, Tande KS, Pasternak AF, Ellertsen B (2004) Seasonal moulting patterns
415	and the generation cycle of Calanus finmarchicus in the NE Norwegian Sea, as
416	inferred from gnathobase structures, and the size of gonads and oil sacs. Marine
417	<i>Biology</i> , 146 , 119-132.
418	Bagøien E, Kaartvedt S, Aksnes DL, Eiane K (2001) Vertical distribution and mortality of
419	overwintering Calanus. Limnology and Oceanography, 46, 1494-1510.
420	Banks HT, Gregory JM (2006) Mechanisms of ocean heat uptake in a coupled climate model
421	and the implications for tracer based predictions of ocean heat uptake. Geophysical
422	Research Letters, 33 , L07608.
423	Barange M, Merino G, Blanchard JL, et al. (2014) Impacts of climate change on marine
424	ecosystem production in societies dependent on fisheries. Nature Climate Change, 4,
425	211-216.
426	Barnard R, Batten SD, Beaugrand G, et al. (2004) Continuous Plankton Records: Plankton
427	Atlas of the North Atlantic Ocean (1958-1999). II. Biographical charts. Marine
428	Ecology Progress Series, Supplement, 11-75.
429	Beaugrand G, Kirby RR (2010) Climate, plankton and cod. Global Change Biology, 16,
430	1268-1280.
431	Bergvik M, Leiknes Ø, Altin D, Dahl KR, Olsen Y (2012) Dynamics of the lipid content and
432	biomass of Calanus finmarchicus (copepodite V) in a Norwegian fjord. Lipids, 47,
433	881-95.

434	Blanchard JL, Jennings S, Holmes R, et al. (2012) Potential consequences of climate change
435	for primary production and fish production in large marine ecosystems. Philosophical
436	transactions of the Royal Society of London. Series B, Biological sciences, 367, 2979-
437	89.
438	Bonnet D, Harris RP, Hay S, Ingvarsdøttir A, Simon O (2007) Histological changes of the
439	digestive epithelium in Calanus finmarchicus: an index for diapause? Marine Biology,
440	151 , 313-326.
441	Campbell RG, Wagner MM, Teegarden GJ, Boudreau CA, Durbin EG (2001) Growth and
442	development rates of the copepod Calanus finmarchicus reared in the laboratory.
443	Marine Ecology Progress Series, 221, 161-183.
444	Chust G, Castellani C, Licandro P, Ibaibarriaga L, Sagarminaga Y, Irigoien X (2013) Are
445	Calanus spp. shifting poleward in the North Atlantic? A habitat modelling approach.
446	ICES Journal of Marine Science, 71, 241-253.
447	Chust G, Icarus Allen J, Bopp L, et al. (2014) Biomass changes and trophic amplification of
448	plankton in a warmer ocean. Global Change Biology, 20, 2124-2139.
449	Collins M, Knutti R, Arblaster J, et al. (2013) Long-term Climate Change: Projections,
450	Commitments and Irreversibility. Climate Change 2013: The Physical Science Basis.
451	Contribution of Working Group I to the Fifth Assessment Report of the
452	Intergovernmental Panel on Climate Change, pp. 1029-1136.
453	Davis SJ, Socolow RH (2014) Commitment accounting of CO ₂ emissions. <i>Environmental</i>
454	Research Letters, 9, 084018.
455	Doney SC, Ruckelshaus M, Duffy JE, et al. (2012) Climate change impacts on marine
456	ecosystems. Annual review of marine science, 4, 11-37.
	27

457	Drijfhout S, van Oldenborgh GJ, Cimatoribus A (2012) Is a decline of AMOC causing the
458	warming hole above the North Atlantic in observed and modeled warming patterns?
459	Journal of Climate, 25 , 8373-8379.

