

1 405 nm light technology for the inactivation of pathogens and its potential
2 role for environmental disinfection and infection control

3

4 Michelle Maclean^{1*}, Karen McKenzie¹, John G Anderson¹, George Gettinby² & Scott J MacGregor¹

5

6 ¹ The Robertson Trust Laboratory for Electronic Sterilisation Technologies, University of
7 Strathclyde, Glasgow, Scotland UK. ² Department of Mathematics and Statistics, University of
8 Strathclyde, Glasgow, UK

9

10 * Corresponding author. Mailing address: ROLEST, Department of Electronic and Electrical
11 Engineering, University of Strathclyde, Royal College Building, 204 George Street, Glasgow,
12 Scotland, G1 1XW. Phone: +44 (0)141 548 2891. Fax: +44 (0)141 552 5398. E-mail:
13 michelle.maclean@strath.ac.uk.

14

15

16

17 Running Title: 405nm light environmental disinfection

18

19

20

21

22

23

24

25

26 **Summary**

27 **Background:** Although the germicidal properties of UV light have long been known it is only
28 comparatively recently that the antimicrobial properties of visible violet-blue 405 nm light have
29 been discovered and utilised for environmental disinfection and infection control applications.

30 **Aim:** To review the antimicrobial properties of 405 nm light and describe its application as an
31 environmental decontamination technology with particular reference to disinfection of the
32 hospital environment.

33 **Methods:** Extensive literature searches for relevant scientific papers and reports.

34 **Findings:** A large body of scientific evidence is now available that provides underpinning
35 knowledge of the 405 nm light induced photodynamic inactivation process involved in the
36 destruction of a wide range of prokaryotic and eukaryotic microbial species including resistant
37 forms such as bacterial and fungal spores. For practical application, an environmental
38 disinfection system (HINS-light EDS) has been developed and tested in hospital isolation rooms.
39 The trial results have demonstrated that this 405 nm light system can provide continuous
40 disinfection of air and exposed surfaces in occupied areas of the hospital, thereby substantially
41 enhancing standard cleaning and infection control procedures.

42 **Conclusions:** Violet-blue light, particularly 405 nm light, has significant antimicrobial properties
43 against a wide range of bacterial and fungal pathogens and, although germicidal efficacy is
44 lower than UV-light, this limitation is offset by its facility for safe continuous use in occupied
45 environments. Promising results on disinfection efficacy have been obtained in hospital trials
46 but the full impact of this technology on reduction of HAI has yet to be determined.

47

48 **Keywords:**

49 Violet-blue 405 nm light; Hospital acquired infection; Infection control; Pathogens;
50 Environment; Decontamination; Disinfection; Photodynamic inactivation; Air disinfection;
51 Surface disinfection.

52

53

54

55 **Introduction**

56 Although intensive efforts over recent years are making an impact, healthcare-associated
57 infections (HAI) still regularly occur and continue to pose a major challenge. In addition to the
58 significant morbidity and financial costs, concern over contraction of a HAI is one of the greatest
59 fears of patients being admitted to hospital.¹ Infection control procedures such as hand washing
60 are of critical importance in addressing the HAI problem, however greater awareness of the
61 hospital environment as a source of nosocomial pathogens has led to renewed focus on hospital
62 cleaning and disinfection. Whilst effective physical cleaning remains essential for infection
63 control and aesthetic reasons, there has been an upsurge of interest in the development of new
64 cleaning and decontamination technologies.^{2,3} A number of these employ novel methods of
65 delivering antimicrobial chemicals, whereas others use the antimicrobial properties of light to
66 enhance disinfection,^{4,5,6} and it is this latter approach that forms the topic of this review.

67 The most germicidal wavelengths of light fall within the ultraviolet (UV) range and UVC
68 (between 240-260 nm) irradiation has traditionally been used for disinfection, particularly for
69 air and medical device decontamination applications.^{7,8,9} More recently the antimicrobial
70 properties of violet-blue visible light has emerged as an area of increasing research interest.
71 Although less germicidal than UVC light, violet-blue light with wavelengths in the region of
72 405 nm, has proved effective for inactivation of a range of microbial species, and exploitation of
73 these wavelengths may provide alternative methods of antimicrobial treatment for infection
74 control applications. This paper will provide a brief background on the use of light for
75 environmental decontamination applications within hospitals before presenting a detailed
76 description of the broad spectrum antimicrobial effects of violet-blue light and how this
77 knowledge has led to the development and clinical evaluation of a 405 nm light environmental
78 disinfection system. In addition to environmental decontamination applications, other potential
79 uses of violet-blue light for infection control proposes such as skin and wound treatment have
80 been highlighted in recent literature but these topics are out with the scope of the current
81 review.¹⁰⁻¹⁷

82 **Inactivation of microorganisms by light in the hospital environment**

83 Records of observations on the antibacterial effects of light go back to the latter part of the 19th
84 century and these early historical observations have been documented by Kowalski.¹⁸ The
85 germicidal effects of light received further attention during the early part of the 20th century and
86 the appreciation of the decontamination effect of light was translated into early hospital design
87 features where natural ventilation and exposure to sunlight were regarded as beneficial.¹⁹ The
88 roles of sunlight and natural ventilation for controlling the transmission of infections within

89 healthcare settings has recently been reviewed by Hobday and Dancer, who provide a detailed
90 record of the early – mid 20th century observations on the effects of natural sunlight on a wide
91 range of nosocomial pathogens.²⁰ Whilst natural light and ventilation were originally
92 considered beneficial, modern hospital design has tended to reduce these features. Recent
93 interest in the application of ‘artificial’ lighting within hospitals has been with regard to energy
94 reduction issues but also how lighting can affect the mood and circadian rhythm of patients.^{21,22}
95 Light from artificial sources with wavelength emission in the UV range, can have significant
96 antimicrobial effects and new technologies for hospital decontamination have been developed
97 around this concept.^{6,23-25}

98 The most widespread applications of ultra-violet germicidal irradiation (UVGI) has been for air
99 and water disinfection, as well as for decontamination of devices.²⁶⁻²⁸ More recently, with the
100 increased emphasis that has been directed towards enhanced decontamination of the hospital
101 environment, novel technologies have been developed for the rapid delivery of UVC radiation to
102 exposed surfaces in clinical areas. Several of these are automated or manually positioned
103 robotic systems using either continuous or pulsed UV emission sources.^{6,25} Detailed information
104 on UVGI and other ‘no-touch’ automated room disinfection systems is provided in a recent
105 review by Otter *et al.*⁶

106 **Antimicrobial Effects of Violet-Blue Light**

107 Until relatively recently light within the visible spectrum (400–700 nm) was considered to have
108 little biocidal effect compared to UVC light due to the lower photon energy of these wavelengths.
109 Wavelengths of violet-blue light, particularly around 405 nm, have however been shown to
110 possess antimicrobial capabilities, and there is scope for exploiting these wavelengths for the
111 control of problematic microorganisms in many areas of application including the disinfection
112 of air and exposed surfaces in the clinical environment. The following section provides an
113 overview of the antimicrobial inactivation mechanism, and the antimicrobial efficacy of high-
114 intensity 405 nm violet-blue light.

