# **Development of Wearable Sensors for Tailored Patient Wound Care**

Stephen D. Milne, Patricia Connolly, Hanadi Al Hamad and Ihab Seoudi

Abstract— In recent years a specialist interest has developed worldwide in advanced wound management for difficult to heal chronic wounds. Further progress in advanced wound management will require an improvement in personalized medicine for the patient and in particular an improvement in the availability of diagnostic tests and parameters that fulfil clinical need in wound management decisions. However, without easy to use sensors for nurses and carers these potentially important near-patient diagnostic parameters will not enter clinical diagnostics. This study focuses on a number of metrics for wound condition and wound healing: wound fluid pH, wound moisture, and wound matrix metalloproteinases (MMP) enzyme activity. To observe these important markers state of the art sensors have been developed that are based on inexpensive sensing technologies that can be integrated within wound dressings. These sensors will enable the wound healing markers to be studied and profiled in clinics which will further enhance the understanding of these markers and their relationship in the complex healing process involved in chronic wound healing.

## I. INTRODUCTION

Chronic wounds are a growing problem worldwide with the annual spend close to £190 million on wound dressings alone by the National Health Service, in England, during 2011 [1]. In the USA chronic wounds accounted for 31.4% of all direct treatment costs for dermatological diseases [2]. Chronic wounds mostly affect the over 60's and with an aging population this problem is set to rise year on year [3]. The high costs associated with chronic wounds are due to their complex healing patterns which do not follow the standard healing path of an acute wound and result in delayed healing. Currently there is a real lack of analytical tools available that assess the real time state of the wounds. At presents clinicians rely on subjective observations to evaluate the healing of the wound and use treatments selected from their experience on a trial and error basis to achieve healing in chronic wounds. Clinical staff are thus faced with a plethora of choices in dressings and treatments with many competing manufacturers' protocols to consider during treatment selection. Each visual examination of a

Ihab Seoudi and Hanadi Al Hamad are with Hamad Medical Corporation, Doha, Qatar. (e-mail: iseoudi@hmc.org.qa).

wound to judge its stage of healing requires removal of the wound dressing. Wearable sensors to monitor the condition of a wound under the wound dressing will enable the state of the wound to be observed without disturbing the wound by removing the dressing.

The use of analytical measurement tools for measuring healing parameters in wounds is still an emerging field in wound management and treatment. Analytical measurements such as wound pH, wound size, protease level and moisture balance would enable an objective measurement of healing to provide the clinician with the most efficient and effective techniques to heal the wound.

These sensors will enable the wound healing markers of moisture, pH and MMP enzyme activity to be studied simultaneously and profiled in wound healing clinics which will further the understanding of their relationship in the complex healing process involved in chronic wound healing. The real-time measurement of these parameters is important in ensuring that the wound microenvironment is maintained in its optimum condition for healing to occur. These parameters, if measured and made available to clinicians, can provide a patient-specific, personalized, data-based approach to the selection of appropriate wound dressing and wound management protocols.

#### II. MARKERS OF WOUND CONDITION

## A. pH in wounds

When the skin is broken the internal pH of the body is exposed moving the pH from the acidic skin towards the alkali pH of the internal body. There is a large range of reported data on pH level of wounds with the range reported from 5.45-8.9 [4]-[6]. The large range of pH values is interesting as the level of pH directly and indirectly affects all of the processes that take place in the wound. The pH of a wound varies as it heals moving from an alkaline pH towards an acidic pH during re-epithelialization as the skin layer reforms [7]. Research has shown that non healing wounds and those with a high bacterial load generally have a pH level above 7.6 [8]. The trend in recent advanced wound dressings in reducing pH suggests that a lower wound pH helps start the healing process of chronic wounds. In a recent study by the authors it was found that certain wound dressings reduced the pH of a model wound bed significantly [9]. A lower wound pH would increase the wound oxygen, reduce MMP levels and reduce the proliferation of bacteria. While this trend toward lower pH may help with some aspects of the healing process the overall role of pH in a

Research supported by The Qatari national research foundation (QNRF) NPRP 4-603-3-79.