- 460 Durbin EG, Garrahan PR, Casas MC (2000) Abundance and distribution of *Calanus*
- *finmarchicus* on the Georges Bank during 1995 and 1996. *ICES Journal of Marine Science*, **57**, 1664-1685.
- Edvardsen A, Pedersen JM, Slagstad D, Semenova T, Timonin A (2006) Distribution of
 overwintering *Calanus* in the North Norwegian Sea. *Ocean Science*, 2, 87-96.
- Falk-Petersen S, Mayzaud P, Kattner G, Sargent JR (2009) Lipids and life strategy of Arctic
 Calanus. Marine Biology Research, 5, 18-39.
- Fiksen Ø, Varpe Ø, Kaartvedt S (2004) Reply to Horizons Article 'Some ideas about the role
 of lipids in the life cycle of *Calanus finmarchicus*' Irigoien (2004): II. *Journal of Plankton Research*, 26, 980-981.
- Friedlingstein P, Andrew RM, Rogelj J, *et al.* (2014) Persistent growth of CO₂ emissions and
 implications for reaching climate targets. *Nature Geoscience*, 7, 709-715.
- Gatten RR, Sargent JR, Forsberg TEV, O'Hara SCM, Corner EDS (1980) On the nutrition
 and metabolism of zooplankton. XIV. Utilization of lipid by *Calanus helgolandicus*during maturation and reproduction. *Journal of Marine Biology Association UK*, 60,
 391-399.
- Head EJH, Harris LR, Yashayaev I (2003) Distributions of *Calanus* spp. and other
 mesozooplankton in the Labrador Sea in relation to hydrography in spring and
 summer (1995-2000). *Progress in Oceanography*, **59**, 1-30.

479	Head E, Pepin P (2008) Variations in overwintering depth distributions of Calanus
480	finmarchicus in the slope waters of the NW Atlantic continental shelf and the
481	Labrador Sea. Journal of Northwest Atlantic Fishery Science, 39, 49-69.
482	Heath MR, Boyle P, Gislason A, et al. (2004) Comparative ecology of over-wintering
483	Calanus finmarchicus in the northern North Atlantic, and implications for life-cycle
484	patterns. ICES Journal of Marine Science, 61, 698-708.
485	Heath MR, Fraser JG, Gislason A, Hay SJ, Jønasdøttir SH, Richardson K (2000) Winter
486	distribution of Calanus finmarchicus in the Northeast Atlantic. ICES Journal of
487	Marine Science, 57 , 1628-1635.
488	Heath MR, Jønasdøttir SH (1999) Distribution and abundance of overwintering Calanus
489	finmarchicus in the Faroe-Shetland Channel. Fisheries Oceanography, 8, 40-60.
490	Heath MR, Rasmussen J, Ahmed Y, et al. (2008) Spatial demography of Calanus
491	finmarchicus in the Irminger Sea. Progress in Oceanography, 76, 39-88.
492	Hind A, Gurney WSC, Heath MR, Bryant AD (2000) Overwintering strategies in Calanus
493	finmarchicus. Marine Ecology Progress Series, 193 , 95-107.
494	Hinder SL, Gravenor MB, Edwards M, et al. (2013) Multi-decadal range changes vs. thermal
495	adaptation for north east Atlantic oceanic copepods in the face of climate change.
496	Global Change Biology, 20, 140-146.
497	Hirche HJ (1996) Diapause in the marine copepod, Calanus finmarchicus - A review.

498 *Ophelia*, **44**, 129-143.

500	Hygum BH, Rey C, Hansen BW, Tande K (2000) Importance of food quantity to structural
501	growth rate and neutral lipid reserves accumulated in Calanus finmarchicus. Marine
502	<i>Biology</i> , 136 , 1057-1073.
503	Ingvarsdøttir A, Houlihan DF, Heath MR, Hay SJ (1999) Seasonal changes in respiration
504	rates of copepodite stage V Calanus finmarchicus (Gunnerus). Fisheries
505	<i>Oceanography</i> , 8 , 73-83.
506	Irigoien X (2004) Some ideas about the role of lipids in the life cycle of Calanus
507	finmarchicus. Journal of Plankton Research, 26, 259-263.
508	Johnson C, Pringle J, Chen C (2006) Transport and retention of dormant copepods in the Gulf
509	of Maine. Deep-Sea Research Part II, 53, 2520-2536.
510	Johnson CL, Checkley DM (2004) Vertical distribution of diapausing Calanus pacificus
511	(Copepoda) and implications for transport in the California undercurrent. Progress in
512	<i>Oceanography</i> , 62 , 1-13.
513	Johnson CL, Leising AW, Runge JA, Head EJH, Pepin P, Plourde S, Durbin EG (2008)
514	Characteristics of Calanus finmarchicus dormancy patterns in the Northwest Atlantic.
515	ICES Journal of Marine Science, 65 , 339–350.
516	Jønasdøttir SH (1999) Lipid content of Calanus finmarchicus during overwintering in the
517	Faroe-Shetland Channel. Fisheries Oceanography, 8, 61-72.
518	Jønasdøttir SH, Visser AW, Richardson K, Heath MR (2015) A seasonal copepod lipid pump
519	promotes carbon sequestration in the deep North Atlantic. Proceedings of the National
520	Academy of Sciences of the United States of America, 112, 12122-12126.