115 ***Violet-Blue Light Inactivation Mechanism***

116 Investigations into the mechanism of action of 405 nm violet-blue light indicate that
117 photodynamic inactivation occurs as a result of the photo-excitation of intracellular porphyrin
118 molecules within the exposed bacterial cells. Laboratory studies have shown that a range of
119 violet-blue light wavelengths in the region 400-425 nm can be used for bacterial inactivation,²⁹⁻
120 ³⁴ however, optimal antimicrobial activity has been found at 405 nm.^{34,35} This peak in activity
121 correlates with the absorption maximum of porphyrin molecules, termed the soret band, being

122 in this wavelength region.³⁶ Exposure to light of this wavelength induces an oxygen dependent
123 photo-excitation reaction within exposed microorganisms, where excited porphyrins react with
124 oxygen or cell components to produce reactive oxygen species (ROS) causing oxidative damage
125 and microbial cell death.^{29,37-41} Cell death has been accredited to oxidative damage to the cell
126 membrane, with a recent study demonstrating disruption of the cytoplasmic content and cell
127 walls of exposed *S. aureus*,¹⁰ and it is likely that, due to the non-selective nature of ROS, multi-
128 target damage will be induced in the microbial cells.

129 ***Antimicrobial Effects of Violet-Blue Light***

130 Extensive laboratory studies have shown that 405 nm light, and the wider violet-blue light
131 wavelengths, have a broad spectrum of activity, with successful inactivation demonstrated for a
132 wide range of organisms, including antibiotic-resistant bacterial strains such as methicillin-
133 resistant *Staphylococcus aureus* (MRSA).³⁰⁻³² Bacterial species which have demonstrated
134 susceptibility include HAI-associated organisms, including *Staphylococcus aureus*, *Clostridium*
135 *difficile*, *Acinetobacter baumannii*, *Escherichia coli*, *Staphylococcus epidermidis*, *Pseudomonas*
136 *aeruginosa*, *Klebsiella pneumoniae*, *Streptococcus pyogenes* and *Mycobacterium* species.^{29-33,42,43}
137 Bacterial sensitivity to violet-blue light inactivation tends to be species dependent, however the
138 general trend suggests that Gram positive bacteria tend to be more susceptible to inactivation
139 than Gram negative species.^{32,44}

140 Two of the most significant pathogens associated with HAI are MRSA and *C. difficile*, and
141 vegetative cells of these species both show susceptibility to violet-blue light inactivation.
142 Vegetative cells of *C. difficile* are particularly sensitive to inactivation, and this is likely to be due
143 to this organism being an obligate anaerobe, giving it increased sensitivity to oxidative
144 damage.³³ *C. difficile* spores are a significant issue for infection control, particularly due to their
145 prolonged survival in the environment, and their resilience to disinfection technologies is well
146 documented.⁴⁵⁻⁴⁷ *C. difficile* spores can be successfully inactivated by exposure to 405 nm light,
147 however as expected, significantly higher doses (~50 times) are required for inactivation
148 compared to vegetative cells.³³

149 Laboratory studies have demonstrated the successful antimicrobial efficacy of violet-blue light
150 for the inactivation of bacterial contamination in liquid,^{10,11,29,32,34} artificially seeded on
151 surfaces,^{30,31,42,48} and most recently, in biofilms.⁴⁴ Within the clinical environment, biofilm
152 formation is a major cross-contamination risk, with the presence of patient fluids such as saliva,
153 blood and urine influencing biofilm adhesion and development on surfaces.⁴⁹ Indeed, a recent
154 study attributed the presence of *Pseudomonas aeruginosa* biofilms on sinks to the acquisition of
155 infections, with a 33% death rate.⁵⁰

156 Although the germicidal efficacy of blue light is lower than that of ultraviolet light – UV
157 inactivation typically required doses of the order of mili-joules rather than joules as is the case
158 with violet-blue light^{51,52} – significant bacterial inactivation can still be demonstrated, with up
159 to 9-log₁₀ orders of reduction being achieved in one study.³² A major advantage of violet-blue
160 light inactivation is that the susceptibility of strains isolated from the clinical environment is
161 similar to their laboratory type strain counterparts i.e. clinical isolates do not show enhanced
162 resistance and thus can be inactivated by 405 nm light with no inherent problems.³² Also, it has
163 recently been demonstrated that sublethally damaged bacterial cells are more susceptible to
164 light inactivation⁴⁸, therefore, there is great potential for bacterial contamination that has been
165 sub-lethally stressed by desiccation and disinfectants during routine cleaning of the hospital
166 environment to be more susceptible to inactivation by exposure to violet-blue light.

167 In addition to clinically relevant bacteria, the effectiveness of 405 nm light for microbial
168 inactivation has also been demonstrated against bacterial species associated with foodborne
169 infection including *Listeria*, *Campylobacter*, *Shigella* and *Salmonella* species;^{32,34,53} pathogens
170 *Helicobacter pylori*, *Chlamydia* and *Propionibacterium acnes*;^{29,37,43} oral periodontal
171 pathogens;^{54,55} and fungal organisms including moulds and yeasts such as *Candida*.⁵⁶ To date
172 the effect of violet-blue light on viruses has not been fully determined, however it is expected
173 that due to the hypothesised involvement of porphyrins in the inactivation mechanism, it is
174 unlikely that viruses will be highly susceptible to light exposure alone, and may require the
175 addition of photosensitising material to enhance viricidal activity.⁵⁷

176 **Use of 405 nm Violet-Blue Light for Hospital Disinfection**

177 The wide antimicrobial spectrum of activity combined with the ability to apply light intensities
178 safe for human exposure make violet-blue light ideal for decontamination of occupied
179 environments, and the development of a system which utilises high-intensity narrow spectrum
180 (HINS) 405 nm light for environmental disinfection of the clinical environment has been
181 recently described.⁵⁸⁻⁶⁰ This new disinfection technology, termed the HINS-light Environmental
182 Decontamination System (EDS) is a ceiling-mounted lighting system designed for the reduction
183 of environmental contamination in hospital wards and other areas of the healthcare
184 environment. The antimicrobial light from the system is generated from a matrix of light-
185 emitting diodes (LEDs) which emit low irradiance violet-blue light with a narrow spectral
186 profile centred on 405 nm.⁵⁸ The output of the antimicrobial light has been set to ensure, with
187 reference to international guidelines,^{61,62} that the light source does not pose a blue light hazard
188 and is safe for use in occupied environments. Whilst biocidal, the 405 nm wavelengths is well
189 below the blue light wavelengths which can impact on human health, particularly in the region

190 of 440 nm which is associated with photoretinitis, and 480 nm which influences mood and
191 circadian rhythm in humans, as shown in Figure 1. Whilst satisfying safety standards as an
192 installed light source it is interesting to also note that, when comparing the susceptibility of
193 mammalian cells and bacteria to 405 nm light, mammalian keratinocytes and osteoblasts were
194 considerably more resistant and could be exposed to bactericidal levels of 405 nm light with no
195 loss of cell viability.^{10,11,63} The increased resistance of mammalian cells is likely due to the fact
196 that these cells have much more advanced mechanisms for coping with oxidative damage
197 compared to the more primitive microbial cells.

198 For practical application as an overhead light source, incorporation of white LEDs into the HINS-
199 light EDS system ensures the illumination output is predominantly white, thus blending with
200 the standard room lighting.⁵⁸ The system is designed to be operated continuously, providing
201 on-going disinfection of the air and all exposed environmental surfaces within the treated area,
202 with no disruption to day-to-day hospital procedures or patient care. Laboratory testing of the
203 system confirms the efficacy for inactivation of a range of bacterial pathogens associated with
204 HAI.⁶⁴ As mentioned, the low irradiance levels employed by the system were deliberately
205 selected to enable continuous disinfection in occupied environments, and therefore require
206 sufficient time to exert the antimicrobial effect. Significant inactivation of microbial
207 contamination on simulated laboratory surfaces can be achieved by approximately 1-2 h light
208 exposure,⁶⁴ however, inactivation kinetics are likely to be significantly enhanced in the 'real'
209 clinical environment due to the stressed and desiccated state of the microorganisms.⁴⁸

210 ***Clinical Assessment of 405nm Light for Environmental Disinfection***

211 A number of published studies have presented results from clinical assessment of this 405 nm
212 light system for continuous environmental decontamination of single-bed isolation rooms.⁵⁸⁻⁶⁰
213 Evaluation of the technology has been carried out in isolation rooms within two main clinical
214 areas: a Burns Unit and an Intensive Care Unit (ICU).