Stephen Milne and Patricia Connolly are with the Department of Biomedical Engineering, University of Strathclyde, Wolfson building, Glasgow, G4 0NW, UK (phone: (+44)1415482855; fax: (+44)141 552 6108; e-mail: s.milne@ strath.ac.uk).

wound is not fully understood and further research needs to be conducted. There is currently no data to suggest the most effective pH level for wound healing. The lack of data on this subject is somewhat related to the difficulties in measuring wounds. The standard glass pH electrode is still the only effective way to measure the pH of a wound. They are expensive and difficult to sterilize. The pH electrode has not been successfully adapted into smaller and alternate packaging because the membrane that forms the selective part of the electrode, being made of silicon glass, is fragile, thus limiting manufacture, sterilization and practical application. A tool to measure pH level in an easy, cost effective and reliable way will enable the pH of a wound to be studied further to find its overall influence on the healing process and a results from a prototype early device are presented in Section IV.

# B. Moisture of wounds

Moisture of the wound is now accepted as a key element in the condition of the wound bed [10] based upon the evidence that cell proliferation and migration is facilitated by a moist environment. International clinical practice guidelines known as the TIME guidelines now stress the importance of moisture control in the wound healing cycle [11].

Modern wound dressings provide better control and absorbance of wound exudate, however, the actual moisture status inside the dressing at the wound surface is not known. Achieving a moist wound environment relies on clinical judgment to determine the correct therapeutic levels, since too little moisture will desiccate the wound and too much will lead to maceration of the wound bed and surrounding tissue. A study by McColl et al [12] using a new artificial wound bed and in situ moisture sensors showed some surprising results as some popular wound dressings dried out the wound and others allowed liquid to pool on the wound surface. The importance of moisture balance in wound care continues to be the subject of great interest in wound healing.

# C. Protease level in wound

Chronic wound fluid contains a greatly increased level of protease activity up to 30 times that of an acute wound[13]. Matrix metalloproteinases (MMPs) are involved in every phase of wound healing [14]. In acute wound healing they act to remove necrotic tissue, promote migrations of fibroblasts and keratinocytes, regulate the activity of some growth factors, contract the wound matrix and promote angiogenesis. During the final stages of re-epithelialization they are active in remodeling the newly formed granulation tissue. As a chronic venous ulcer wound moves towards healing the activity of MMPs decreases. If the balance of the proteases is upset within a wound it can upset the cellular processes preventing healing and eventually causing further damage to the wound. It has been suggested that MMP2, MMP8 and MMP9, as they degrade type 1 fibrin collagen, have the biggest role in degrading the extracellular matrix and have been found at high levels in chronic wound fluid

[11] [15]. The measurement and study of these important wound healing markers will enable clinicians to have a clear idea of the protease activity in the wound and tailor patent treatment accordingly.

## III. METHOD OF DETECTING WOUND CONDITION

# A. pH sensor

The pH sensor presented in Fig. 1 and developed by the authors is based on inexpensive screen printing technology. The sensor is based on potentiometric method of detecting pH in liquid samples. The sensor has been developed so that it can accurately measure pH accurately with as little as  $10\mu$ L of wound fluid sample. The sensor has been used in a trial looking at how wound dressings influence the pH of the wound environment [9].

To test for practical use a sterile cotton bud swap was used to absorb fluid then pressed onto the sensor to transfer some liquid. During the test this method worked successfully and gave a stable pH reading over the pH range 4-10. The sensor hardware device was designed for simple operation by a nurse or carer to give accurate readings with small sample volume. The sensor could be modified to allow it to be placed within a wound dressing and provide real time pH conditions of the wound.



Figure 1. Hand held measurement device with sensor shown at top of device.

# B. Moisture sensor

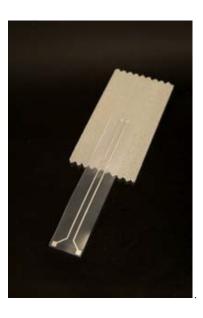


Figure 2. Moisture sensor, Ohmedics

The author group at the University of Strathclyde have now developed a sterile, disposable moisture sensor, suitable for use in any dressing worn by a patient, which allows the dressing moisture status to be checked without the need to disturb the dressing as shown in Fig. 2. This allows clinical staff to optimally time the dressing changes and dressing type and avoids pain for the patient while reducing the risk of infection. It was released on the healthcare market in 2011 and is now in clinical use in a number of centers.