521	Kattner G (1989) Lipid composition of C. finmarchicus from the North Sea and the Arctic, a
522	comparative study. Comparative Biochemistry and Physiology Part B: Comparative
523	Biochemistry, 94, 185-188.
524	Kattner G, Hagen W (1995) Polar herbivorous copepods - different pathways in lipid
525	biosynthesis. ICES Journal of Marine Science, 52, 329-335.
526	Kattner G, Hagen W (2009) Latitudinal characteristics and perspective to global warming. In:
527	Arts MT, Brett MT, Kainz M (eds) Lipids in aquatic ecosystems. Springer, New
528	York, pp 257–280.
529	Li C, von Storch JS, Marotzke J (2013) Deep-ocean heat uptake and equilibrium climate
530	response. Climate Dynamics, 40, 1071-1086.
531	Locarnini RA, Mishonov AV, Antonov JI, et al. (2013) World Ocean Atlas 2013. Vol. 1:
532	Temperature. A. Mishonov, Technical Ed. NOAA Atlas NESDIS, 73, 40.
533	Madec G (2012) NEMO ocean engine. Note du Pole de mod elisation, Institut Pierre-Simon
534	Laplace (IPSL), France.
535	Madsen SD, Nielsen TG, Hansen BW (2001) Annual population development and production
536	by Calanus finmarchicus, C. glacialis and C. hyperboreus in Disko Bay, western
537	Greenland. <i>Marine Biology</i> , 139 , 75–93.
538	Maps F, Record NR, Pershing AJ (2014) A metabolic approach to dormancy in pelagic
539	copepods helps explaining inter- and intra-specific variability in life-history strategies.
540	Journal of Plankton Research, 36, 18-30.

541	Melle W, Runge J, Head E, et al. (2014) The North Atlantic Ocean as habitat for Calanus
542	finmarchicus: Environmental factors and life history traits. Progress in
543	<i>Oceanography</i> , 129 , 244-284.
544	Miller CB, Crain JA, Morgan CA (2000) Oil storage variability in Calanus finmarchicus.
545	ICES Journal of Marine Science, 57, 1786-1799.
546	Miller CB, Morgan CA, Prahl G, Sparrow MA (1998) Storage lipids of the copepod Calanus
547	finmarchicus from Georges Bank and the Gulf of Maine. Limnology and
548	<i>Oceanography</i> , 43 , 488-497.
549	Møller EF, Maar M, Jønasdøttir SH, Gissel Nielsen T, Tonnesson K (2012) The effect of
550	changes in temperature and food on the development of Calanus finmarchicus and
551	Calanus helgolandicus populations. Limnology and Oceanography, 57, 211-220.
552	Pasternak A, Arashkevich E, Tande K, Falkenhaug T (2001) Seasonal changes in feeding,
553	gonad development and lipid stores in Calanus finmarchicus and C. hyperboreus from
554	Malangen, northern Norway. Marine Biology, 138, 1141-1152.
555	Pepin P, Head EJH (2009) Seasonal and depth-dependent variations in the size and lipid
556	contents of stage 5 copepodites of Calanus finmarchicus in the waters of the
557	Newfoundland Shelf and the Labrador Sea. <i>Deep-Sea Research I</i> , 56 , 989-1002.
558	Pepin P, Parrish CC, Head EJH (2011) Late autumn condition of Calanus finmarchicus in the
559	northwestern Atlantic: evidence of size-dependent differential feeding. Marine
560	Ecology Progress Series, 423, 155-166.
561	Planque B, Fromentin JM (1996) Calanus and environment in the eastern North Atlantic. I.
562	Spatial and temporal patterns of C. finmarchicus and C. helgolandicus. Marine
563	Ecology Progress Series, 134, 101-109.