215 For evaluation, systems were installed within isolation rooms, and used as a complementary
216 disinfection procedure, being operated continuously during daylight hours in occupied rooms,
217 under conditions where normal clinical care and infection control measures were implemented.
218 The effect of the system was assessed through contact-plate sampling of bacterial levels on a
219 range of frequently touched contact surfaces (e.g. locker top, bed table, bed rails, bin lids, light
220 switches & door handles) which are commonly associated with being 'high-risk' surfaces for
221 cross-transmission of HAIs, as well as surfaces likely to have high contamination levels due to
222 aerial deposition, such as ledges. Samples were typically collected (i) before use, (ii) during use,
223 and (iii) a period after the HINS-light EDS units had been switched off, with the same contact

224 surfaces sampled throughout each study. Bacterial levels were assessed using 55 mm contact
225 agar plates, with a surface area of 23.76 cm², which were inoculated by pressing the agar
226 surface onto the environmental surface. Studies monitored the levels of staphylococcal bacteria
227 (a good indicator of contamination of human origin),⁵⁸⁻⁶⁰ and the total viable bacteria levels⁶⁰ in
228 order to establish the effect of the system for reducing levels of bacterial contamination around
229 the isolation room. For collection of staphylococcal organisms, Baird Parker with egg yolk
230 telurite agar (BPA), a selective medium for the growth of staphylococcal-type organisms,
231 contact plates were used. Tryptone soya agar contact plates (TSA), which use non-selective
232 growth medium, were used to obtain total viable bacterial counts (TVC). Microbiological
233 assessment, as colony forming unit (CFU) counts, was based upon growth on the contact agar
234 plates after incubation at 37°C for 24 hours (TSA plates) or 48-hours (BPA plates).

235 A number of studies also characterised the staphylococcal isolates by subculturing selected
236 isolates and then testing using Staphaurex Plus (Remel Europe Ltd, Dartford, UK) and PBP2
237 Latex Agglutination Test (Oxoid Ltd), to identify *S. aureus* and methicillin *S. aureus* isolates,
238 respectively.

239 ***Inpatient Studies***

240 An initial study evaluated use of the system for disinfection of an unoccupied isolation room,
241 and results demonstrated a significant 90% reduction (P=0.000) in the staphylococcal
242 contamination on surfaces around the room after 24-hour use.⁵⁸ Studies in burns isolation
243 rooms occupied by MRSA positive patients, with treatment periods ranging from 2-7 days,
244 demonstrated that staphylococcal contamination on surfaces around the rooms were
245 significantly reduced by 56 to 86%, over and above the reductions achieved by cleaning alone.
246 Levels of presumptive *S. aureus* and MRSA showed similar reductions.⁵⁸ Significantly, once use
247 of the system ceased, recontamination of the room was observed, to levels similar to pre-
248 treatment contamination levels.

249 An example of the data from one published study is shown in Figure 2, which demonstrates the
250 mean reductions in the total staphylococcal counts and the presumptive *S. aureus* levels in an
251 occupied burns unit isolation room, before, during and after 5-day use of HINS-light EDS.
252 Samples (n=70) were collected twice during each of the three phases, and the results from all
253 sampled surfaces have been pooled to demonstrate the overall decontamination effect the
254 system had across the room. In this study, data demonstrated that a significant 62% decrease in
255 total staphylococcal counts, and 50% decrease in presumptive *S. aureus* was achieved (P<0.05)
256 after 5-days use of the system. 'After use' samples, collected during a 6-day period after the
257 system had been turned off, showed that contamination around the room had significantly

258 risen, with a 126% and 98% increase in the total staphylococci and presumptive *S. aureus*
259 counts, respectively ($P < 0.05$), thus reinforcing the recontamination effect that occurs after
260 removal of the light-treatment.⁵⁸ Extended use of the system also proved to further reduce the
261 bacterial contamination around the room, supporting the continuous use of this system for
262 maintaining low contamination levels around isolation rooms.⁵⁸ Importantly, studies were
263 performed to show that the decontamination effect was not patient or room dependent.⁵⁹

264 Studies carried out in an ICU isolation room also demonstrated system efficacy, with 60 to 70%
265 reductions in both the staphylococcal and the total bacterial contamination across the entire
266 sampled room environment.⁶⁰ In addition to demonstrating an overall reduction in
267 contamination around the room, results demonstrated that exposed surfaces had reduced
268 contamination levels as a result of use of the system, and an example of this is shown in Figure
269 3. Levels of bacteria on various surfaces around an occupied ICU isolation room were
270 determined before use of the HINS-light EDS, and resampled after a 5-day exposure period.
271 Results demonstrated that despite marked variation in the initial bacterial bioburden there was
272 a marked decrease in levels of bacterial contamination at all tested sites.

273 In addition to these findings, a significant factor noted in the studies carried out in the ICU
274 isolation room was that despite asymmetrical positioning of the EDS units within the room,
275 results demonstrated that the special distribution of bacterial contamination was reduced
276 almost uniformly across all the sampled contact surfaces. This suggested that disinfection of
277 airborne bacteria contributes to the reductions in bacterial contamination levels, and the
278 installation positions of the systems may not be critical.⁶⁰

279 ***Outpatient studies***

280 In addition to its use for disinfection of occupied inpatient isolation rooms, the HINS-light EDS
281 has also proved effective when used in an outpatient clinic.⁵⁹ Communal use of outpatient clinic
282 rooms provides a recognised risk of cross-contamination between subsequently treated
283 patients, therefore it is important to maintain cleanliness in these areas throughout the day.
284 Studies carried were carried out to evaluate the environmental bacterial levels at the start and
285 end of 8-hour clinic sessions, with and without use of the EDS. Results demonstrated that a
286 statistically significant 61% efficacy was achieved ($P = 0.02$), and these successful results lead to
287 the suggestion that use of this system would be beneficial in other similar communal patient
288 rooms such as the bathroom or physiotherapy room, where decontamination of all surfaces is
289 unachievable between each patient due to time limitations.⁵⁹

290 Overall, results have been successful, showing evidence that use of 405 nm light achieves
291 significant reductions in bacterial contamination levels around isolation room environments.⁵⁸⁻
292 ⁶⁰ Results also demonstrated that when switched off, the decontamination effect ceases and
293 bacterial contamination levels return to around pre-treatment levels, further confirming the
294 effectiveness of the 405 nm light. It is important to note that these results were achieved under
295 a range of clinical conditions within a busy city hospital environment, and that the bacterial
296 disinfection results obtained were over and above those achieved by the hospital's normal
297 stringent infection control procedures which remained fully in place throughout the study.⁵⁸⁻⁶⁰
298 Further studies are still required to establish the effectiveness of 405 nm light for disinfection of
299 larger communal environments.

300

301 **Comparison of 405 nm Light with Other Environmental Decontamination Systems**

302 The increased awareness of the importance of the hospital environment as a source of
303 nosocomial pathogens has focused attention not only on improving the efficiency of
304 conventional cleaning and disinfection procedures, but has led to the development of a range of
305 novel technologies for enhanced decontamination of whole room environments, including new
306 UV systems (as discussed earlier), steam cleaning, hydrogen peroxide vapour and super-
307 oxidised water fogging.^{7,65-67} Although these systems are effective for widespread disinfection of
308 the room environment, they require, for safety reasons, experienced operator supervision and
309 their use is restricted to unoccupied, sealed rooms, thereby resulting in rooms being out-of-
310 commission for periods of time – a consequence which can be costly and undesirable in busy
311 ward areas. Additionally, whilst these systems provide effective decontamination, studies have
312 found that once treatment has finished, there is rapid and widespread recontamination of the
313 room.⁶⁸ In addition to human safety considerations, another problem associated with UV-light
314 and chemically-based technologies is the potential for long-term material degradation of
315 furniture and equipment within the treated room if these are repeatedly exposed.^{69,70} Therefore
316 these methods are best-suited for terminal- and deep-cleaning procedures, but are ineffective
317 for maintaining low levels of contamination.