The sensor was fabricated by screen printing silver chloride ink onto a biocompatible polymer. To stop the sensor from adhering to the wound during measurements a porous, nonadherent layer of material was added to the sensor to cover the AgCl measurement electrodes.

# C. Protease sensor

To move towards creating a protease sensor the authors developed an ELISA sandwich assay for measuring MMP-9 at physiologically relevant levels. Using the MMP-9 ELISA assay as a gold standard a new point of care sensor is in development to allow the user to detect the MMP level in a wound using an electrochemical based method.

## IV. SENSOR PERFORMANCE

## A. pH sensor response

The sensor is able to detect the pH with as little  $10\mu$ L of fluid in an operational range of pH 3.5-pH 10 as shown in Fig. 3. The sensor is stable to other small molecules, such as Na<sup>+</sup> and K<sup>+</sup>, and they do not interfere with the measurement of the pH.

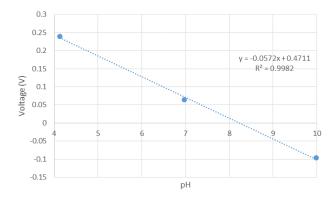


Figure 3. Response of a printed pH electrode each point was recorded with a 10uL sample size of pH buffer solution.

The pH sensor was able to effectively measure pH in a horse serum mixture with measured values only within 0.04pH of a glass electrode measurement. This result indicates that the pH electrode with screen printed Ag/AgCl reference electrode is able to accurately measure the pH of human wound fluid. The sensor has also been incorporated into a different form that is capable of being placed directly in a wound. This sensor type was used to record the real-time changes that occur after application of wound dressing to a simulated wound bed model [9]. During this research it was found that certain wound dressings could change the pH of a model wound environment to low pH values. This reduction in pH will have influence on the various different molecules and processes vital to wound healing. Further research into its effects needs to be conducted to understand pH influence in wound healing.

## B. protease assay performance

The MMP-9 ELISA was successfully produced to show linear results over a clinical relevant range (0.01-100ng/ml) of MMP-9 concentration (Fig. 4). Investigations into preservation solutions which will enable the ELISA plates coated in antibodies to be stored at long periods at room temperature have been investigated. The ELISA will act as a gold standard for MMP detection using a electrochemical method.

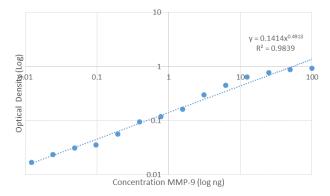


Figure 4. Results of 100ng/ml poly concentration log scale of results (n=3) averaged with average zero subtracted.

## C. Wound moisture sensor

The moisture sensor has a CE mark and is currently available for sale in the European and Middle Eastern markets with the university spin out company Ohmedics. WoundSense product is a single-use, sterile, disposable, moisture sensor which is placed on the wound at dressing change (Fig. 5). The sensor is read for moisture level using a small, handheld meter as required. Because the dressing is not disturbed the clinician, having read the moisture level, can take decisions as to the most appropriate time to change the dressing, decide on a change of dressing type or that the wound requires extra moisture if necessary [16]. The sensor has been trialed with a smartphone based telehealth system to provide clinician with information on moisture content and indicate when a wound dressing needs changed.



Figure 5. Moisture sensor, Ohmedics

# V. CONCLUSION

Each of the sensors developed gives a window to the condition of the wound bed. These wound based metrics will allow the clinician to observe the condition of the wound and apply different treatments to ensure the wound is at its optimum healing condition. The aim of the project is to use these sensors clinically to observe the effect of pH, moisture, and protease levels affect the wound healing condition. The wound moisture monitor can already be placed in situ in a wound and with further development the pH and protease sensors will follow to enable continuous sensing of these metrics of wound condition.