564	Pond DW, Tarling GA (2011) Phase transitions of wax esters adjust buoyancy in diapausing
565	Calanoides acutus. Limnology and Oceanography, 56, 1310-1318,
566	Pond DW (2012) The physical properties of lipids and their role in controlling the
567	distribution of zooplankton in the oceans. Journal of Plankton Research, 34, 443-453
568	Preziosi BM, Runge JA (2014) The effect of warm temperatures on hatching success of the
569	marine planktonic copepod, Calanus finmarchicus. Journal of Plankton Research, 36,
570	1381-1384.
571	Rey-Rassat C, Irigoien X, Harris R, Carlotti F (2002) Energetic cost of gonad development in
572	Calanus finmarchicus and C. helgolandicus. Marine Ecology Progress Series, 238,
573	301-306.
574	Reygondeau G, Beaugrand G (2011) Future climate-driven shifts in distribution of Calanus
575	finmarchicus. Global Change Biology, 17 , 756-766.
576	Riahi K, Rao S, Krey V, et al. (2011) RCP 8.5-A scenario of comparatively high greenhouse
577	gas emissions. Climatic Change, 109, 33-57.
578	Richardson K, Jónasdóttir SH, Hay SJ, Christoffersen A (1999) Calanus finmarchicus egg
579	production and food availability in the Faroe-Shetland Channel and northern North
580	Sea: October-March. Fisheries Oceanography, 8 (Suppl. 1), 153-162.
581	Runge JA, Plourde S, Joly P, Niehoff B, Durbin E (2006) Characteristics of egg production
582	of the planktonic copepod, Calanus finmarchicus, on Georges Bank: 1994-1999.
583	Deep-Sea Research Part II, 53, 2618-2631.

584	Runge JA, Ji R, Cameron RS, et al. (2015) Persistence of Calanus finmarchicus in the
585	western Gulf of Maine during recent extreme warming. Journal of Plankton Research,
586	37. 221-232.

587 Sanford T, Frumhoff PC, Luers A, Gulledge J (2014) The climate policy narrative for a

588 dangerously warming world. *Nature Climate Change*, **4**, 164-166.

- Saumweber WJ, Durbin EG (2006) Estimating potential diapause duration in *Calanus finmarchicus*. *Deep-Sea Research Part II*, **53**, 2597-2617.
- Schmidt-Nielsen K (1997) Animal physiology: adaptation and environment. Cambridge
 University Press.
- 593 Speirs DC, Gurney WSC, Heath MR, Horbelt W, Wood SN, de Cuevas BA (2006) Ocean-
- scale modelling of the distribution, abundance, and seasonal dynamics of the copepod *Calanus finmarchicus. Marine Ecology Progress Series*, **313**, 173-192.
- 596 Svetlichny LS, Hubareva ES, Arashkevich E (1998) Physiological and behavioural response
- to hypoxia in active and diapausing stage V copepodites of *Calanus* euxinus. *Archiv fur Hydrobiologie Special Issues Advances in Limnology*, **52**, 507-519.
- Tarrant AM, Baumgartner MF, Verslycke T, Johnson CL (2008) Differential gene expression
 in diapausing and active *Calanus finmarchicus* (Copepoda). *Marine Ecology Progress Series*, 355, 193-207.
- Varpe O (2012) Fitness and phenology: annual routines and zooplankton adaptations to
 seasonal cycles. Journal of Plankton Research, 34, 267–276.
- Visser AW, Jønasdøttir SH (1999) Lipids, buoyancy and the seasonal vertical migration of
 Calanus finmarchicus. Fisheries Oceanography, **8**, 100-106.

606	Wilson RJ, Speirs DC, Heath MR (2013) Solid evidence or fluid ideas on the importance
607	lipid phase transitions to diapausing copepods. Journal of Plankton Research, 35,
608	438–440.

Wilson RJ, Speirs DC, Heath MR (2015) On the surprising lack of differences between two
 congeneric calanoid copepod species, *Calanus finmarchicus* and *C. helgolandicus*.
 Progress in Oceanography. 134, 413-431.

- 612 Yool A, Popova EE, Coward AC, Bernie D, Anderson TR (2013) Climate change and ocean
- acidification impacts on lower trophic levels and the export of organic carbon to the

614 deep ocean. *Biogeosciences*, **10**, 5831-5854.