318 Whilst UV irradiation and 405 nm light technology share some similar features they are, in
319 many respects quite distinct technologies both in their modes of action and methods of
320 application (Table I and Fig. 1). Whilst UV light is strongly germicidal it is dangerous to humans
321 and the different UV waveband regions corresponding to UVC, UVB and UVA can cause a wide
322 range of detrimental effects on the human eye and skin.⁷⁰ Violet-blue within the visible
323 spectrum can also cause harmful effects at high irradiance levels but these are particularly at

324 440 nm which can cause photoretinitis,^{61,62} and 480 nm which is the peak sensitivity of
325 mammalian photosensitive retinal ganglion cells (pRGCs) which modulate diverse physiological
326 responses to light, including circadian physiology and pupil constriction.⁷⁶ A comparison of the
327 biological effects of radiation extending from the UV and into the visible light regions is
328 presented in Figure 1. Whilst 405 nm light is germicidal it falls within a relatively benign
329 wavelength region and if operated at appropriate irradiance levels it is safe for human
330 exposure.^{61,62}

331 The above features explain why the 405 nm light environmental disinfection technology, in
332 comparison with other whole-room decontamination systems including UV technology, can be
333 operated continuously in the presence of patients and staff, thus facilitating a background
334 decontamination effect which maintains low levels of contamination.⁵⁸⁻⁶⁰ Continuous operation
335 of the 405 nm light system ensures that there is a level of disinfection concurrently being
336 applied even during periods of high activity, such as visiting hours, and bed and bandage
337 changing.^{77,78} Whilst disinfectant cleaning and hand hygiene are critical for maintaining a clean
338 environment and minimising the spread of potential pathogens, compliance with hand-washing
339 tends to be low after direct contact with a patient, and significantly, healthcare workers are
340 even less likely to wash their hands after being in contact with the environmental surfaces
341 around the patients, even though these surfaces can be reservoirs of potential pathogens.⁷⁹ Use
342 of the 405 nm light technology can strategically augment this by enhancing the low levels of
343 contamination achieved with intermittent cleaning, and also provide decontamination of
344 surfaces within rooms, such as walls and high ledges, as well as delicate equipment, which may
345 not be routinely cleaned using disinfectants. Moreover the system can be automatically
346 operated with no user training required, and consequently problems with staff and patient
347 compliance do not apply.⁵⁸⁻⁶⁰

348 As with all methods of cleaning and disinfection there are inherent disadvantages with any
349 procedure. A limitation of the 405 nm light technology is that, to ensure that patient friendly
350 room illumination conditions are used, relatively low irradiance levels are applied and this
351 impacts on microbial inactivation rates which are inevitably lower than can be achieved with
352 other decontamination technologies albeit only in short term comparisons. The high doses of
353 405 nm light required for inactivation of endospores means it is unlikely that 405 nm light alone
354 could be realistically applicable for the specific environmental decontamination of *C. difficile*
355 spores, however enhancement of the inactivation may be achieved when combined with other
356 decontamination methods such as oxidative biocides, due to the similar oxidative damage that is
357 exerted on the bacteria by both treatments.³³ In addition to the resilience of spores, the
358 antiviral efficacy of violet-blue light has not been fully established, therefore further research in

359 this area is required. Also, similar to UVC technology, 405 nm light effectively treats hospital air,
360 but only surfaces that are directly or reflectively exposed to the light are treated, and the effects
361 on occluded or darkly shadowed areas are limited. It is also the case that whilst all of the new
362 technologies including 405 nm light can claim to have demonstrated enhanced disinfection of
363 the hospital environment translation of this potential benefit into a significant reduction in
364 infection rates will be required to ensure the widespread uptake of these new disinfection
365 technologies.

366 **Further commentary regarding the application of 405 nm light for hospital** 367 **disinfection**

368 Regarding the deployment of the HINS-light system within hospitals, although important issues
369 such as disinfection efficacy and patient safety have been addressed, other questions relating to
370 the use of such a novel light source in clinical settings must also be considered. Undoubtedly
371 enrichment of room lighting with additional violet-blue light will alter the normal lighting effect.
372 This could have some impact on patient and staff comfort levels, and possible effects on medical
373 procedures that involve colour perception must also be considered. In the hospital trials
374 already conducted with the HINS-light EDS no such issues have been problematic (unpublished
375 observations) but monitoring for such effects must remain during uptake of this technology.
376 Further hospital-based studies, funded by the Scottish Infection Research Network and the Chief
377 Scientist Office, are currently being initiated to investigate the acceptability of the technology,
378 and to ensure the technology is optimised with staff and patient comfort fully taken into
379 account. There may conceivably also be implications for colours employed in hospital
380 furnishings and fabrics as these may serve to amplify or suppress the reflection or absorption of
381 violet-blue light.

382 As already discussed, a benefit of 405 nm light over UV-light for disinfection purposes is that,
383 unlike UV-light, 405 nm light, because of its lower photon energy, does not cause photo-
384 degradation of photosensitive materials such as rubbers and plastics used in the hospital
385 environment and equipment.⁶⁹ However strong visible light can cause photochemical changes
386 in light-sensitive solutions, and this aspect requires consideration if such solutions were to be
387 exposed for long periods. At the relatively low 405 nm light intensities used⁵⁸⁻⁶⁰, and
388 considering the fact that light intensity reduces upon transmission through materials e.g. plastic
389 tubing or IV bag material, then this issue is not anticipated to be problematic but nevertheless
390 must remain a consideration if highly light-sensitive pharmaceuticals were introduced.

391 The HINS-light system utilises LED-based technology and as such it benefits from the well-
392 established characteristics of LED lighting, namely reduced energy requirements, long

393 operational (lifetime) use, and low maintenance characteristics. In the hospital trials already
394 conducted, the HINS-light EDS unit is designed to be easily retrofitted into the ceiling in place of
395 a ceiling tile. Installed units have remained maintenance-free and fully-operational over the
396 trial period which now extends to several years. From a lighting technology perspective, it is
397 interesting that the introduction of this LED-based disinfection system is concurrent with major
398 potential changes taking place in general lighting technology. Considerable debate is underway
399 regarding the advantages and disadvantages of replacing conventional fluorescent lighting with
400 LED sources, a discussion that is mainly being driven by potential energy efficiency gains
401 associated with LED lighting. Another potential advantage of LED technology is the capacity to
402 blend different colours to 'fine tune' the colour spectrum to suit different environments and
403 applications. In this context it is interesting that it is now appreciated, and as previously
404 discussed in this review, that the nature of the light spectrum can affect circadian rhythmicity,
405 sleep and mood and that this is associated with photosensitive retinal ganglion cells in the eye.⁷⁶
406 Such effects are not only important in the home and workplace but also for patients in the
407 hospital environment, where it has been suggested that more research is required to better
408 understand how lighting in the hospital environment can influence sleep, mood and pain in
409 medical inpatients.²² Future development of the HINS-light EDS system will undoubtedly be
410 influenced by the various considerations outlined above.