#### ACKNOWLEDGMENT

This research was funded by the Qatari National Research Foundation (QNRF) NPRP 4-603-3-79.

## REFERENCES

- NHS Authority Business Services, "National Wound Management Charts," NHSBSA, 2012. [Online]. Available: http://www.nhsbsa.nhs.uk/PrescriptionServices/Documents/PPDPresc ribingAnalysisCharts/Wound\_Management\_National\_Jan\_2012.pdf.
- [2] D. R. Bickers, H. W. Lim, D. Margolis, M. a Weinstock, C. Goodman, E. Faulkner, C. Gould, E. Gemmen, and T. Dall, "The burden of skin diseases: 2004 a joint project of the American Academy of Dermatology Association and the Society for Investigative Dermatology.," J. Am. Acad. Dermatol., vol. 55, no. 3, pp. 490–500, Sep. 2006.
- [3] Frost and Sullivan, "European Advanced Wound," Frost&Sullivan Manag. Mark., no. August 2003, 2003.
- [4] B. Greener, A. A. Hughes, N. P. Bannister, and J. Douglass, "Proteases and pH in chronic wounds," J Wound Care, vol. 14, no. 2, pp. 59–61, 2005.
- [5] J. Dissemond, M. Witthoff, T. C. Brauns, D. Haberer, and M. Goos, "pH values in chronic wounds. Evaluation during modern wound therapy," Hautarzt, vol. 54, no. 10, pp. 959–965, 2003.
- [6] G. Gethin, "The significance of surface pH in chronic wounds," Wounds A Compend. Clin. Res. Pract., vol. 3, no. 3, pp. 52–55, 2007.
- [7] M. Romanelli, G. Gaggio, M. Coluccia, F. Rizzello, and A. Piaggesi, "Technological advances in wound bed measurements," Wounds-a Compend. Clin. Res. Pract., vol. 14, no. 2, pp. 58–66, 2002.
- [8] G. T. Gethin, S. Cowman, and R. M. Conroy, "The impact of Manuka honey dressings on the surface pH of chronic wounds," Int Wound J, vol. 5, no. 2, pp. 185–194, 2008.
- [9] S. D. Milne and P. Connolly, "The influence of different dressings on the pH of the wound environment," J. Wound Care, vol. 23, no. 2, pp. 53–57, 2014.
- [10] J. Bryan, "Moist wound healing: a concept that changed our practice. Journal of wound care," J. Wound Care, vol. 13(6), 227, 2004.
- [11] G. Schultz, D. Mozingo, M. Romanelli, and K. Claxton, "Wound healing and TIME; new concepts and scientific applications," Wound Repair Regen., vol. 13, no. 4, pp. S1–S11, 2005.
- [12] D. McColl, B. Cartlidge, and P. Connolly, "Real-time monitoring of moisture levels in wound dressings in vitro: an experimental study.," Int. J. Surg., vol. 5, no. 5, pp. 316–22, Oct. 2007.
- [13] N. J. Trengove, M. C. Stacey, S. Macauley, N. Bennett, J. Gibson, F. Burslem, G. Murphy, and G. Schultz, "Analysis of the acute and chronic wound environments: the role of proteases and their inhibitors," Wound Repair Regen., vol. 7, no. 6, pp. 442–452, 1999.
- [14] M. Muller, C. Trocme, B. Lardy, F. Morel, S. Halimi, and P. Y. Benhamou, "Matrix metalloproteinases and diabetic foot ulcers: the ratio of MMP-1 to TIMP-1 is a predictor of wound healing.," Diabet. Med., vol. 25, no. 4, pp. 419–26, Apr. 2008.
- [15] M. J. Reiss, Y. P. Han, E. Garcia, M. Goldberg, H. Yu, and W. L. Garner, "Matrix metalloproteinase-9 delays wound healing in a murine wound model," Surgery, vol. 147, no. 2, pp. 295–302, 2010.
- [16] D. McColl and M. MacDougall, "Monitoring moisture without disturbing the wound dressing," Wounds UK, vol. 5, no. 3, pp. 2–6, 2009.