411 **Conclusions**

412 Although the germicidal effects of sunlight and UV-light have been known for well over a
413 century it is only comparatively recently that the antimicrobial properties of visible light in the
414 violet-blue region of the spectrum have been recognised and studied in a number of
415 laboratories. Given the severity of current and anticipated future microbiological problems
416 faced by society, the development of any new antimicrobial weapon is to be welcomed. Violet-
417 blue light, with particular efficacy at 405 nm, has been shown to possess broad spectrum
418 photodynamic antimicrobial activity, and as such its use has been suggested for a range of
419 potential clinical and medical applications.

420 One such application is the use of 405 nm light for environmental disinfection. The increased
421 safety of 405 nm light wavelengths compared to UV-light, has facilitated development of this
422 light technology for safe continuous disinfection of occupied environments, and results have
423 shown the successful application of this system for environmental disinfection of hospital
424 isolation rooms. This technology termed, the HINS-light EDS, has demonstrated a significant
425 capability for reducing environmental bacterial contamination in clinical patient areas, over and
426 above reductions achieved using the conventional cleaning and infection control strategies

427 alone. In common with the aspirations of other novel whole room disinfection systems, it is
428 intended that this intervention technology, when used in conjunction with conventional
429 infection control procedures, can help reduce levels of pathogens in the environment, thereby
430 limiting the likelihood of pathogen transmission from the environment to patients, and thus
431 contribute to reducing levels of HAIs.

432 Whilst violet-blue 405 nm light irradiation represents a new antimicrobial approach, the
433 physical nature of this light source and the limitations of its antimicrobial effects must be
434 understood. Inevitably microbial inactivation rates using 405 nm light are slower than can be
435 achieved with the typical application of many other physical and chemical disinfection and
436 sterilisation treatments. This limitation is however mitigated by its operational facility for
437 continuous application to disinfect air and all illuminated surfaces in occupied environments
438 and by the biochemical mechanism of 405 nm light inactivation. The photodynamic inactivation
439 process induced by 405 nm light exposure involves a multi-targeted intracellular killing effect
440 resulting from the generation of reactive oxygen species, a killing mechanism that is not
441 conducive to microbial resistance development. Given these unique features, it is evident that
442 405 nm violet-blue light technology represents a novel antimicrobial approach that hopefully
443 can make some contribution to tackling the challenge posed by ubiquitous environmental
444 contamination, and to the ongoing health and resource problems associated with healthcare-
445 associated infections (HAI).

446

447 **Conflict of Interest Statement**

448 The intellectual property rights of the HINS-light EDS belong to the University of Strathclyde.
449 The University has made all systems for research purposes only and no commercial company
450 manufactures this technology.

451

452 **References**

- 453 1. Stone PW. Economic burden of healthcare associated infections: an American
454 perspective. *Expert Rev Pharmacoecon Outcomes Res* 2009; **9**: 417-422
- 455 2. Rutala WA, Weber DJ. Are room decontamination rooms needed to prevent transmission
456 of environmental pathogens. *Infect Control Hosp Epidemiol* 2011; **32**: 743-747

- 457 3. Dancer SJ. Hospital cleaning in the 21st century. *Eur J Microbiol Infect Dis* 2011; **12**:
458 1473-1481
- 459 4. Sharma M, Hudson JB. Ozone gas is an effective and practical antibacterial agent. *Am J*
460 *Infect Control* 2008; **36**: 559-563
- 461 5. Merandzic MN, Cadnum JL, Eckart KE, Donskey CJ. Evaluation of a handheld far-ultra
462 violet radiation device for decontamination of *Clostridium difficile* and other healthcare-
463 associated pathogens. *BMC Infect Dis* 2012; **12**: 120-125
- 464 6. Otter JA, Yezli S, Perl TM, Barbut F, French GL. The role of 'no-touch' automated room
465 disinfection systems in infection prevention control. *J Hosp Infect* 2013; **83**: 1-13
- 466 7. Andersen BM, DrScient HB, Boe E, BcEcon, Bjordal O, Drangsholt F. Comparison of UV-C
467 lights and chemicals for disinfection of surfaces in hospital isolation units. *Infect Control*
468 *Hosp Epidemiol* 2006; **27**: 729-734
- 469 8. Nardell EA, Bucher SJ, Brickner PW, *et al.* Safety of upper room ultra violet germicidal air
470 disinfection for room occupants: Results from the Tuberculosis ultra violet shelter
471 study. *Public Health Rep* 2008; **123**: 52-60
- 472 9. Reed NG. The history of ultra violet germicidal irradiation for air disinfection. *Public*
473 *Health Rep* 2010; **125**: 15-27
- 474 10. Dai T, Gupta A, Huang YY, *et al.* Blue light eliminates community acquired methicillin
475 resistant *Staphylococcus aureus* in infected mouse skin abrasions. *Photomed Laser Surg*
476 2013; **31**: 531-538
- 477 11. Dai T, Gupta A, Huang YY, Yin R, Murray CK, Vrahas MS, Sherwood ME, Tegos GP,
478 Hamblin MR. Blue light rescues mice from potentially fatal *Pseudomonas aeruginosa*
479 burn infection: efficacy, safety and mechanism of action. *Antimicrob Agents Chemother*
480 2013; **57**: 1238-1245
- 481 12. Elman M, Slatkine M, Harth Y. The effective treatment of acne vulgaris by a high intensity
482 narrow band 405-420nm light source. *J Cosmet Laser Ther* 2003; **5**: 111-116
- 483 13. Shalita AR, Harth Y, Elman M, *et al.* Acne phototherapy using UV free high intensity
484 narrow band blue light: a three centre clinical study. *Progress Biomed Optics Imaging*
485 2001; **2**: 61-73

- 486 14. Kleinpenning MM, Otero ME, van Erp PEJ, Gerritsen R, van de Kerkhof PCM. Efficacy of
487 blue light versus red light in the treatment of psoriasis: a double blind, randomized
488 comparative study. *J Eur Acadamy Dermatol Venereol* 2012; **26**: 219-225
- 489 15. Ganz RA, Viveiros J, Ahmad A, Ahmadi A, Khalil A, Tolckoff MJ, Nishioka NS, Hamblin MR.
490 *Helicobacter pylori* in patients can be killed by visible light. *Lasers Surg Med* 2005; **36**:
491 260-265
- 492 16. Lembo AJ, Ganz RA, Sheth S, *et al.* Treatment of *Helicobacter pylori* infection with intra
493 gastric violet light phototherapy: A pilot clinical trial. *Lasers Surg Med* 2009; **41**: 337-
494 344
- 495 17. McDonald R, MacGregor SJ, Anderson JG, Maclean M, Grant MH. Effect of 405-nm high-
496 intensity narrow-spectrum light on fibroblast populated collagen lattices – an in vitro
497 model of wound healing. *J Biomed Optics* 2011; **16**: 048003
- 498 18. Kowalski W. Ultraviolet Germicidal Irradiation Handbook UVGI for Air and Surface
499 Disinfection. Springer Heidelberg Dordrecht, London, New York 2009
- 500 19. Hobday RA. Sunlight therapy and solar architecture. *J Med Hist* 1997; **4**: 455-472
- 501 20. Hobday RA, Dancer SJ. Roles of sunlight and natural ventilation for controlling infection:
502 historical and current perspectives. *J Hosp Infect* 2013; **84**: 271-282
- 503 21. Lieverse R, van Someren JW, Nielen MA, Uitdehaag BMJ, Smit JH, Hoogendijk WJG. Bright
504 light treatment in elderly patients with non-seasonal major depressive disorder: a
505 randomized placebo-controlled trial. *Am Med Assoc* 2011; **68**: 61-70
- 506 22. Bernhofer EI, Higgins PA, Daily BJ, Burant CJ, Hornick TR. Hospital lighting and its
507 association with sleep, mood and pain in medical inpatients. *J Adv Nurs* 2014; **70**(5):
508 1164-1173
- 509 23. Kent A. News and views from the literature. *Rev Obstet Gynecol* 2013; **6**: 25-38
- 510 24. Simmons S, Morgan M, Hopkins T, Helsabeck K, Stachowiak J. Impact of a multi-hospital
511 intervention utilising screening, hand hygiene education and pulsed xenon ultra violet
512 (PX-UV) on the rate of hospital associated Methicillin *Staphylococcus aureus*. *J Infect Prev*
513 2013; **14**: 172-174
- 514 25. Boyce JM, Havill NL, Moore BA. Terminal decontamination of patient rooms using an
515 automated mobile UV light unit. *Infect Control Hosp Epidemiol* 2011; **32**: 737-742

- 516 26. Davies A, Pottage T, Bennett A, Walker J. Gaseous and air decontamination technologies
517 for *Clostridium difficile* in the healthcare environment. *J Hosp Infect* 2011; **77**: 199-203
- 518 27. Shin GA, Linden KG, Arrowood MJ, Sobsey MD. Low pressure UV inactivation and
519 DNA repair potential of *Cryptosporidium parvum* oocysts. *Appl Environ Microbiol* 2001;
520 **67**: 3029-3032
- 521 28. Kac G, Gueneret M, Rodi A, et al. Evaluation of new disinfection procedure for ultrasound
522 probes using ultraviolet light. *J Hosp Infect* 2007; **65**: 163-168
- 523 29. Hamblin MR, Viveiros J, Yang C, Ahmadi A, Ganz RA, Tolkoﬀ MJ. Helicobacter pylori
524 accumulates photoactive porphyrins and is killed by visible light. *Antimicrob Agents*
525 *Chemother* 2005; **49**: 2822-2827
- 526 30. Guffey JS, Wilborn J. *In vitro* bactericidal effects of 405-nm and 470-nm blue light.
527 *Photomed Laser Surg* 2006; **24**: 684-688
- 528 31. Enwemeka CS, Williams D, Hollosi S, Yens D, Enwemeka SK. Visible 405nm SLD photo
529 destroys methicillin-resistant *Staphylococcus aureus* (MRSA) *in vitro*. *Laser Surg Med*
530 2008; **40**:734-737
- 531 32. Maclean M, MacGregor SJ, Anderson JG, Woolsey GA. Inactivation of bacterial pathogens
532 following exposure to light from a 405nm LED array. *Appl Environ Microbiol* 2009; **75**:
533 1932-1937
- 534 33. Maclean M, Murdoch LE, MacGregor SJ, Anderson JG. Sporicidal effects of high-intensity
535 405 nm visible light on endospore-forming bacteria. *Photochem Photobiol* 2013; **89**:
536 120-126
- 537 34. Endarko, Maclean M, Timoshkin IV, MacGregor SJ, Anderson JG. High intensity 405nm
538 light inactivation of *Listeria monocytogenes*. *Photochem Photobiol* 2012; **88**: 1280-1286
- 539 35. Maclean M, MacGregor SJ, Anderson JG, Woolsey GA. High-intensity narrow-spectrum
540 light inactivation and wavelength sensitivity of *Staphylococcus aureus*. *FEMS Microbiol*
541 *Lett* 2008; **285**: 227-232
- 542 36. Goldoni A. Porphyrins: fascinating molecules with biological significance. *ELETTRA*
543 *Laboratory, Research Highlights 2001-2002: Atomic, Molecular and Supramolecular*
544 *Studies* 2002; 64-65.

- 545 37. Ashkenazi H, Malik Z, Harth Y, Nitzan Y. Eradication of *Propionibacterium acnes* by its
546 endogenic porphyrins after illumination with high intensity blue light. *FEMS Immunol*
547 *Med Microbiol* 2003; **35**: 17-24
- 548 38. Feuerstein, O, Ginsburg I, Dayan E, Veler D, Weiss EI. Mechanism of visible light
549 phototoxicity on *Porphyromonas gingivalis* and *Fusobacterium nucleatum*. *Photochem*
550 *Photobiol* 2005; **81**: 1186-1189
- 551 39. Maclean M, MacGregor SJ, Anderson JG, Woolsey GA. The role of oxygen in the visible-
552 light inactivation of *Staphylococcus aureus*. *J Photochem Photobiol B* 2008; **92**: 180-184
- 553 40. Lipovsky A, Nitzan Y, Friedmann H, Lubart R. Sensitivity of *Staphylococcus aureus*
554 strains to broadband visible light. *Photochem Photobiol* 2009; **85**: 255-260
- 555 41. Dai T, Gupta A, Murray CK, Vrahas MS, Tegos GP, Hamblin MR. Blue light for infectious
556 diseases: *Propionibacterium acnes*, *Helicobacter pylori*, and beyond? *Drug Resist Update*
557 2012; **15**: 223-236
- 558 42. Murdoch LE, Maclean M, Endarko, MacGregor SJ, Anderson JG. Bactericidal effects of
559 405-nm light exposure demonstrated by inactivation of *Escherichia*, *Salmonella*, *Shigella*,
560 *Listeria* and *Mycobacterium* species in liquid suspensions and on exposed surfaces.
561 *ScientificWorldJournal* 2012; Article ID 137805.
- 562 43. Wasson CJ, Zourelis JL, Aardsma NA, Eells JT, Ganger MT, Schober JM, Skwor TA.
563 Inhibitory effects of 405 nm irradiation on *Chlamydia trachomatis* growth and
564 characterization of the ensuing inflammatory response in HeLa cells. *BMC Microbiol*
565 2012; **12**:176-186
- 566 44. McKenzie K., Maclean M, Timoshkin IV, Endarko, MacGregor SJ, Anderson JG.
567 Photoinactivation of bacteria attached to glass and acrylic surfaces by 405nm light:
568 Potential application for biofilm decontamination. *Photochem Photobiol* 2013; **89**: 927-
569 935
- 570 45. Wilcox MH, Fawley WN. Hospital disinfectants and spore formation by *Clostridium*
571 *difficile*. *The Lancet* 2000; **356**: 1324
- 572 46. Rupnik M, Wilcox MH, Gerding DN. *Clostridium difficile* infection: new developments in
573 epidemiology and pathogenesis. *Nature* 2009; **7**: 526-536

- 574 47. St Denis TG, Dai T, Izikson L, et al. All you need is light antimicrobial photo inactivation
575 as an evolving and emerging discovery strategy against infectious disease. *Landes*
576 *Bioscience* 2011; **2**: 509-520
- 577 48. McKenzie K, Maclean M, Timoshkin IV, MacGregor SJ, Anderson JG. Bactericidal effect of
578 405nm light on *Escherichia coli* and *Listeria monocytogenes* in the presence of sub-lethal
579 stress. *Int J Food Microbiol* 2014; **170**: 91-98
- 580 49. Donlan MR. Biofilms: Microbial Life on Surfaces. *Emerg Infect Dis* 2002; **8**: 881-890
- 581 50. Hota S, Hirji Z, Stockton K, et al. Outbreak of multidrug resistant *Pseudomonas*
582 *aeruginosa* colonization and infection secondary to imperfect intensive care unit room
583 design. *Infect Control Hosp Epidemiol* 2009; **30**: 25-33
- 584 51. Chang JCH, Ossoff SF, Lobe DC, et al. UV inactivation of pathogenic and indicator
585 microorganisms. *Appl Environ Microbiol* 1985; **49**: 1361-1365
- 586 52. Hijnen WAM, Beerendonk EF, Medema GJ. Inactivation credit of UV radiation for viruses,
587 bacteria and protozoan (oo)cysts in water: a review. *Water Res* 2006; **40**:3-22
- 588 53. Murdoch LE, Maclean M, MacGregor SJ, Anderson JG. Inactivation of *Campylobacter jejuni*
589 by exposure to high-intensity 405nm visible light. *Foodborne Path Dis* 2010; **7**: 1211-
590 1216.
- 591 54. Feuerstein O, Persman N, Weiss EI. Phototoxic effect of visible light on *Porphyromonas*
592 *gingivalis* and *Fusobacterium nucleatum*: an *in vitro* study. *Photochem Photobiol* 2004;
593 **80**:412-415
- 594 55. Soukos NS, Som S, Abernethy AD, Ruggiero K, Dunham J, Lee C, Doukas AG, Goodson JM.
595 Phototargeting oral black-pigmented bacteria. *Antimicrob Agents Chemother* 2005;
596 **49**:1391-1396
- 597 56. Murdoch LE, McKenzie K, Maclean M, MacGregor SJ, Anderson JG. Lethal effects of high
598 intensity violet 405-nm light on *Saccharomyces cerevisiae*, *Candida albicans* and on
599 dormant and germinating spores of *Aspergillus niger*. *Fungal Biol* 2013; **117**: 519-527
- 600 57. Yin R, Dai T, Avci P et al. Light based anti-infectives: ultra violet C irradiation,
601 photodynamic therapy, blue light and beyond. *Curr Opin Pharmacol* 2013; **13**: 1-32

- 602 58. Maclean M, MacGregor SJ, Anderson JG, *et al.* Environmental decontamination of a
603 hospital isolation room using high-intensity narrow-spectrum light. *J Hosp Infect* 2010;
604 **76**: 247-251.
- 605 59. Bache SE, Maclean M, MacGregor SJ, *et al.* Clinical studies of the HINS-light
606 Environmental Decontamination System for continuous disinfection in the burn unit
607 inpatient and outpatient settings. *Burns* 2012; **38**: 69-76
- 608 60. Maclean M, Booth M, MacGregor SJ, Anderson JG, Woolsey GA, Coia JE, Hamilton K,
609 Gettinby G. Continuous decontamination of an intensive care isolation room during
610 patient occupancy using 405nm light technology. *J Infect Prevent* 2013; **14**: 176-181.
- 611 61. International Commission of Non-Ionizing Radiation Protection (ICNIRP). Guidelines on
612 limits of exposure to optical radiation from 0.38-3.9mm. *Health Physics* 1997; **73**: 539-
613 554
- 614 62. International Commission on Non-Ionizing Radiation Protection (ICNIRP). Guidelines on
615 limits of exposure to ultra violet radiation of wavelengths between 180nm and 400nm
616 (incoherent radiation). *Health Physics* 2004; **87**: 171-186
- 617 63. McDonald RS, Gupta S, Maclean M, *et al.* 405nm light exposure of osteoblasts and
618 inactivation of bacterial isolates from arthroplasty patients: potential for new
619 decontamination applications? *Eur Cell Mater* 2013; **25**: 204-214
- 620 64. Bache SE, Maclean M, Anderson JG, *et al.* Laboratory inactivation of healthcare-
621 associated isolates by a visible HINS-light source and its clinical application in the burns
622 unit (Abstract). *Burns* 2011; **37**: S6.
- 623 65. French GL, Otter JA, Shannon KP, Adams NMT, Watling D, Parks MJ. Tackling
624 contamination of the hospital environment by methicillin resistant *Staphylococcus*
625 *aureus* (MRSA): a comparison between conventional terminal cleaning and hydrogen
626 peroxide vapour decontamination. *J Hosp Infect* 2004; **57**: 31-37
- 627 66. Clark J, Barrett SP, Rogers M, Stapelton R. Efficacy of super oxidised water fogging in
628 environmental decontamination. *J Hosp Infect* 2006; **64**: 386-390
- 629 67. Department of Health. An integrated approach to hospital cleaning: microfiber cloth and
630 steam cleaning technology. DH London 2007

- 631 68. Hardy KJ, Gossain S, Henderson N. Rapid recontamination with MRSA of the
632 environment of an intensive care unit after decontamination with hydrogen peroxide
633 vapour. *J Hosp Infect* 2007; **66**: 360-368
- 634 69. Andrady, AL, Hamid SH, Hu X, Torikai A. Effects of increased solar ultraviolet radiation
635 on materials. *J Photochem Photobiol B* 1998; **46**:96-103.
- 636 70. Matsumura Y, Ananthaswamy HN. Toxic effects of ultra violet radiation on the skin.
637 *Toxicol Appl Pharmacol* 2004; **195**: 298-308
- 638 71. Sinah RP, Hader DP. UV-induced DNA damage and repair: a review. *Photochem Photobio*
639 *Sci* 2002; **1**: 225-236
- 640 72. Oguma K, Katayama H, Ohgaki S. Photoreactivation of Escherichia coli after low or
641 medium pressure UV disinfection determined by an endonuclease sensitive site assay.
642 *Appl Environ Microbiol* 2002; **68**: 6029-6035
- 643 73. Hamblin MR, Hasan T. Photodynamic therapy: a new antimicrobial approach to
644 infectious disease? *Photochem Photobiol Sci* 2004; **3**: 436-450
- 645 74. Nitzan Y, Kauffman M. Endogenous porphyrin production in bacteria by δ -
646 aminolaevulinic acid and subsequent bacterial photoeradication. *Laser Med Sci* 1999;
647 **14**: 269-277
- 648 75. Donnelly RF, McCarron PA, Tunney MM. Antifungal photodynamic therapy. *Microbiol Res*
649 2008; **163**: 1-12
- 650 76. Foster RG. The 'third' photoreceptor system of the eye – photosensitive retinal ganglion
651 cells. *Eur Ophthal Rev* 2009; **2**(1):84-86
- 652 77. Shiomori T, Miyamoto H, Makishima K, *et al.* Evaluation of bedmaking-related airborne
653 and surface methicillin-resistant *Staphylococcus aureus* contamination. *J Hosp Infect*
654 2002; **50**: 30-35
- 655 78. Bache SE, M Maclean, G Gettinby, JG Anderson, SJ MacGregor, I Taggart. Quantifying
656 bacterial transfer from patients to staff during burns dressing and bed changes:
657 implications for infection control. *Burns* 2013; **39**: 220–228
- 658 79. Bhalla A, Pultz NJ, Gries DM *et al.* Acquisition of nosocomial pathogens on hands after
659 contact with environmental surfaces near hospitalized patients. *Infect Control Epidemiol*
660 2004; **25**: 164-166

661

662

663 **Table I.** Comparison of the properties of ultraviolet C (UVC) and 405 nm light for environmental
664 disinfection applications.

	UVC LIGHT	405nm LIGHT
Typical/Potential use	Terminal clean of air and light exposed surfaces	Continuous disinfection of air and light exposed surfaces
Safety	Significant safety hazards associated with human exposure; can cause DNA mutations, erythema ⁷⁰	Can be utilised safely in the presence of people at recommended irradiation levels ⁵⁸⁻⁶⁰
Mechanism of Action	DNA damage kills cells. Sub-lethally damaged cells can recover using photoreactivation mechanism to repair DNA ^{71,72}	Photoexcitation of intracellular molecules induces oxidation of microbial cells. No known repair mechanism ^{73,74}
Antimicrobial Activity	Broad spectrum action against a range of microorganisms including spores and viruses ^{51,52}	Effective against bacteria, fungi, yeasts and spores; antiviral activity not yet fully established ^{32,33,56}
Antimicrobial Efficacy	Rapid inactivation rate within treatment zone ^{6,24}	Comparably slower inactivation rate within treatment zone ⁵⁸⁻⁶⁰
Materials Compatibility	UV light associated polymer damage ⁶⁹	Lower energy 405 nm wavelengths more materials compatible ⁶⁹
Ease of Use for Environmental Disinfection	Rooms/wards need to be vacated during use; operator training required ^{6,24}	Can be safely used during room occupation; no operator safety training required ⁵⁸⁻⁶⁰
Microbial Mutagenic Potential	Powerful mutagen that may encourage resistance development	Multi-target oxidative action mitigates against resistance development ⁷⁵
Penetrability	Does not penetrate through plastics and glass and weakly penetrates into water and fabrics	Can penetrate through plastics and glass and penetrates into water and fabrics ⁴⁴

665

666

667
668
669
670
671
672
673
674
675
676
677
678
679
680
681
682
683
684
685
686
687
688
689
690
691
692
693
694

Figure Legends

Figure 1. *UV, visible light and infrared regions of the electromagnetic spectrum. Highlighted are key UV and violet/blue wavelengths with detail of their germicidal action and safety aspects.*

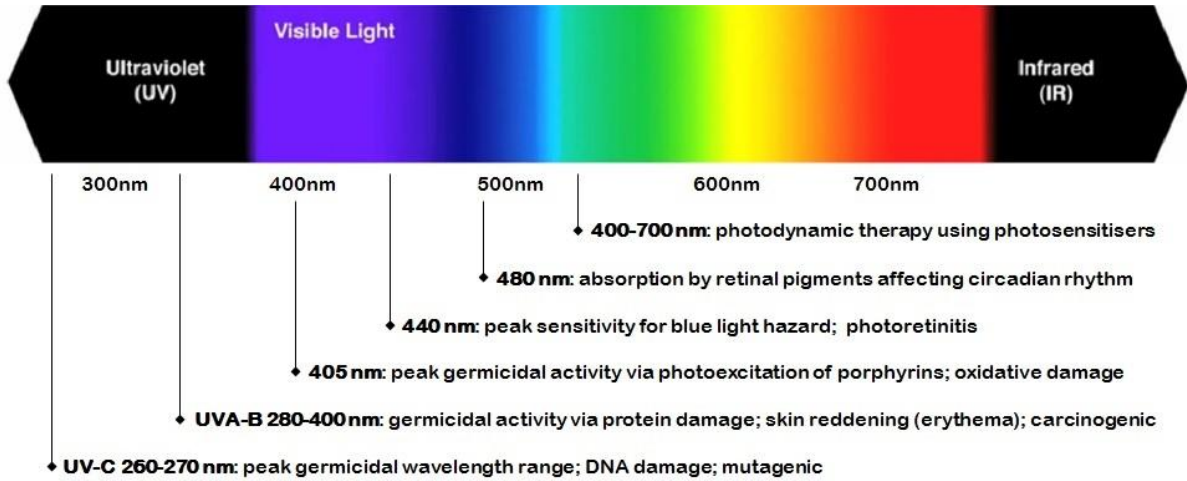
Figure 2. *Mean reductions in the total staphylococcal counts and the presumptive S. aureus levels across an occupied burns unit isolation room, before, during and after 5-day use of HINS-light EDS. Contact plate samples (n=70) were collected twice during each phase and the results pooled to assess the overall decontamination effect. A significant 62% decrease in total staphylococcal counts, and 50% decrease in presumptive S. aureus was achieved (P<0.05). 'After use' samples, showed that contamination around the room had significantly risen over the 6-days after the system was switched off: 126% and 98% increase in the total staphylococci and presumptive S. aureus counts, respectively (P<0.05). (Data adapted from ⁵³).*

Figure 3. *Reductions in the mean levels of environmental bacteria on a range of surfaces in an ICU isolation room before and after 5-day use of the HINS-light EDS. Tryptone soya agar contact plate samples were collected from each surface and results pooled to show the mean reduction in contamination on the sampled surface (data adapted from ⁵⁵).*

695

696

697



698

699

700

701

702

703

704

705

706

707

708

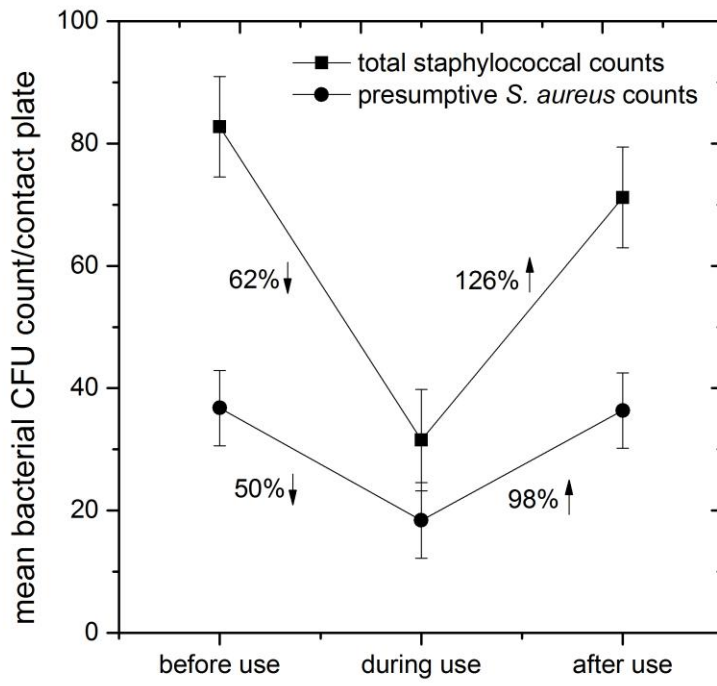
709

710

711

712

713



714

715

716

717

718

719

720

721

722

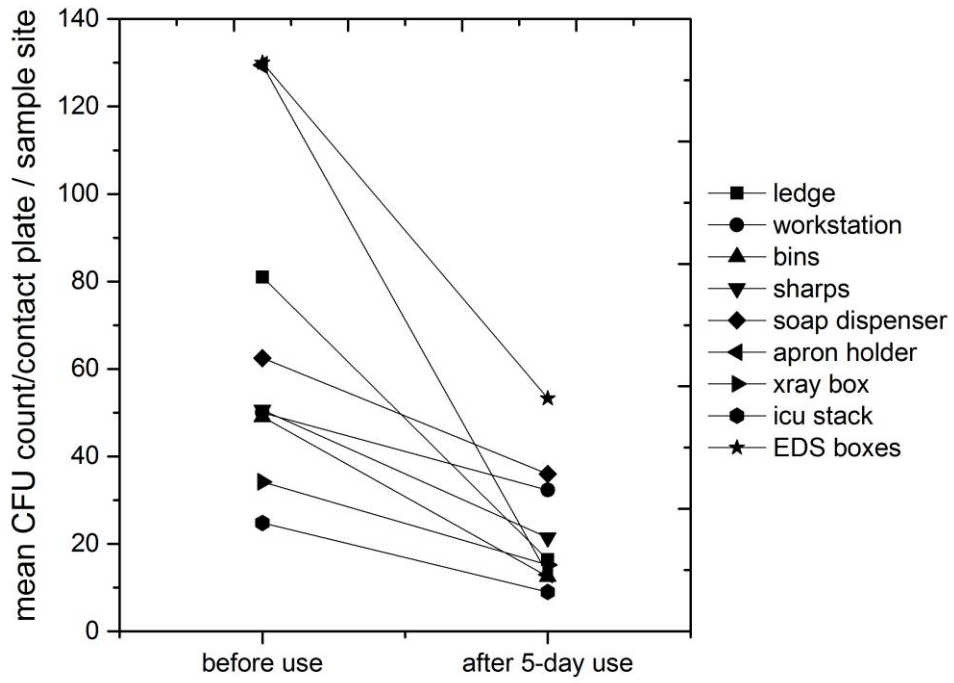
723

724

725

726

727



728

729

730